Interim Measures Investigation Report

EXIDE ENVIRONMENTAL RESPONSE TRUST Frankfort Indiana Site 555 North Hoke Avenue EPA ID No. IND001647460

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PROFESSIONAL GEOLOGIST CERTIFICATION

Interim Measures Investigation Report EXIDE ENVIRONMENTAL RESPONSE TRUST Frankfort Indiana Site 555 North Hoke Avenue EPA ID No. IND001647460

By affixing my seal, I am certifying that information which was developed by Montrose for use in this report is true and correct, and that information obtained from other parties was utilized without intentional modification. I further certify that I am licensed to practice in the State of Indiana and that it is within my professional expertise to verify the correctness of the appropriate hydrogeologic information.

STATE OF MOINTINGS STATE OF MOINTINGS No. 1562

Gregory Smoot, P.G. (License No. 1562)

Signed and sealed this day 29 July 2022



ABBREVIATIONS

AOC area of concern

BTEX Benzene, Toluene, Ethyl Benzene, and xylenes

CA Corrective Action
cis-1,2-DCE cis-1,2-Dichloroethene
CSM Conceptual Site Model

CVOCs chlorinated volatile organic compounds

GC gas chromatograph HQ Hazard Quotient

IDEM Indiana Department of Environmental Management

ICM Interim Corrective Measures
IMI Interim Measures Investigation
IMWP Interim Measures Work Plan

L liter

MCL Maximum Contaminant Limit

m meter

PCE Perchloroethylene

PRT Geoprobe's Post Run Tubing RFI RCRA Facility Investigation

RISC Risk Integrated System of Closure
SOP Standard Operating Procedure
SLVE Screening Level Vapor Exposure
SSI Supplemental Site Investigation

TCE Trichloroethene ug microgram

USEPA United State Environmental Protection Agency

VISL Vapor Intrusion Screening Level VOCs volatile organic compounds

VI Vapor intrusion VC Vinyl Chloride



EXECUTIVE SUMMARY

Advanced GeoServices Corp., dba Montrose Environmental Solutions LLC (Montrose), on behalf of the Exide Environmental Response Trust (Trust) performed an Interim Measures Investigation (IMI) to address impacts to groundwater from volatile organic compounds (VOCs) and chlorinated VOCs (CVOCs) at the former Exide Technologies (Exide) facility located at 555 North Hoke Avenue in Frankfort, Indiana (EPA ID# IND 001 647 460) (i.e., the Site). The work was performed in accordance with the Interim Measures Work Plan (IMWP or Work Plan) issued by Montrose on August 30, 2021.

Montrose prepared a Comprehensive RCRA Facility Investigation (RFI) Report dated February 24, 2021, as part of the RCRA Corrective Action (CA) process. The RFI Report recommended conducting focused investigation activities. The focused IMI activities included installing additional monitoring wells and groundwater sampling and analysis for VOCs and CVOCs to define the movement of groundwater and groundwater contamination in the area north and east of MW-4, for the ultimate purpose of proceeding with interim measures for chlorinated VOCs.

The IMI activities included the collection of representative soil gas samples using permanent soil gas sampling points and probes and direct push methods from select locations (including three samples directly above or adjacent to the sanitary sewer bedding). Vapor samples were also collected from sanitary and storm water manholes for analysis for VOCs and CVOCs. Vapor sampling was conducted to assess the subsurface vadose zone, local utilities, and local sewer bedding for the presence of VOC and CVOC vapors that could be moving along preferred pathways.

Groundwater in on-Site monitoring wells is impacted with VOCs and CVOCs. Recently, the TCE concentrations detected were 49.1 ug/L at MW-1, and 187,000 ug/L at MW-4. Off-Site monitoring well MW-9 contained 1.8 ug/L TCE, 3,210 ug/L cis-1,2-DCE, and 957 ug/L vinyl chloride. The TCE impact at MW-1 appears to be discontinuous with the CVOC plume at MW-4. MW-1 is over 500 feet from MW-4 and it is unlikely that the TCE in MW-1 is a result of contaminant transport by groundwater flow down hydraulic gradient. The CVOC concentrations decrease quickly with distance away from MW-4 which indicates that the plume of TCE impacted groundwater is localized and likely contained by low-permeability soils. The source of CVOCs is suspected to be associated with AOC-3/UST-2 because CVOC impacts were within 20 feet of UST-2. The In Situ Microcosm study suggests there is a strong potential for the complete anaerobic reductive dechlorination of PCE and TCE under bioaugmentation with SDC-9 and SRS amendment at this Site. The complicating issue may be that the subsurface may not be able to adequately transmit amendments through the area of impact given the extensive low-permeability soil in the subsurface.

Groundwater was evaluated to assess the CVOC plume, groundwater flow direction, and groundwater velocity to evaluate the potential for groundwater in exceedance of the standards to migrate off-site. The groundwater flow rate is extremely slow due to laterally extensive low-permeability glacial tills combined with a low groundwater gradient. Data indicates that it can take decades for groundwater to flow 1.0 foot horizontally at the Site.



The results of soil gas samples indicate that there are no exceedances of the Target Sub-Slab and Near Source Soil Gas Concentration VISL.

The sewer gas samples collected from manhole locations exceeded the Target Sub-Slab and Near Source Soil Gas Concentration VISL for CVOCs. No unique utility bedding fill material (i.e., sand/gravel) was identified during the investigation at three locations and it appears the utility trench was backfilled and compacted using the local excavated material. The soil gas samples taken adjacent to the sewer pipe did not contain soil gas results above VISL. The issue of CVOC vapors in the sewer system is complex because the sewer is the receptor of pollutants from a large number of potential sources. CVOC vapors in sewer lines should not be a source for indoor CVOC vapor intrusion because modern plumbing systems has sewer traps to prevent gases contained in wastewater, the public sewer, or septic tanks from escaping and entering residential housing.

Recommendations

Elevated lead in the shallow surface soils at the surface water discharge locations should be delineated and removed, or stabilized using capping, fencing, silt fence or similar techniques as Corrective Action alternatives. Additionally, appropriate institutional and deed controls can be put into place.

Based on the results of this study, the CVOC Remediation Evaluation dated January 19, 2021, will be revisited to further evaluate the potential groundwater remedial actions that may be employed at the Site.

Additional sampling and/or investigation to define the extent of trichloroethene (TCE) contamination in the area of groundwater near monitoring wells MW-1 and MW-13 will be considered under a separate future work plan.

Future sampling and/or investigations will be necessary to evaluate and identify the permanent solutions to assure protection of groundwater at, and beyond, the property boundary and also identify how potential exposures to off-site receptors will be mitigated, if warranted.

A prudent method to further investigate for evidence of COVC impacted groundwater from potentially infiltrating the sewer would be to run a camera through the sewer, possibly after waterjetting the line, to assess for potential pipe connections from the Site or obvious breaches in the sewer line. This can be conducted during Interim Remedial Action.



1.0 OVERVIEW

Advanced GeoServices Corp., dba Montrose Environmental Solutions LLC (Montrose), on behalf of the Exide Environmental Response Trust (Trust) performed an Interim Measures Investigation (IMI) to address impacts to groundwater from volatile organic compounds (VOCs) and chlorinated VOCs (CVOCs) at the former Exide Technologies (Exide) facility located at 555 North Hoke Avenue in Frankfort, Indiana (EPA ID# IND 001 647 460) (i.e., the Site). The work was performed in accordance with the Interim Measures Work Plan (IMWP or Work Plan) issued by Montrose on August 30, 2021.

Montrose conducted a Comprehensive RCRA Facility Investigation (RFI) at the Site and presented the results of the RFI and Interim Corrective Measures (ICM) activities in a report titled Comprehensive RFI Report dated February 24, 2021 (RFI Report). The RFI was completed as part of the RCRA Corrective Action (CA) process. Investigation of groundwater in May and July 2018 showed impact to groundwater in on-site monitoring wells from VOCs and CVOCs. The RFI Report recommended conducting focused investigation activities to define the movement of groundwater and groundwater contamination in the area north and east of MW-4, for the ultimate purpose of proceeding with interim measures for chlorinated VOCs. The focused investigation activities included additional groundwater sampling and analysis for VOCs and CVOCs as detailed herein.

The investigation also included the collection of representative soil gas samples using permanent soil gas sampling points and probes and direct push methods from select locations (including three samples directly above or adjacent to the sanitary sewer bedding). Vapor samples were also collected from sanitary and storm water manholes for analysis for VOCs and CVOCs.

1.1 Facility Location and Description

The Site is located in central Indiana within Clinton County, approximately 50 miles northwest of Indianapolis (see Figure 1). The Site is bounded by North Hoke Avenue to the west, Kelley Avenue to the east, Michigantown Road to the north (also referred to as Washington Street on some maps), and Norfolk Southern railroad tracks to the south. The Site consists of eighteen (18) contiguous parcels now owned by the Trust which encompass approximately 13.7 acres. All but three of the parcels are located within a perimeter security fence. The majority of the area (12.1 acres) lies within the perimeter security, and with the exception of grass and a few shrubs along North Hoke Avenue is covered with former building pads, pavement or crushed stone. The facility was formerly a manufacturing plant that produced lead-acid automotive batteries. The plant was closed and the aboveground infrastructure was decontaminated and demolished January 2013.

1.2 Geologic Setting

As the Laurentide ice sheet began to retreat from present day Northern Indiana and Northwest Ohio between 14,000 and 15,000 years ago, it receded into three distinct lobes. The eastern or Erie Lobe sat atop and behind the Fort Wayne Moraine. Meltwater from the glacier fed into two



ice-marginal streams, which became the St. Joseph and St. Marys Rivers. Their combined discharge was probably the primary source of water for the proglacial Wabash River system. Rich prairie soils extend over central Indiana. Some clays in east-central Indiana are compact with poor drainage and frequent ponding of water. Till (material deposited directly by glaciers) forms flat to hummocky plains that dominate the central portion of the state. The thickness of the glacial deposits ranges from 100 to 400 feet throughout Clinton County.

Bedrock beneath the Site is located at approximately 550 feet MSL; or approximately 300 feet below ground surface (bgs). The Site appears to fall near the contact between Devonian and Silurian bedrock units. Devonian formations typically have a carbonaceous shale on the upper portion and are underlain by limestone, dolostone and shale. Silurian bedrock contains the carbonates limestone and dolostone. Indiana bedrock geology features a broad anticline with a slight plunge to the northwest.

1.3 <u>Local Geology</u>

The soil and unconsolidated materials in the subsurface of the Site consist of glacial till which is unsorted glacial sediment that washes off of retreating glaciers. The soil has been mapped as the Fincastle-Crosby soils. This is a silty loam with slow infiltration rates (Class C) and is somewhat poorly drained. The area is characterized by swell and swale topography. Fincastle soils are typically observed on rises and have a brown silt loam surface layer, and yellowish brown, mottled silty clay loam to clay loam subsoil. Crosby soils are found on high rises and have a brown silt loam surface layer, and yellowish brown, mottled silty clay loam, clay loam, and loam subsoil. The hydraulic conductivity (K) in these regional silt and clay deposits is low which makes these deposits semi-pervious. Slug testing of select monitoring wells has indicated that vertical hydraulic conductivities range from 0.0004 to 0.0088 feet/day (See Section 2.6) and are consistent with glacial till.

1.4 <u>Hydrogeologic Setting</u>

The Tipton Complex Aquifer System is characterized by unconsolidated deposits that are quite variable in materials and thickness. Aquifers within the system range from thin to thick and include single or multiple intra-till sands and gravels. The aquifers are highly variable in depth and lateral extent and are typically confined by thick clay layers. The total unconsolidated thickness of the Tipton Complex Aquifer System generally ranges from about 200 feet to over 400 feet in Clinton County. The potentiometric surface of the regional unconsolidated aquifer is approximately 800 feet MSL; or approximately 50 feet bgs.

Aquifer layers utilized in the Tipton Complex Aquifer System are generally 5 to 10 feet thick sands and/or gravels. These sands and gravels are overlain by a till cap which is commonly 65 to 190 feet thick with thin intratill sand and gravel layers. Wells in this system are typically completed at depths ranging from 68 to 195 feet. Domestic well yields are commonly 15 to 65 gallons per minute (gpm) and static water levels are generally 15 to 35 feet below the surface. There are 8 registered significant ground-water withdrawal facilities (29 wells) in this system in Clinton County. High-capacity well yields of up to 1,200 gpm are reported.



1.5 <u>Local Hydrogeology</u>

Shallow groundwater is present in the till cap that is 65 to 195 feet thick and overlays the layers of sand and gravel. Shallow groundwater is present as local perched zones of saturation in clay and small local sand and silt layers. The depth and thickness of the saturated layers varied from 4 to 10 feet below ground surface, and appear to be laterally discontinuous. The depth to the water table measured in the fourteen (14) monitoring wells at the site range from 1.89 to 8.83 feet below top of casing. The underlining aquiclude was encountered in all of the groundwater monitoring well locations and consisted of a very stiff to hard gray clayey Silt to silty Clay with trace amounts of sand and/or gravel. The perched groundwater near MW-4 was 4.43 feet below top of casing in December 2021 and groundwater flows very slowly generally towards the north (see Section 3.10.4).

Based on the well gauging data collected on December 8, 2021, the hydraulic gradient in the central portion of the Site is 0.007 toward the north northwest. In the northern portion of the site near MW-1 and MW-2, the hydraulic gradient steepens slightly to 0.013 and turns to the north northeast. On the eastern portion of the Site in the area of groundwater contamination near MW-4, groundwater flows north under a hydraulic gradient of 0.013. In the area along North Kelley Avenue, the groundwater flow turns to the northeast under a gradient of 0.010. The groundwater flow rate calculated in Section 3.11.4 is extremely slow due to laterally extensive low-permeability glacial tills combined with a low groundwater gradient. Data indicates that it can take decades for groundwater to flow 1.0 foot horizontally at the Site.

The low groundwater velocity indicates that the groundwater at the Site is "old" groundwater, meaning there has been long period since the water recharged the subsurface. In this setting, it is not uncommon for overturned groundwater ages, which is where younger groundwater is under older groundwater. The significance is that high age groundwater accumulates and contains contaminants.

1.6 <u>Community Relations Activities</u>

Community involvement and outreach consisted of the preparation of a fact sheet mailer sent to residents near the Site, updated language in the website (https://www.exidefrankfortclosure.com/), and installing signage on the fence of the Site.

1.7 Reference Documents

The focused investigation activities described herein were performed under the framework of the following site documents:

1. Supplemental Site Investigation Work Plan prepared for Exide Technologies by Advanced GeoServices Corp. dated December 21, 2018



- 2. Quality Assurance Project Plan for the RCRA Facility Investigation, Exide Technologies Former Manufacturing Facility, Frankfort, Indiana, prepared for Excide Technologies by Advanced GeoServices Corp. dated October 12, 2017
- 3. Sampling and Analysis Plan for the RCRA Facility Investigation, Former Exide Manufacturing Facility, Frankfort, Indiana, prepared for Excide Technologies by Advanced GeoServices Corp. dated October 12, 2017
- 4. *CVOC Remediation Evaluation* prepared by Advanced GeoServices / Montrose Environmental Group, dated January 19, 2021
- 5. *Interim Measure Work Plan, Frankfort Indiana Site,* prepared for Exide Environmental Response Trust by Montrose dated August 30, 2021

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2.0 INVESTIGATION ACTIVITIES

2.1 Objectives

The objectives of the investigation outlined in the IMWP dated August 30, 2021, were to:

- 1. Conduct a focused investigation activities to define the movement of groundwater and groundwater contamination in the area north and east of MW-4, for the ultimate purpose of proceeding with interim measures for chlorinated VOCs.
- Assess the subsurface vadose zone, local utilities, and local sewer bedding for the presence of VOC and CVOC vapors that could be moving along preferred pathways.

2.2 <u>Investigation Overview</u>

Montrose mobilized to the Site on October 12, 2021 to perform the majority of the field activities discussed in the Work Plan. Montrose mobilized to the site a second time on December 8, 2021 to perform the groundwater sampling and initiate the in-situ microcosm study. Field notes are provided in Appendix A. The following paragraphs discusses the field activities performed.

Montrose contracted Cascade Technical Services (Cascade) to provide all powered sub-surface installation and sampling (i.e., Hydropunch activities, well install, vapor port install, etc.). Samples collected were analyzed by Pace Analytical Laboratories at either an on-Site mobile lab or at their permanent location in Indianapolis, Indiana.

In order to achieve the objectives stated in Section 2.1, the IMWP developed a set of sampling locations for soil gas, sewer vapor, and groundwater samples to be collected and analyzed using an on-Site Pace Analytical® Services mobile laboratory gas chromatograph (GC) for the following parameters:

- Benzene, Toluene, Ethyl Benzene, and xylenes (BTEX);
- cis-1,2-Dichloroethene (cis-1,2-DCE);
- Perchloroethylene (PCE);
- Trichloroethene (TCE); and
- Vinyl Chloride (VC)

Eighteen (18) sample locations were designated for Hydropunch sampling and/or soil gas sampling in the IMWP designated Location 1 through Location 18. These locations are shown on Figure 2.

The locations designated as L1 through L6 were six vapor ports for the purpose of sampling soil gas along the east side of Kelley Avenue as described in Section 2.13. Montrose installed soil gas probes as fixed/permanent flush-mount wells, with bolted lids.



The locations designated as L7 through L11 were Hydropunch groundwater sampling locations that were analyzed on-Site for TCE, PCE, cis-1,2-DCE, VC, and BTEX.

Four "step-in" locations, designated as L7A, L8A, L9A, and L10A were additional Hydropunch groundwater sampling locations based on the results in accordance with the IMWP and described in Section 2.3.

The groundwater monitoring well network was enhanced by installing a new monitoring well at Location L12 (MW-11) on the south side of E. McClurg Street (approximately 50 feet east of MW-9) as shown on Figure 2. Additionally, three new groundwater monitoring wells (MW-12, MW-13, and MW-14) were installed at the fringe of the TCE plume around MW-4. The wells were sampled and analyzed at the on-Site laboratory. Surface water and sediment samples were collected at Outfall Z shown on Figure 2 and analyzed at an on-Site laboratory.

In addition, Montrose collected step out soil gas samples in the locations shown on the attached map designated as:

- Location 13 Along the unpaved road north of East McClurg Street (soil gas);
- Location 14 Near the curve close to 1609 Goder Drive (soil gas);
- Location 15 East of North Kelley Ave. and north of MW-3 (soil gas and Hydropunch).

Montrose obtained utility maps from the City of Frankfort and reviewed the available information (Appendix B). Montrose collected three (3) soil gas samples from soil gas probes and temporary Geoprobe vapor sampling at Locations 16, 17, and 18 which to collect soil gas samples from immediately above the saturated bedding for the sanitary sewer and storm sewer.

Eight (8) grab vapor samples were collected from sanitary manholes (B, C, E, F, G, H, and I). Storm sewer manhole A was an open grate manhole identified along Kelley Avenue as shown on the attached Figure 2. Montrose also collected VOC/CVOC vapor grab samples from sanitary manholes J and L along Washington Avenue shown on Figure 3.

2.3 Decision Tree Document

Pace® mobile laboratory testing services (Pace® Mobile Labs) was utilized so that data could be obtained quickly and decisions could be made based on the Decision Tree document contained in Appendix A of the IMWP. The Decision Tree document is summarized below. Table 1 summarizes the Decision Tree actions.

Groundwater samples were obtained using a Hydropunch sampler. If the groundwater sample results were between 100 and 600 ug/L TCE, sampling was deemed complete and no offset samples were collected. If the number was less than 100 ug/l, Montrose stepped in and installed an offset sample location closer to the source (MW-4). These locations are designated with an "A" on Figure 2. Four step-in Hydropunch locations (L7A, L8A, L9A, and L10A) were conducted. At a value of less than 100 ug/L, the extent of impact can be reasonably estimated employing



multiple lines of evidence, including, but not limited to, analytical data; extrapolation or modeling based on existing data; application of a conceptual site model; or other means for determining the extent of the contamination for the purposes of interim remedial action. If the number was over 600 ug/L, Montrose would have stepped out and completed a step-out offset Hydropunch sample. These locations are designated with an "B" on Figure 2. No step-out Hydropunch locations were warranted based on the on-Site laboratory data.

2.4 <u>Groundwater Sampling – Temporary Sampling Points</u>

Samples of the groundwater were collected in the general area of monitoring well MW-4. These samples were collected to better understand the horizontal extent of the observed groundwater VOC plume located at MW-4. Collection of groundwater samples was initially attempted using standard Geoprobe Hydropunch sampling techniques. However, water was not observed to recharge sufficiently to allow sampling using this methodology on site. Therefore, samples were collected by advancing a Geoprobe rod to a target depth of 11 feet (i.e., within the upper groundwater interval) and installing a temporary 1-inch diameter piezometer that was allowed to recharge overnight prior to sampling. Samples for VOCs were collected from these piezometers using a peristaltic pump.

The results provided by the mobile lab were reviewed for the concentration of TCE. Locations L8, L9, L10, and L11 had TCE concentrations less than 100 ug/L. Therefore, a second Hydropunch sample was collected at an offset boring located closer to the source (i.e., Locations L8A, L9A, L10A, and L11A) in order to better define the TCE extent around MW-4. Location L7 had a TCE result of 280 ug/L, which is within the target concentration of 100-600 ug/L, therefore, no additional step out sampling was performed (i.e., Location L7A).

2.5 Outfall Sampling

One stormwater outfall location is present along Kelley Ave. north of the Site. This outfall (designated Outfall Location Z on Figures 2 and 9) was observed to be flowing on October 19, 2021. A sample of the water was collected for VOC and Lead analysis. Additionally, a soil sample from the unnamed tributary the outfall pipe drains to was collected for VOCs and Lead analysis. Samples from the outfall (water and sediment) were relinquished to Pace – Indianapolis for analysis.

Discharge from the MH-10 pipe was not able to be sampled during the IMI field work.

2.6 Slug Testing

Slug testing was performed at monitoring wells MW-1, MW-3, MW-4, MW-7, MW-9, and MW-10 on October 13, 2021. The slug tests were performed by installing automated data logging devices in the selected wells (i.e., Eijkelkamp TD-Diver system [Divers]) and then adding sufficient deionized water to the well to fill the casing. The Divers were set to record the water level in the well at 5 second intervals. Once the water level in the well had regained 80% of its initial static level, the Divers were removed from the well and the data downloaded using the Eijkelkamp software.



A K value for each well was calculated using the USGS published Spreadsheets for the Analysis of Aquifer-Test and Slug-Test Data by Halford and Kuniansky (2002). This spreadsheet uses the Bouwer and Rice (1976) modeling to calculate the K value. During the analysis, it was found that the use of a 5 second data collection interval resulted in a significant amount of data (over 17,000 data points per well), while the spreadsheet used is only capable of analyzing 600 data points. Therefore, the data was systematically reduced by Montrose to focus on the first three hours of the testing when the well had regained 80% of its initial static level during that time. One well (MW-7) took a longer period than the other wells to regain this level, therefore additional points collected at an hourly or bihourly interval for the remaining time period (i.e., beyond the initial three hours) were incorporated into the data set for the calculation of the hydraulic conductivity (K) value for this well. The final summary sheet produced by the spreadsheet and a data plot for each well is presented in Appendix C. Calculated hydraulic conductivities range from 0.0004 to 0.0088 and are summarized on Table 3. The values are consistent with glacial till.

2.7 Monitoring Well Installation

Monitoring well installation activities were performed by Cascade using hollow-stem auger drilling techniques. The drill tools were decontaminated using a steam cleaner in between well locations to minimize the potential of cross contamination. A Montrose field geologist monitored the well installation activities. The wells were installed between October 19 and 21, 2021. Well installation logs are provided as Appendix D. A well construction summary table for all Site groundwater wells is included as Table 4.

One (1) groundwater monitoring well (MW-11) was installed off of the Site in the downgradient direction from well MW-9. This location was selected to better understand an eastward flow pattern noticed in sampling events following the installation of MW-9 as well as define the extent of the groundwater impacts observed in MW-9 sample results.

Three (3) groundwater monitoring wells were installed on-Site in the general area of existing well MW-4. One well (MW-12) was installed in the upgradient direction of groundwater flow. The other two wells (MW-13 and MW-14) were positioned in the down- and side-gradient directions to define the extent of the groundwater impacts at MW-4.

During monitoring well installation, a soil sample was collected from the auger tailings from the 2 to 3-foot depth interval for analysis of VOCs and lead. A sample could not be collected from the installation of MW-14 due to no tailings being generated during the installation process. The samples were relinquished to Pace Indianapolis for analysis.

These monitoring wells were constructed using a 2-inch ID, flush-threaded, Schedule 40 PVC riser with a factory-slotted 0.010-inch PVC well screen. The wells were installed using 10 feet of well screen. A sand pack was placed to a minimum of 2 feet above the top of the monitoring well screen with No. 1 sand. The annulus of the borehole above the sand was sealed to the ground surface using bentonite. Wells on site were completed as stick-up style wells with a 4-inch square protective casing extending approximately 3 feet below ground and projecting approximately 2



feet above ground. Monitoring well MW-11 was installed as a flush mount style well to allow for vehicular traffic in the road. A 2 by 2-foot square well pad was installed so that the surface slopes away from the well for all wells.

The wells were developed using a submersible pump and purged until the water removed from the well ran clear. Due to the yield of the wells and the flow rate of the pump, the wells ran dry multiple times during the development. The wells were allowed to recharge and development resumed until the desired visual clarity was achieved.

Soils and purged groundwater generated during well installation and development activities were containerized in 55-gallon drums and stored on site. A composite sample was taken for each matrix (i.e., one sample for soils and one sample for water) for off-site waste disposal characterization. The laboratory results are summarized on Table 5 and the laboratory reports are contained in Appendix H. The waste is classified as non-hazardous. The USEAP Generator ID Number is IND001647460. The drums of material were removed from the site by US Ecology on March 9, 2022 for disposal at EQ Detroit, Inc., ID Number MID980991566.

2.8 Synoptic Water Level Measurement

Depth-to-water was measured by Montrose in each well on the Site using an electronic water level indicator prior to the initiation of groundwater sampling activities. The synoptic measurements included the measurement of water levels in the monitoring wells. The wells were allowed to equilibrate to atmospheric pressure and the data was collected over a two-hour period to determine the potentiometric surface across the Site. The field personnel measured the water levels in the wells to the nearest 0.01 foot using the surveyed point at the top of the inner well casing for reference. An updated groundwater potentiometric map is presented on Figure 4 using the results of the synoptic water level measurement performed on December 8, 2021. Depth to bottom measurements were collected following completion of the groundwater sampling activities to prevent unnecessary disturbance of the settled material in the well that may affect the sampling results.

2.9 Groundwater Sampling

During December 2021, one round of groundwater sampling was performed from site monitoring wells MW-1, MW-2, MW-3, MW-4, MW-7, MW-9, MW-10, MW-11, MW-12, MW-13 and MW-14, using low-flow sampling techniques. The Site well construction and water level readings conducted during the installation and groundwater sampling events are presented in Table 4. Monitoring wells were purged and sampled from the suspected least contaminated well to the most contaminated well to minimize the potential for cross-contamination.

The wells were purged using a stainless-steel low-flow bladder pump placed at the midpoint of the screen in each well. A flow-through cell was used to measure pH, temperature, conductivity, redox potential, and dissolved oxygen prior to contact with oxygen at 3 to 5-minute intervals during purging. Turbidity was also measured at the same interval. The wells were purged until the field



parameters stabilize to within 10% over three readings and pH readings differed less than 0.1 unit.

Once the field parameters had stabilized, the flow rate was reduced to 100 ml/min. to collect volatile organic compounds (VOC) samples. Samples were collected directly from the pump discharge line into laboratory-supplied bottles containing the necessary preservatives. An equipment blank sample was collected from the sampling equipment each day of the sampling event.

Purge water was contained in 55-gallon drums for off-site disposal as discussed in Section 2.7.

2.10 Survey

Randall Miller & Associates, Inc., an Indiana licensed surveyor, located all new monitoring wells, permanent vapor ports, temporary piezometer, and manhole locations for the purpose of accurately representing the horizontal datum of the sampling locations in the state-plane coordinate system. Elevations and locations presented on Table 4 and figures for this report are based on these survey results.

2.11 Sediment and Surface Water Sampling

A sediment sample and a surface water sample were be collected at the discharge point of the Site storm water sewer to unnamed tributary along Michigantown Road, near Kelley Avenue. This is designated as Outfall Z on Figure 2. The sample was collected and sent to Pace Analytical for analysis for VOCs using EPA Method 8260 and lead analysis.

2.12 Utility Investigation

Montrose contacted the Frankfort Street Department to acquire available utility maps for sewer lines and utilities in Kelley Avenue, Hoke Avenue, Washington Avenue (aka Michigantown Road), and E. Morrison Street. Additionally, the manholes selected for vapor sampling (discussed in Sections 2.15) were surveyed by a professional surveyor following sample collection. The utilities have been incorporated into the figures of this report as appropriate. Drawings and summarized observations are contained in Appendix B.

Montrose subcontracted Bloodhound Underground Utility Locators (Bloodhound) to scan the work areas with ground penetrating radar (GPR) to determine the locations of any utilities or other underground features that may interfere with the proposed subsurface investigation activities. The field scan was performed on October 12, 2021. Minor adjustments were made following the GPR scan to the proposed subsurface investigation locations, however no individual point required significant movement from the proposed location (i.e., less than 5 feet). Sampling from and next to underground utilities was conducted as described in Section 2.14 and Section 2.15.



2.13 Kelly Avenue and Step-Out Soil Gas Sampling

On October 13 to 14, 2021, Montrose oversaw as Cascade installed six (6) soil vapor ports along North Kelley Ave (L1 through L6), Goder Drive (L13), East McClurg Street (L14) and west of North Kelly Ave. (L15). Montrose also had Cascade install three (3) sampling ports within the sewer utility bedding along the east side of Kelley Avenue (locations L16, L17, and L18).

The locations were hand-augured until groundwater was encountered. The 8-inch long, stainless-steel screen of the port was positioned approximately 1 foot above the encountered groundwater in the borehole. Teflon tubing was connected to the screen and extended to the surface to allow sampling. The screen area was filled with sand, bentonite was placed and hydrated to the surface grade, and a bolted flush mount well vault was installed at the surface. Observations during installation are summarized on Table 2.

Sampling of these permanent vapor port locations was performed on October 18, 2021. A shroud was placed over the sampling port and filled with helium. A pre-sample was collected in a Tedlar bag and field screened for the presence of significant (>10%) helium. No pre-sample contained significant helium. Collection of the pre-sample also allowed for the purging of more than 3 volumes of vapor from the sample port prior to collection of the vapor sample. The sample was collected into a new Tedlar bag using a hand operated pump and relinquished to the on-Site Pace mobile lab for VOC analysis.

Locations L1, L3, L14, L15, and L18 were the only locations that could be sampled for soil gas using this procedure. The other seven vapor point locations L2, L4, L5, L6, L13, L16, and 17 were discovered to be saturated and produce only groundwater. Upon determining that the procedures described in the August 30, 2021 Interim Measures Work Plan were inadequate and impractical at these locations, another procedure had to be used to obtain a soil gas samples. Geoprobe's Post Run Tubing (PRT) system of sample collection was selected to collect soil gas samples at a depth of 1.5 feet at The PRT system is a USEPA-approved method of collecting soil gas and is described in the Region 4 document "Operating Procedure: Soil Gas Sampling" dated February 24, 2020 (LSASDPROC-307-R4).

Geoprobe's Post Run Tubing (PRT) system of sample collection was selected to collect soil gas samples at a depth of 1.5 feet at locations L2, L4, L5, L6, L13, L16, and 17. This system collects soil vapors from a temporary sampling port. The PRT system is a predominant and industry accepted method of collecting soil gas samples. Using this system, soil gas samples can be collected with a high degree of assurance that the samples are representative shallow (1.5 foot) depth. The change in sampling procedures from a vapor point to the PRT system is not expected to have an impact on the quality and usability of the soil gas data for the following reasons:

• The PRT sample ports were placed as closely as possible to the permanent sample ports allowing for equipment restrictions (i.e., overhead wires). The PRT samples were collected within 5 feet of the permanent port locations.



- The post run tubing and PRT adapter with O-rings eliminates concerns about system leaks at threaded joints and ensures that the entire sampling train is air-tight. No ambient air is introduced into the sample during collection.
- Tubing/equipment used to collect the sample was either Teflon® or stainless steel.
- The samples were analyzed using an on-Site mobile laboratory eliminating concerns associated with storing and shipping samples.

A total of twelve (12) soil gas samples were collected and analyzed for BTEX, cis-1,2-DCE, PCE, TCE, and VC using a field GC operated by Pace Analytical. One duplicate sample was also collected. The results indicate no exceedances of the Target Sub-Slab and Near Source Soil Gas Concentration Vapor Intrusion Screening Level (VISL).

Aqueous samples were collected at the saturated probe locations L2, L4, L5, L6, L13, L16, and 17 for analysis for BTEX, cis-1,2-DCE, PCE, TCE, and VC using a field GC operated by Pace Analytical. Additionally, locations L3, L4, and L6 were selected to have groundwater sampled using the Geoprobe Hydropunch procedure. The Hydropunch samples were collected in the rough center of Kelley Ave. perpendicular to the corresponding soil gas location. The groundwater samples were collected at these locations for analysis for BTEX, cis-1,2-DCE, PCE, TCE, and VC using a field GC operated by Pace Analytical.

During December 10, 2021, Montrose field technicians attempted to collect soil gas samples from the permanent soil gas sampling ports (Locations L1 through L6). Two of these six locations (L1 and L3) could be sampled for soil gas while the other four could not be sampled due to the presence of significant amounts of water in the sample port. As this was the same conditions previously observed during October 2021, Montrose did not re-sample the two available ports as the resulting small dataset would not be of significant value to the overall Site model.

2.14 Utility Bedding Vapor Sampling

Montrose installed three (3) sampling ports within the sanitary sewer utility bedding along the east side of Kelley Avenue (locations L16, L17, and L18). No unique utility bedding fill material (i.e., sand or gravel) was identified during the installation of these ports, and it is Montrose's observation that the utility trench was backfilled using the local excavated material. Hand auger techniques were used to install these ports and prevent damage to the utility line. The ports were constructed such that the stainless-steel screen was placed adjacent to the utility within the reworked backfill material. Teflon tubing was used to allow sampling. The screen area of the port was backfilled with sand, and the borehole was sealed using hydrated bentonite to the ground surface. A permanent bolted flush-mount well vault was installed at the surface. Two of these ports (L16 and L17) were found to produce water and could not be sampled. Only location L18 could be sampled. The PRT method of collecting a soil gas samples was implemented to collect soil gas at a depth of 1.5 feet at L16 and L17.

The results indicate no exceedances of the Target Sub-Slab and Near Source Soil Gas Concentration VISL.



2.15 Manhole Sampling

The accumulated vapors within sewer manhole access points were collected along Kelley Ave, Washington Ave, and within the Site property. A dedicated length of tubing was lowered into the culvert and positioned so that the tubing was approximately 2 feet above the bottom of the culvert. If the manhole was an open grate-style manhole (i.e., a storm sewer inlet point), the location was not sampled under the assumption that the culvert is open to the atmosphere and therefore would not accumulate vapors. Manholes with multiple small holes (i.e., 2-inch diameter) were sampled by lowering the tubing through the hole. Manholes without any integral openings were opened sufficiently (i.e., 'cracked') to allow sample tubing access. The vapors were collected using a hand operated pump into a new Tedlar bag. The manholes are designated as Manhole B, C, E, F, G, H, I, J, L, and MH-10. The sewer vapor samples were for analyzed for BTEX, cis-1,2-DCE, PCE, TCE, and VC using a field GC operated by Pace Analytical.

Manhole A shown on Figure 2 was not sampled because it was an open grate manhole. Manhole C was initially believed to be a storm water manhole but is now believed to be a sanitary sewer manhole located where the sanitary sewer branches from North Kelley Avenue eastward along East McClurg Street. Manhole C was sampled. Based on site reconnaissance, Manhole N shown on Figure 3 is likely to be the same manhole that has been previously designated as "O" or "MH-11" on Site sketches. The exact location of the piping for stormwater Manhole N is not well understood but records indicate the piping makes a corner toward the North and then empties into the "K" manhole.

2.16 Passive In-Situ Microcosm Study

Upon completion of the groundwater sampling event, an in-situ microcosm study was completed by Montrose. This study entails collecting microbial, chemical and geochemical data to determine if monitored natural attenuation (MNA), bio-enhancement or bio-augmentation are appropriate interim measures. The in-situ study comprises Bio-Trap® Sampler and CENSUS analysis provided and performed by Microbial Insights, Inc. Bio-Trap® samplers are in-well study units that contain a sampling matrix favorable to colonization by microorganisms, which can later be harvested, quantified and assessed in the laboratory for microbial characterization and contaminant degradation potential (i.e. CENSUS analysis). The CENSUS assessment tests for the presence and growth of Dehalococcoides, intrinsic and augmented, and the microbial reductase genes necessary for complete conversion of chlorinated VOCs (i.e., TCE) to non-toxic end products (i.e. ethene or ethane). Specifically, the parameters assessed with CENSUS are used as metrics for assessing biodegradation potential and include the following:

- 1. Dehalococcoides population (>104 cells/milliliter groundwater benchmark for effective bioremediation strategy);
- Expression of tceA Reductase gene (TCE degradation);
- 3. Expression of bvcA Reductase gene (VC degradation, prevents cis-DCE accumulation); and



4. Expression of vcrA Reductase gene (VC degradation, prevents cis-DCE accumulation).

In addition to the CENSUS analysis, Bio-Trap® samplers also collect groundwater for characterization of secondary lines of evidence indicative of microbial growth and contaminant degradation including dissolved gas production (ethane, ethene and methane), volatile fatty acids ("VFA") production, standard geochemical parameters and contaminant of concern concentration profiles. Detection of VFAs indicates active fermentation of intrinsic or added organic substrates (i.e. biostimulation) by microbial organisms has occurred and microbial growth is favorable. Changes to the innate contaminant of concern concentration profile, with the production of intermediate degradation products (e.g., cis-1,2,-Dichloroethene and vinyl chloride) and non-toxic end products (e.g., ethene and ethane), can provide a direct line of evidence that bioremediation is occurring. Lastly, the Bio-Trap® sampler includes sub-units that assess the benefit of carbon substrate addition, a critical factor in determining full-scale design parameters. The following text is an exert from the SITE LOGIC Report dated March 25, 2022, and briefly describes each sub-unit that was deployed in MW-1, MW-4, and MW-9 on December 10, 2021:

Monitored Natural Attenuation (MNA) Unit: The purpose of the Control Bio-Trap Unit is to quantify contaminant degrading bacteria and daughter product formation under monitored natural attenuation (MNA) conditions and to serve as a baseline for comparison to BioStim and/or BioAug Units.

BioStim Slow Release Substrate (SRS) Unit: The Biostimulation Bio-Trap Unit is designed to test the hypothesis that electron donor addition will stimulate growth of dechlorinating bacteria and enhance biodegradation.

BioAug SRS Dehalococcoides-containing microbial consortium (SDC-9TM)Unit: The Bioaugmentation Bio-Trap Unit is designed to evaluate bioaugmentation as a treatment technology. The MICRO sampler contains Bio-Sep® beads pre-inoculated with the desired commercial culture. An amendment supplier may also be used to deliver an amendment.

The deployment guidance provided by Microbial Insights, Inc. (Appendix F). The units were retrieved after three months on March 9, 2022, and shipped to Microbial Insights laboratory in Knoxville, Tennessee. Upon receiving the results, Montrose utilized the Microbial Insights Database to assess the degree that bioremediation may be successful based on the measured concentrations of contaminant degrading microorganisms. The Microbial Insights Database allows comparison to more than 32,000 unique groundwater, soil, and sediment sample results in order to answer the questions as to whether the result is low, medium, or high.



3.0 RESULTS EVALUATION

3.1 <u>Data Validation</u>

All data reported by Pace (both the mobile lab and Indianapolis) was reviewed by a Montrose data validation specialist. Pace analytical sheets and laboratory packages are provided in Appendix G. Validation was performed in accordance with the Quality Assurance Project Plan to verify compliance with the required analytical protocols and to determine the qualitative and quantitative reliability of the data. A Montrose Level IIA validation was performed for:

- Holding Time Compliance;
- Laboratory Method Blank;
- Field Blank Contamination;
- Initial and Continuing Calibration Accuracy;
- Matrix Spike/Matrix Spike Duplicate Precision and Recovery;
- Field Duplicate Precision;
- Quantitation/Detection Limits;
- Instrument Blanks;
- Laboratory Control Sample; and,
- Laboratory Duplicate Precision.

All data were acceptable as reported with data validation qualifiers, if necessary. The data validation check sheets and the raw data packages (including copies of chain-of-custody documentation) from the laboratory are included in Appendix G and H, respectively.

3.2 Groundwater Monitoring Results

The analytical results for groundwater samples collected during December 8 and 9, 2021, from the on-Site monitoring wells are presented on Table 6. Eleven (11) monitoring wells were sampled: MW-1, MW-2, MW-3, MW-4, MW-7, MW-9, MW-9D (duplicate), MW-10, MW-11, MW-12, MW-13, and MW-14.

The groundwater sampling data was used to develop isoconcentration maps shown on Figures 5, 6, and 7 to further understand the extent and magnitude of the CVOC plume in groundwater. The IDEM VISL based definition 2021 Screening Level Vapor Exposure (SLVE) was used to interpreting the overall extent of the VOCs plume:



		2021 SLVE ² (ug/L)			
Compound	Standard ¹ (ug/L)	Residential	<u>Industrial</u>		
Benzene	5	28	120		
Ethyl Benzene	700	-	-		
Toluene	1,000	-	-		
Xylenes	10,000	-	-		
cis-1,2-DCE	70	-	-		
Tetrachloroethene	5	6.5	28		
TCE	5	9.1	38		
Vinyl Chloride	2	2.1	35		

^{1. 2018} Remediation Closure Guide (RCG) GW Tap Limit, 2009 RISC TPH Closure Limits

The IDEM 2021 SLVE was used to interpreting the overall extent of the VOC plumes. The results were also compared to 2018 RCG GW Tap Limits because not all compounds have IDEM 2021 SLVE.

Trichloroethene (TCE) was detected at concentrations above the 2018 SLVE at two (2) of the monitoring well locations (MW-1 and MW-4) during the December 2021 groundwater sampling event. The TCE concentrations detected were 49.1 ug/L at MW-1, and 187,000 ug/L at MW-4. Both the 2021 SLVE residential and Industrial standards were exceeded at MW-1 and MW-4. Several other VOCs were detected in the groundwater sample collected at MW-4 including 1,1-Dichloroethane at 795 J ug/L, 1,1-Dichloroethene at 554 ug/L, Chloroform at 149 J ug/L, cis-1,2-Dichloroethene at 267,000 ug/L, toluene at 239 J ug/L, trans-1,2-Dichloroethene at 1,700 J ug/L and Vinyl Chloride at 22,900 ug/L.

Monitoring wells MW-9 and MW-13 were the only other monitoring wells on Site to contain VOCs (other than TCE) in excess of their compound specific limits. Cis-1,2-DCE and Vinyl Chloride were detected in MW-9 at 3,210 ug/L and 957 ug/L, respectively at MW-9. The 2018 RCG GW Tap Limits for cis-1,2-DCE and VC are 70 ug/L and 2 ug/L, respectively. There are no 2021 SLVE residential and Industrial standards for cis-1,2-DCE. VC was detected at 29.1 ug/L at MW-13. The 2021 SLVE residential and Industrial standards for VC are 2.1 ug/L and 35. Monitoring well MW-13 is above the residential SLVE but below the Industrial SLVE.

3.3 Hydropunch Sampling Results

Locations L3-HP, L4-HP, L6-HP, L7, L8, L8A, L9, L9A, L10, L10A, L11, and L11A were sampled using the Geoprobe Hydropunch procedure. The groundwater samples were analyzed for BTEX, cis-1,2-DCE, PCE, TCE, and VC using a field GC operated by Pace Analytical. The analytical results for groundwater samples collected during October 18 - 19, 2021 are shown on Table 7.

^{2.} Indiana Department of Environmental Management 2021 Residential Screening Level Vapor Exposure



The Hydropunch results were used to refined the locations of the new monitoring wells. In addition, the Hydropunch data was used in conjunction with the monitoring well results to develop the isoconcentration discussed in Section 3.5. The data extracted from Table 7 for use in contouring is summarized below and the results that exceeded the 2021 SLVE residential and Industrial standards for TCE and VC (or cis-1,2-DCE Tap Limit) are shown in bold.

Hydro- punch	Location	TCE (ug/L)	Cis-1,2- DCE (ug/L)	VC (ug/L)
L3-HP	Off-Site downgradient and northeast from MW-4 across Kelley Avenue	1	1	
L4-HP	Off-Site downgradient and east-northeast from MW-4 across Kelley Avenue	-	180	270
L6-HP	Off-Site upgradient and south-southeast from MW-4 across Kelley Avenue	6.7	15	3
L7	On-Site upgradient and south of MW-4	280	2,400	430
L8	On-Site upgradient and southwest of MW-4	66	420	200
L8A	L8A was a step-in sample location based on the L8 results.		1,500	330
L9	On-Site cross-gradient and west of MW-4	0.2	8.5	10
L9A	L9A was a step-in sample location based on the L9 results.		71	75
L10	On-Site downgradient northwest from MW-4	0.49	19	23
L10A	L10A was a step-in sample location based on the L10 results.	510	380	170
L11	On-Site downgradient north from MW-4		2.7	2.1
L11A	L11A was a step-in sample location based on the L11 results.	1.1	21	14

To summarize the basic plume configuration, MW-4 is the highest concentration of CVOCs. MW-9 cross-gradient to the east has relatively high concentrations of cis-1,2-DCE and VC. The newly installed monitoring wells are located along the plume edges and bound the plume except upgradient toward the southwest. On-Site monitoring well MW-13 meets the Industrial SLVE for VC, but not the residential SLVE. Sampling the Hydropunch locations and decision tree step-in sample locations provided information on the extent and magnitude of the CVOC plume(s).

3.4 Aqueous Samples from Soil Gas Probes

Aqueous samples were collected at the saturated probe locations L2, L4, L5, L6, L13, L16, and 17 for analysis for BTEX, cis-1,2-DCE, PCE, TCE, and VC using a field GC operated by Pace Analytical. The results are shown on Table 8 and indicate that there are no exceedances of the Target Sub-Slab and Near Source Soil Gas Concentration VISL. Because there were no



exceedances, and because the aqueous samples were collected from vapor pins, this data was not used in developing the isoconcentration maps.

3.5 Extent and Magnitude of VOC/CVOC Impacts

Isoconcentration maps have been developed for the three compounds that exceed the 2018 RCG GW Tap Limit at the Site: Trichloroethene (Figure 5), cis-1,2-Dichloroethene (Figure 6), and Vinyl Chloride (Figure 7). Additional data gathered from the temporary piezometers installed and sampled during the Interim Measures Investigation were used to further define the extents of the groundwater impacts.

As has been observed previously, and as is shown on the isoconcentration maps, the primary area of groundwater impact is localized at MW-4. There is an increase of several orders of magnitude from the upgradient wells (i.e., MW-12) as well as a decrease of several orders of magnitude in the downgradient direction (i.e., MW-3, MW-13, and MW-14). This interpreted area of the groundwater impacts has been largely stable for the previous monitoring events and is comparable for the compounds for which isoconcentration maps were developed. The CVOC concentrations decrease quickly with distance away from MW-4 which indicates that the plume of highly TCE-impacted groundwater is localized and likely contained by low-permeability soils.

Off-Site monitoring well MW-9 contained 1.8 ug/L TCE, 3,210 ug/L cis-1,2-DCE, and 957 ug/L vinyl chloride. The 957 ug/L result for vinyl chloride exceeds the USEPA VISL (55.9 ug/L) and the residential and industrial IDEM SLVEs for vinyl chloride. Groundwater flow is toward the northeast from MW-9 toward MW-11 which is approximately 70 feet downgradient from MW-9. MW-11's groundwater sample was non-detect for CVOCs, including vinyl chloride. The extent of CVOC in groundwater appears to be unbounded to the southeast, but given the low permeability of the subsurface and low rate of groundwater velocity, additional delineation of off-Site impacts at MW-9 is not needed for the purpose of developing and implementing Interim Measures on-Site.

A future work plan will include language to perform routine monitoring of MW-1, MW-9, MW-13, and other wells on an regular basis, and potentially additional delineation, if warranted.

It should be noted that an additional area of CVOC groundwater impact is present around monitoring well MW-1 at the north extent of the Site. TCE (49.1 ug/L) in exceedance of the screening level (5 ug/L) has been noted in this well during the December 2021 event, as well as several historical sampling events. However, the surrounding wells (MW-2, MW-7, and MW-10) do not show a corresponding elevated result. Additional sampling and/or investigation to define the extent of trichloroethene (TCE) contamination in the area of groundwater monitoring well MW-1 will be considered under a separate future work plan.

3.6 Soil Sampling Results

During monitoring well installation, a soil sample was collected from the auger tailings from the 2 to 3-foot depth interval from MW-11, MW-12, and MW-13 for analysis of VOCs and lead. A sample could not be collected from the installation of MW-14 due to no tailings being generated. The



results are presented on Table 9. No VOCs above IDEM RISC Screening levels were detected in the soil samples. The sample from MW-12 contained a lead level of 560 mg/kg which exceeds the Residential IDEM RISC Screening level for lead of 400 mg/kg but is below the commercial/industrial screening level of 800 mg/kg.

3.7 Outfall Z Results

The results of the samples collected at Outfall Z are presented on Table 10. The screening criteria utilized for Outfall Z was the 2021 IDEM RCG GW Tap Limit for the surface water and the RCG Soil Direct Contact Residential and Non-Residential Limit for the sediment. The lead results were found to be below the appropriate screening levels. No VOCs were detected in the sediment sample. Two VOCs were detected in the surface water sample: cis-1,2-Dichlorethene and Vinyl Chloride at concentrations of 5.3 and 2.1 ug/L, respectively. The Vinyl Chloride result is just above the screening level of 2 ug/L.

3.8 Soil Gas Results

Soil gas results were compared to screening levels derived using the USEPA VISL calculator for "Near Source Soil Screening" HQ=1.0 and Target Risk 1x10⁻⁵ which are shown on the following table:

USEPA Vapor Intrusion Screening Level (VISL)							
Near Source Soil Screening" HQ=1.0 and Target Risk 1x10 ⁻⁵							
	Target Sub-Slab and Near-source Soil Gas Concentration VISL (ug/M³)	Target Groundwater Concentration (ug/L)	Is Target Groundwater Concentration < MCL? (Cgw < MCL?)				
Trichloroethylene (TCE)	69.5	5.18	No (5)				
Tetrachloroethylene (PCE)	1,390	57.6	No (5)				
1,2-dichloroethane (I,2-DCE)	-	-	-				
Vinyl Chloride (VC)	55.9	1.47	Yes (2)				
Benzene	120	15.9	No (5)				
Toluene	174,000	19,200	No (1,000)				
Ethyl Benzene	374	34.9	Yes (700)				
Xylenes	3,480	385	Yes (10,000)				



Notes:

- 1. Output generated 27AUG2021:10:49:19
- 2. There is no VISL value for cis I,2-DCE on the EPA VISL website or in IDEM's 2021 Screening Level Vapor Exposure. Source: https://epa-visl.ornl.gov/cgi-bin/visl_search
- 3. 1 No VISL standard exists for cis-1,2-Dichloroethene

A total of twelve (12) soil gas samples were collected and analyzed for BTEX, cis-1,2-DCE, PCE, TCE, and VC using a field GC operated by Pace Analytical. Locations L1, L3, L14, L15, and L18 were collected from soil vapor pins (after helium leak testing) and the results are shown on Table 11. A duplicate sample was taken of L3. Geoprobe PRT system of sample collection was used to collect soil gas samples at a depth of 1.5 feet at locations L2, L4, L5, L6, L13, L16, and 17. Figure 8 shows the locations of the soil gas sampling and any positive detections. The results are shown on Table 12. The results on Table 11 and Table 12 indicate no exceedances of the Target Sub-Slab and Near Source Soil Gas Concentration VISL.

3.9 Utility Bedding Results

Montrose installed three (3) sampling ports within the sewer utility bedding along the east side of Kelley Avenue (locations L16, L17, and L18). No unique utility bedding fill material (i.e., sand/gravel) was identified during the installation of these ports, and it is Montrose's observation that the utility trench was backfilled using the local excavated material. The results are shown on Table 12 and Table 13 and indicate that there are no exceedances of the Target Sub-Slab and Near Source Soil Gas Concentration VISL.

3.10 Manhole Vapor Sampling Results

The ten (10) manhole sample results are shown on Table 13. The manholes are designated as Manhole B, C, E, F, G, H, I, J, L, and MH-10. All of the ten sampled manholes were found to exceed the Target Sub-Slab and Near Source Soil Gas Concentration VISL for TCE (69.5 ug/m3). Additionally, five of the ten sampled manholes exceeded the VISL for VC (55.9 ug/m3). All manholes had detectable concentrations of DCE ranging from 23 to 8,900 ug/m3. Other compounds detected in at least one manhole sample include Chloroform, Tetrachloroethene, and Toluene.

The vapor concentrations in manholes near the northeast corner of the Site (Manhole J, L, and MH-10) reported TCE concentration roughly five times higher than cis-1,2-DCE and low or no concentration of VC. Manhole J had the highest concentration of TCE at 5,000 ug/m3. Vapor results in manholes along North Kelley Avenue reported TCE concentrations lower than the cis-1,2-DCE in six out of seven samples. The following information was obtained by former employee of the Site and provided to Montrose.



Manhole Information Summary							
Man-Hole	<u>Street</u>	<u>Type</u>	Outlet & Flow Direction	Manhole Inlets Notes			
Α	Kelley	Storm	North	South	NS-Open Grate		
Man-Hole	<u>Street</u>	<u>Type</u>	Outlet & Flow Direction	Manhole Inlets	<u>Notes</u>		
В	Kelley	Sanitary	North	South	Sampled		
С	Kelley	Sanitary	North	South & East	Sampled		
D	Kelley	Storm	North	South & SW & SE	NS-Open		
E	Kelley	Sanitary	North	South & Southwest	Sampled		
F	Kelley	Sanitary	Northeast	South	Sampled		
G	Kelley	Sanitary	Northeast	SWW & SW & (2) East	Sampled		
Н	Kelley	Sanitary	Northwest	Southwest	Sampled		
I	Kelley	Sanitary	North	Southeast	Sampled		
J	Washington	Sanitary	West	East	Sampled		
K	Washington	Storm	West	SW & (2) SSW & Northeast	NS-Open Grate		
L	Washington	Sanitary	West	NE & (2) South Sampled SE			
М	Hoke	Storm	East	Northwest	NS-Open Grate		
N	Hoke	Storm	Southeast East	Northwest West	NS-Open Grate		
Z	Kelley	Storm	North	South	Outfall Sampled		
MH-10	On-Site	Storm	West	South & West	Sampled		

Figure 9 shows the location of the manhole, the location of the sanitary sewer and storm water buried utilities, and the direction of flow based on the information reviewed by Montrose.

3.11 Groundwater Flow Discussion

In order to calculate the rate of groundwater movement (or groundwater horizontal velocity), the following properties must be understood: 1) the horizontal hydraulic conductivity, 2) the groundwater gradient, and 3) the porosity of the subsurface.

3.11.1 Horizontal Hydraulic Conductivity

Hydraulic conductivity is a function of the ability of materials to convey quantity of groundwater under a hydraulic gradient with respect to time. Hydraulic conductivity is a property of soil that describes the ease with which a fluid can move through pore spaces. Slug tests were conducted



to calculate the horizontal hydraulic conductivity (K) proximal to six on-Site groundwater monitoring wells. The results shown on Table 3.

3.11.2 Groundwater Gradient

The hydraulic gradient represents the pressure head differential between two points which keeps groundwater moving from one point to another. As stated in Section 1.3, the well gauging data collected on December 8, 2021, indicates the hydraulic gradient in the central portion of the Site is 0.007 toward the north northwest. In the northern portion of the site near MW-1, MW-2, and MW-7, the hydraulic gradient steepens slightly to 0.013 and turns to the north northeast. On the eastern portion of the Site in the area of groundwater contamination near MW-4, groundwater flows north under a hydraulic gradient of 0.013. In the area along North Kelley Avenue, the groundwater flow turns to the northeast under a gradient of 0.010.

3.11.3 Porosity

Porosity is the ratio of openings and voids to the total volume of geologic material. Clay is the most porous sediment but is the least permeable. Clay usually acts as an aquitard, impeding the flow of water. The porosity of clay can vary from 40 to 70 percent and silt porosity ranges from 35 to 50 percent (Freeze, A. and Cherry, J., *Groundwater*, Prentice Hall, Inc., Englewood Cliffs, New Jersey, 1979, p. 37.). According to the Wikipedia entry

(http://en.wikipedia.org/wiki/Porosity): "Typical bulk density of clay soil is between 1.1 and 1.3 g/cm³. This calculates to a porosity between 0.58 and 0.51." Because of the presence of silts and occasional sand in the clay soil noted in the well logs and boring logs for this Site, Montrose assumed a porosity of 0.50 to calculate groundwater velocity at the site.

3.11.4 Groundwater Velocity

The rate at which groundwater will travel horizontally, or the horizontal groundwater velocity (V), can be estimated using the following equation:

V = <u>K x i</u> where: K= Hydraulic Conductivity (feet/day)
n i = Hydraulic gradient (ft/ft)
n = effective porosity

At MW-1, the groundwater velocity is:

V = 0.0088 feet/day x 0.013 feet/feet or 2.288 x 10⁻⁴ feet per day 0.50

This is equivalent to 8.351×10^{-2} feet per year. At this velocity, the groundwater would need a period of 12 years in order to flow a distance of 1.0 foot. Using the same formula for the remaining wells that were slug tested yields the following summary of groundwater velocities at the Site:



Well	Calculated	Gradient	Groundwater	Groundwater	Time to	Flow
	K	I	Velocity	Velocity	Travel	Direction
	(feet/day)	(feet/feet)	(foot/day)	(foot/year)	1.0 foot	
					Distance in	
					Subsurface	
					(years)	
MW-1	0.0088	0.013	0.0002288	0.083512	12	NNE
MW-3	0.0078	0.013	0.0002028	0.074022	13.5	NNE
MW-4	0.0033	0.010	0.000066	0.02409	41.5	NE
MW-7	0.0004	0.013	0.0000104	0.003796	263.4	NNW
MW-9	0.0015	0.010	0.00003	0.01095	91.3	NE
MW-10	0.0012	0.013	0.0000312	0.011388	87.8	NNE

Note: Based on porosity n = 0.50

The shallow groundwater velocities at this Site are extraordinary slow. According to the USGS, a groundwater velocity of 1.0 foot per day is a high velocity and groundwater velocities can be as low as 1.0 foot per year or 1.0 foot per decade. The groundwater velocities of one foot per several decades or longer indicates that the groundwater at the Site is "old" groundwater, meaning there has been long period since the water recharged the subsurface.

The high-age zone of groundwater is in an unconsolidated aquifer with a laterally extensive low-permeability unit (glacial tills) that also have a low groundwater gradient. This high-age zone of groundwater is not a hydraulic stagnation point but is associated with the 65 to 190-foot thick till cap overlaying the Tipton Complex Aquifer System. In this setting, it is not uncommon for overturned groundwater ages, which is where younger groundwater is under older groundwater. The significance is that high age groundwater accumulates and contains contaminants.

3.12 Microbial Insights Results

The assemblies deployed in all three wells consisted of three Bio-Trap units each: (i) a control monitored natural attenuation (MNA) unit with no exogenous amendment, (ii) a BioStim unit amended with SRS as the electron donor, and (iii) a BioAug unit amended with SRS as the electron donor and the exogenous SDC-9 dechlorinating bacterial culture. The results from the In Situ Microcosm study are summarized on Table 14. The following information is summarized from the Microbial Insights report contained in Appendix F.

MW-1

The microbial and functional gene data indicate that the potential for the complete reductive dechlorination of PCE and TCE to ethene at well MW-1 is moderate under MNA conditions, low in the BioStim unit, and high under the BioAug condition assessed. As previously stated, the TCE impact at MW-1 appears to be discontinuous with the CVOC plume at MW-4. MW-1 is over 500 feet from MW-4 and it is unlikely that the TCE in MW-1 is a result of contaminant transport by groundwater flow down hydraulic gradient.



MW-4

The results indicate that the both SRS amendment and SDC-9 bioaugmentation stimulated the overall growth of DHC and functional genes. The results suggest that complete reductive dechlorination to ethene occurred in all three Bio-Trap ISM units during the deployment period.

MW-9

The results suggest that complete reductive dechlorination to ethene occurred in all three Bio-Trap ISM units during the deployment period. The results indicate an increase in the genetic potential for the complete anaerobic reductive dechlorination of PCE and TCE under bioaugmentation with SDC-9 and SRS amendment at this well location.



4.0 CONCEPTUAL SITE MODEL

A Conceptual Site Model (CSM) outlines potential source areas, transport mechanisms, environmental media affected, potential exposure pathways, and potential exposure routes to potential receptors. The CSM considers current Site conditions and surrounding land use, as well as the most likely future conditions and land use. The CSM is prepared in accordance with IDEM technical guidance created February 14, 2014.

4.1 Site Setting

The Site is situated in the 65 to 190-foot thick till cap overlaying the Tipton Complex Aquifer System. Nearly the entire site is covered by pavement or building pads that were associated with previous site manufacturing operations. Soils are brown silt loam surface layer on silty clay loam to clay loam subsoil. Evidence of shallow filling (typically less than 1 to 2 feet) was observed beneath most of the paved areas and building pads typically in the form of crushed stone that is consistent with material that would be utilized for pavement and building pad subbase. The depth to the water table measured in the fourteen (14) monitoring wells at the site range from 1.89 to 8.83 feet below top of casing. The shallow groundwater velocities are extraordinary slow, indicating the groundwater is old; high-age groundwater accumulates and contains the migration of contaminants.

4.2 Sources of Contaminants

4.2.1 Former Operations

In 1963, General Battery Corporation began the manufacturing of lead-acid batteries for use in automotive, golf cart, marine and industrial applications. At its peak the facility produced over 12,000 automotive batteries per day. The battery manufacturing process used metallic lead that was received at the facility, melted, and cast into grids and posts.

Based on the documented operational history and an understanding of the character of lead mobility and transport, the most significant potential sources of contamination at the facility during its operational history were erosion and transport of lead-bearing solids by storm water runoff; fugitive dust emissions from traffic and production areas; uncovered waste pile areas or miscellaneous spills. In October 2012, the facility was decontaminated followed by demolition of the above grade structures. The decontamination and demolition project was completed in January 2013.

4.2.2 Soil

Comprehensive RI soil sampling across the Site indicates that elevated lead in soil generally is present to the south and east of the former manufacturing areas, in addition to the manufacturing portion of the site. The detections found above the IDEM RISC Non-Residential Direct Contact Standard are only found within the top 2 feet below the surface in shallow fill. Arsenic concentrations in soil are below IDEM RISC Non-Residential Direct Contact Standard, with the



exception of two individual samples found within the top 2 feet below the surface in shallow fill. As stated, nearly the entire site is covered by pavement or building pads. The Site is situated within a fenced and secured area to prevent unauthorized access.

4.2.3 Sediment

There are no surface water features or wetlands on the site. Sediment that had accumulated in the storm water manholes and pipes was flushed and cleaned during site demolition in 2012. During the SSI, sediment samples collected from the stormwater piping network indicated concentrations of lead in two of four locations in excess of the IDEM RISC Non-Residential Direct Contact Standard (800 mg/kg).

During the 2021 IMI, a sediment sample was collected adjacent to Outfall Z and the lead result was 105 mg/kg. No VOCs were detected in the sediment sample. Two VOCs were detected in the surface water sample: cis-1,2-Dichlorethene and Vinyl Chloride at concentrations of 5.3 and 2.1 ug/L, respectively. The Vinyl Chloride result is just above the screening level of 2 ug/L.

4.2.4 Groundwater

Previous investigations of groundwater in May and July 2018 showed impact to groundwater in on-Site monitoring wells from VOCs and CVOCs. Recently, the TCE concentrations detected were 49.1 ug/L at MW-1, and 187,000 ug/L at MW-4. Off-Site monitoring well MW-9 contained 1.8 ug/L TCE, 3,210 ug/L cis-1,2-DCE, and 957 ug/L vinyl chloride. The CVOC concentrations decrease quickly with distance away from MW-4 which indicates that the plume of highly TCE-impacted groundwater is localized and likely contained by low-permeability soils. The 957 ug/L result for vinyl chloride exceeds the USEPA VISL (55.9 ug/L) and the residential and industrial IDEM SLVEs for vinyl chloride. Groundwater flow is toward the northeast from MW-9 toward MW-11 which is approximately 70 feet from MW-9 and is non-detect for CVOCs, including vinyl chloride. The extent of CVOC in groundwater appears to be unbounded to the southeast, but given the low permeability of the subsurface and low rate of groundwater velocity, additional delineation of off-Site impacts at MW-9 is not needed for the purpose of developing and implementing Interim Measures on-Site. A future work plan will include language to perform routine monitoring of MW-9 and other wells on an regular basis, and potentially additional delineation, if warranted.

The source of CVOCs is suspected to be associated with AOC-3/UST-2 because CVOC impacts were within 20 feet of UST-2. The rate of groundwater flow is extremely slow due to laterally extensive low-permeability glacial tills combined with a low groundwater gradient. Slug test data indicates that it can take decades for groundwater to flow 1.0 foot horizontally at the Site. Discontinuous sand stringers, and zones of preferred pathways may provide for faster localized groundwater transport but this is not evident based on the existing monitoring well network, measured potentiometric groundwater surface, and interpreted isoconcentration maps.

The TCE impact at MW-1 (49.1 ug/L) appears to be discontinuous with the impact at MW-4. MW-1 is over 500 feet from MW-4 and it appears unlikely that the TCE in MW-1 is a result of



contaminant transport by groundwater flow down hydraulic gradient. The wells surrounding MW-1 (MW-2, MW-7, and MW-10) do not show a corresponding elevated result. Additional sampling and/or investigation to define the extent of trichloroethene (TCE) contamination in the area of groundwater monitoring well MW-1 will be considered under a separate future work plan.

The In Situ Microcosm study suggests there is a strong potential for the complete anaerobic reductive dechlorination of PCE and TCE under bioaugmentation with SDC-9 and SRS amendment at this Site. The complicating issue may be that the subsurface may not be able to adequately transmit amendments through the area of impact given the extensive low-permeability soil in the subsurface. Future sampling and/or investigations will be necessary to evaluate and identify the permanent solutions to assure protection of groundwater at, and beyond, the property boundary and also identify how potential exposures to off-site receptors will be mitigated, if warranted.

4.2.5 Soil Gas

The results of twelve (12) soil gas samples indicate that there are no exceedances of the Target Sub-Slab and Near Source Soil Gas Concentration VISL. Locations L5 and L17 were close to off-Site monitoring well MW-9 and there were no CVOVs in soil gas these locations. As stated, there is an exceedance of IDEM Vapor Intrusion screening levels in MW-9 for vinyl chloride.

4.2.6 Sewer Gas

The ten (10) sewer gas samples collected from manhole locations exceeded the Target Sub-Slab and Near Source Soil Gas Concentration VISL for CVOCs. Manhole J had the highest concentration of TCE at 5,000 ug/m³. Manhole F had the highest concentration of cis-1,2-DCE at 8,900 ug/m³ and the highest concentration of VC at 1,500 ug/m³. If the on-Site CVOC-impacted groundwater is the source of CVOCs in the sewer, the mechanism of CVOC entry into the interior of the sewer is unknown. The source does not appear to be soil gas infiltration. No unique utility bedding fill material (i.e., sand/gravel) was identified during the investigation at three locations and it appears the utility trench was backfilled and compacted using the local excavated material when the sewer line was installed. The sanitary sewer lines pass through CVOC impacted groundwater near the intersection of North Kelley Avenue and East McClurg Street. The soil gas samples taken adjacent to (L18) or directly above (L16 and L17) the sewer pipe did not contain soil gas results above VISL. No CVOCs were detected in L16, L17, or L18.

It is possible that impacted groundwater from the Site is leaking into the sewer and volatizing the CVOC vapors into the sewer. It is also possible that the CVOCs in the interior of the sewer piping is from an off-Site source. The issue of CVOC vapors in the sewer system is complex because the sewer is the receptor of pollutants from a large number of potential sources.

The soil gas samples taken adjacent to the sewer pipe did not contain soil gas results above VISL. CVOC vapors in sewer lines should not be a source for indoor CVOC vapor intrusion because modern plumbing systems has sewer traps to prevent gases contained in wastewater, the public sewer, or septic tanks from escaping and entering residential housing. A prudent method to further



investigate for evidence of COVC impacted groundwater from potentially infiltrating the sewer would be to run a camera through the sewer, possibly after water-jetting the line, to assess for potential pipe connections from the Site or obvious breaches in the sewer line. This can be considered for inclusion in the Interim Measures Work Plan.

4.3 Pathways

A pathway evaluation identifies which exposure pathways are open and which are incomplete. Potential complete exposure pathways to regulated substances located in soil and groundwater on the Site include:

- Direct contact (inhalation, ingestion) to soil;
- Soil leaching to groundwater;
- Groundwater ingestion; and
- Vapor Intrusion

Shallow soil ingestion, inhalation, and dermal contact is limited to construction worker scenarios and because the impacts are located under pavement, building slabs, or other caps. The potential direct contact pathway to soil is deemed incomplete based on capping and PPE procedures which limit exposure to soil encountered in excavations. Additionally, appropriate institutional and deed controls can be put into place to manage the construction worker exposure scenario.

Soil leaching to groundwater has been assessed through the installation and sampling of the network of groundwater monitoring wells. Elevated lead levels in soil at depth (1 to 2 ft below ground surface), are limited to the a few samples to the south and west of the manufacturing area. Soil leaching is not considered a complete pathway because of the shallow nature of the contaminant, the Site capping, the slow rate of groundwater migration, and the fact that no lead over screening levels was detected in groundwater samples from monitoring wells.

Groundwater in on-Site monitoring wells is impacted with VOCs and CVOCs. Groundwater was evaluated to assess the CVOC plume, groundwater flow direction, and groundwater velocity to evaluate the potential for groundwater in exceedance of the standards to migrate off-site. The groundwater flow rate is extremely slow due to laterally extensive low-permeability glacial tills combined with a low groundwater gradient. Data indicates that it can take decades for groundwater to flow 1.0 foot horizontally at the Site. The Site was supplied with potable water by the City of Frankfort municipal supply (Frankfort Water Works). The municipal water supply lines to the Site were cut and capped in October 22, 2012 as part of the demolition. Due to the plume stability and lack of groundwater users, groundwater ingestion is not a complete pathway.

Vapor intrusion (VI) typically is risk driver for sites with chlorinated impacts. This Site is vacant and all aboveground structures have been demolished. Practical experience has indicated that VI stems from relatively significant sources in close proximity to the slab or the presence of preferential pathways (e.g., sumps, pipes, or openings). There is an off-Site exceedance of IDEM Vapor Intrusion screening levels in MW-9 for vinyl chloride. This exceedance was addressed by soil gas sampling in the vicinity of MW-9 to further assess the potential for vapor exposure.



Locations L5 and L17 were close to off-Site monitoring well MW-9 and there were no CVOVs in soil gas these locations. The results of twelve (12) soil gas samples indicate that there are no exceedances of the Target Sub-Slab and Near Source Soil Gas Concentration VISL. Based on the data collected, the exposure pathway for VI from the Site appears to be incomplete.

CVOC vapors in sewer lines should not be a source for indoor CVOC vapor intrusion because modern plumbing systems has sewer traps to prevent gases contained in wastewater, the public sewer, or septic tanks from escaping and entering residential housing.

4.4 Receptors

4.4.1 On-Site Receptors

The future construction worker scenario is the only contact with a receptor to on-Site CVOC impacts. The Site is essentially capped. As a technology, capping can be quite effective at interrupting the human health exposure.

4.4.2 Off-Site Receptors

Residential properties lie across the street from the Site on North Hoke and Kelly Avenues; as well as on the opposite side of the railroad tracks to the south. Michigantown Road is immediately north of the Site and has several light industrial/ commercial properties located in proximity to the Site.



5.0 CONCLUSIONS AND RECOMMENDATIONS

5.1 Conclusions

Groundwater

Groundwater in on-Site monitoring wells is impacted with VOCs and CVOCs. Recently, the TCE concentrations detected were 49.1 ug/L at MW-1, and 187,000 ug/L at MW-4. Off-Site monitoring well MW-9 contained 1.8 ug/L TCE, 3,210 ug/L cis-1,2-DCE, and 957 ug/L vinyl chloride. The TCE impact at MW-1 appears to be discontinuous with the CVOC plume at MW-4. MW-1 is over 500 feet from MW-4 and it is unlikely that the TCE in MW-1 is a result of contaminant transport by groundwater flow down hydraulic gradient. The CVOC concentrations decrease quickly with distance away from MW-4 which indicates that the plume of TCE impacted groundwater is localized and likely contained by low-permeability soils. The source of CVOCs is suspected to be associated with AOC-3/UST-2 because CVOC impacts were within 20 feet of UST-2.

Groundwater was evaluated to assess the CVOC plume, groundwater flow direction, and groundwater velocity to evaluate the potential for groundwater in exceedance of the standards to migrate off-site. The groundwater flow rate is extremely slow due to laterally extensive low-permeability glacial tills combined with a low groundwater gradient. Data indicates that it can take decades for groundwater to flow 1.0 foot horizontally at the Site, and such high-age groundwater will accumulate and contain contaminants. The In Situ Microcosm study suggests there is a strong potential for the complete anaerobic reductive dechlorination of PCE and TCE under bioaugmentation with SDC-9 and SRS amendment at this Site. The issue is the ability of the subsurface to transmit amendments through the area of impact given the extensive low-permeability soil in the subsurface.

Soil Gas

The results of twelve (12) soil gas samples indicate that there are no exceedances of the Target Sub-Slab and Near Source Soil Gas Concentration VISL.

Sewer vapors

The sewer gas samples collected from manhole locations exceeded the Target Sub-Slab and Near Source Soil Gas Concentration VISL for CVOCs. No unique utility bedding fill material (i.e., sand/gravel) was identified during the investigation at three locations and it appears the utility trench was backfilled and compacted using the local excavated material. The soil gas samples taken adjacent to the sewer pipe did not contain soil gas results above VISL. The issue of CVOC vapors in the sewer system is complex because the sewer is the receptor of pollutants from a large number of potential sources.

CVOC vapors in sewer lines should not be a source for indoor CVOC vapor intrusion because modern plumbing systems has sewer traps to prevent gases contained in wastewater, the public sewer, or septic tanks from escaping and entering residential housing.



In Situ Microcosm study

The In Situ Microcosm study suggests there is a strong potential for the complete anaerobic reductive dechlorination of PCE and TCE under bioaugmentation with SDC-9 and SRS amendment at this Site. The complicating issue may be that the subsurface may not be able to adequately transmit amendments through the area of impact given the extensive low-permeability soil in the subsurface.

5.2 Recommendations

Elevated lead in the shallow surface soils at the surface water discharge locations should be delineated and removed, or stabilized using capping, fencing, silt fence or similar techniques as Corrective Action alternatives. Additionally, appropriate institutional and deed controls can be put into place.

Based on the results of this study, the CVOC Remediation Evaluation dated January 19, 2021, will be revisited to further evaluate the potential groundwater remedial actions that may be employed at the Site.

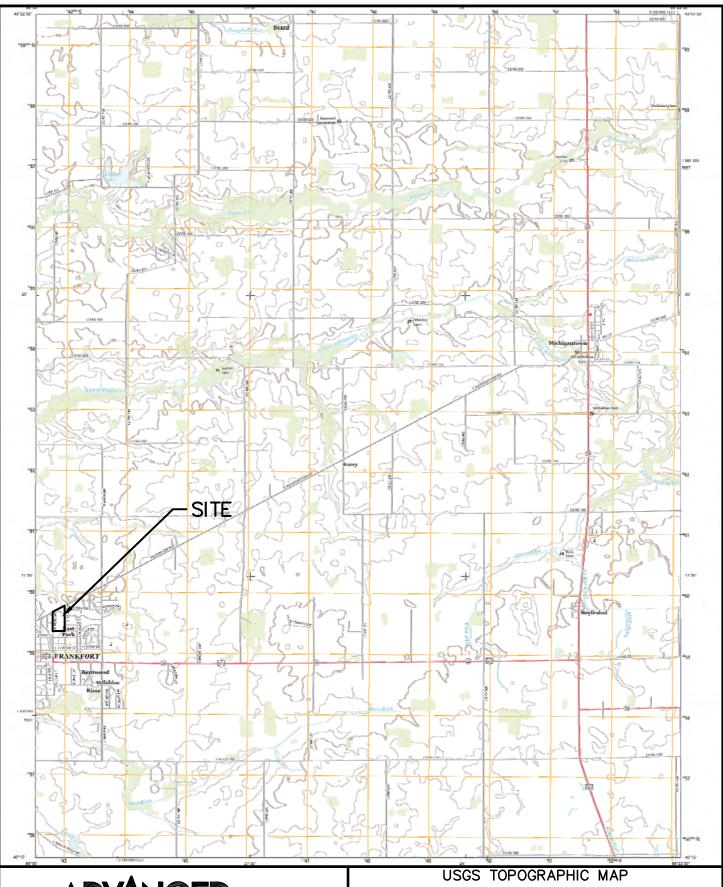
Additional sampling and/or investigation to define the extent of trichloroethene (TCE) contamination in the area of groundwater near monitoring wells MW-1 and MW-13 will be considered under a separate future work plan.

Future sampling and/or investigations will be necessary to evaluate and identify the permanent solutions to assure protection of groundwater at, and beyond, the property boundary and also identify how potential exposures to off-site receptors will be mitigated, if warranted.

A prudent method to further investigate for evidence of COVC impacted groundwater from potentially infiltrating the sewer would be to run a camera through the sewer, possibly after waterjetting the line, to assess for potential pipe connections from the Site or obvious breaches in the sewer line. This can be potentially be conducted during Interim Measures or in a subsequent post Interim Measures investigation phase.



FIGURES

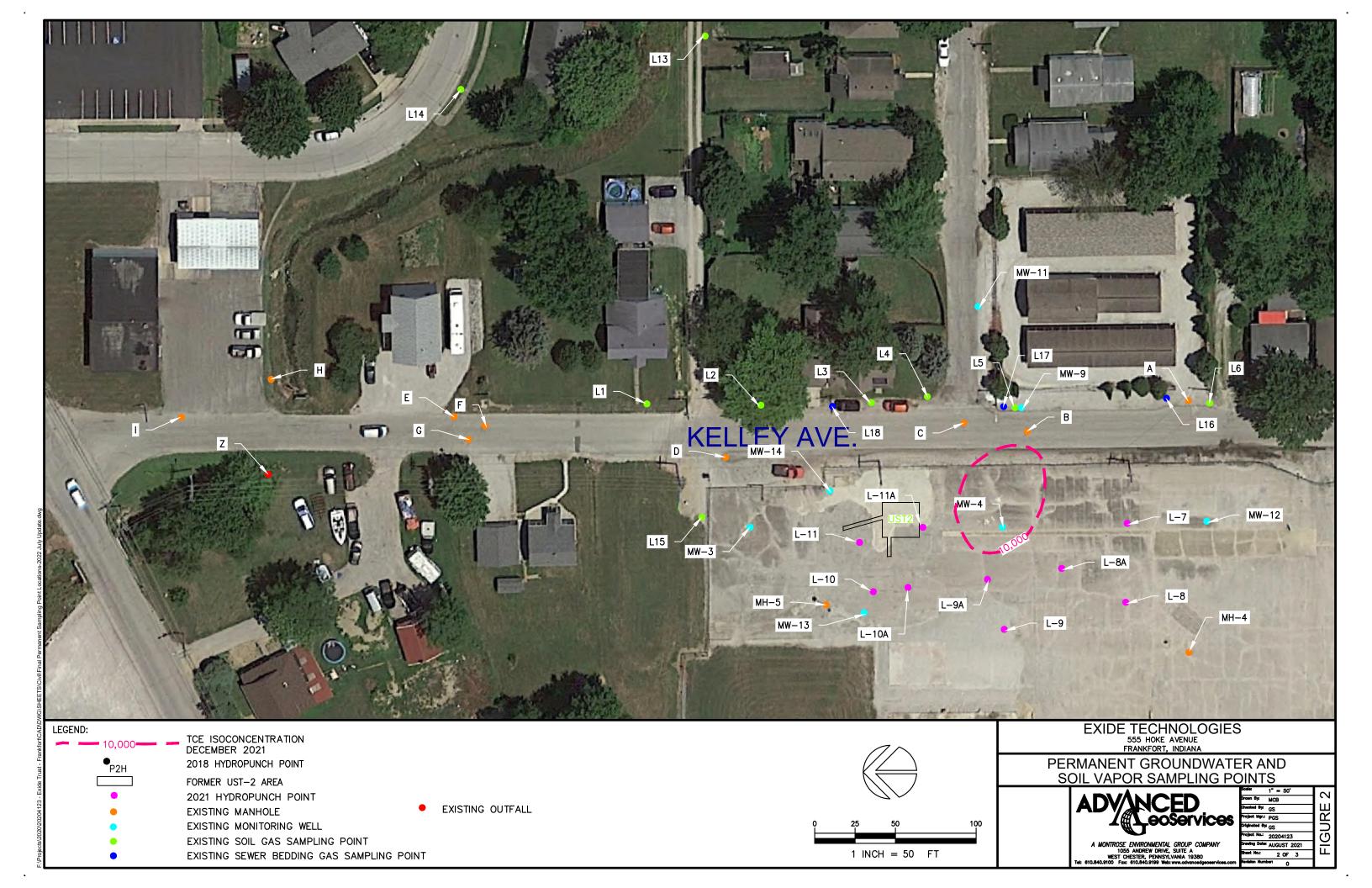


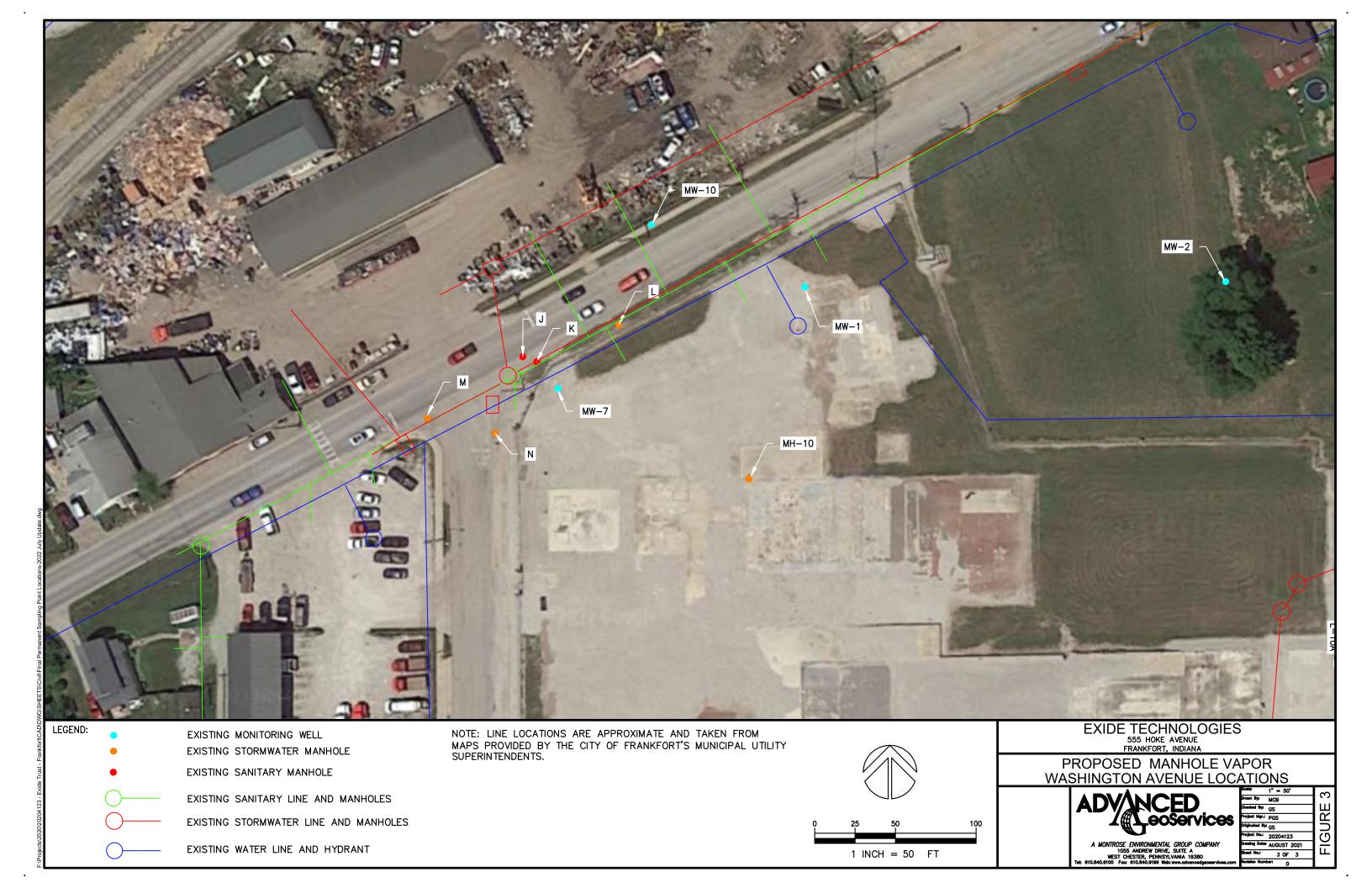


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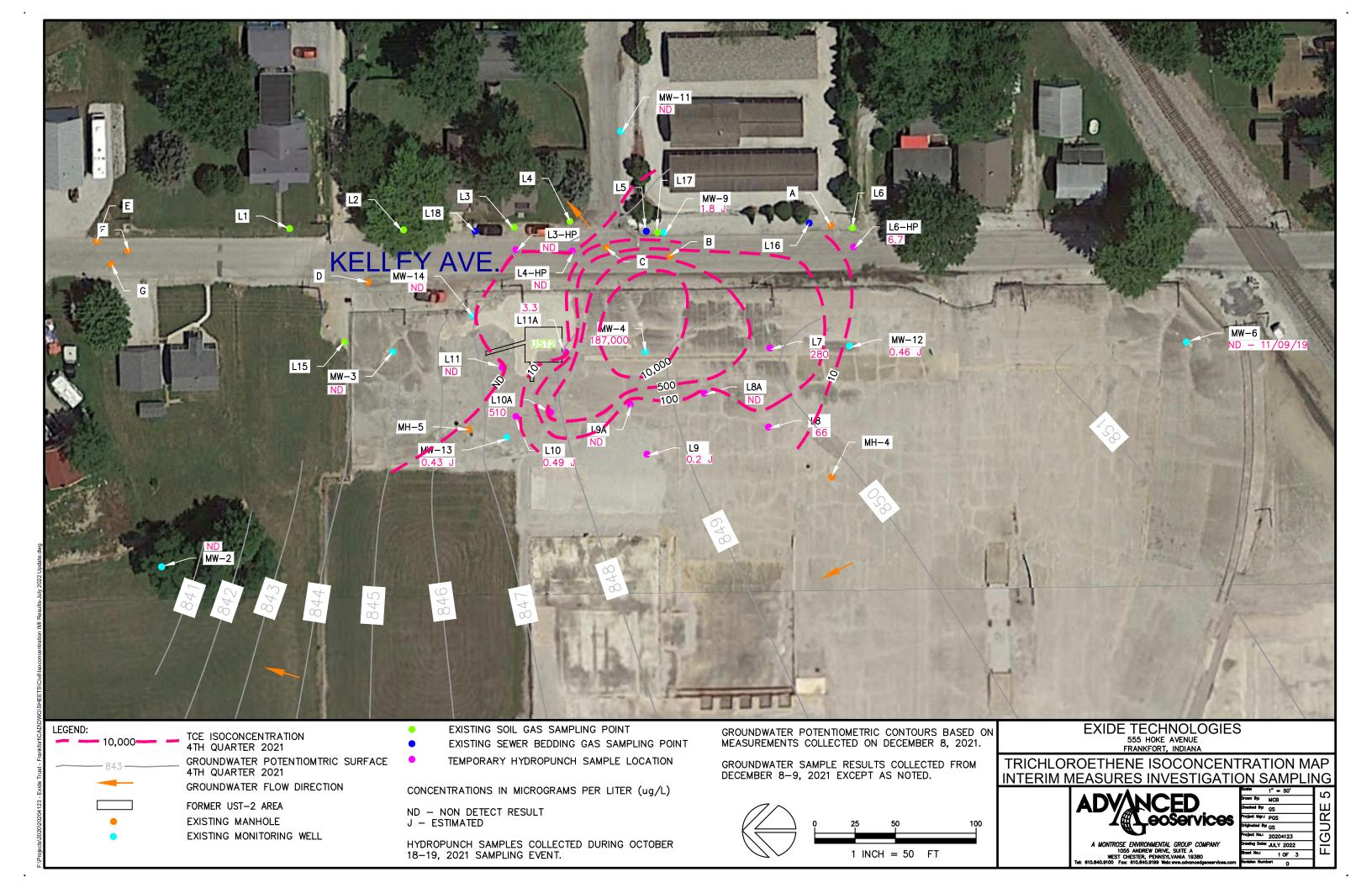


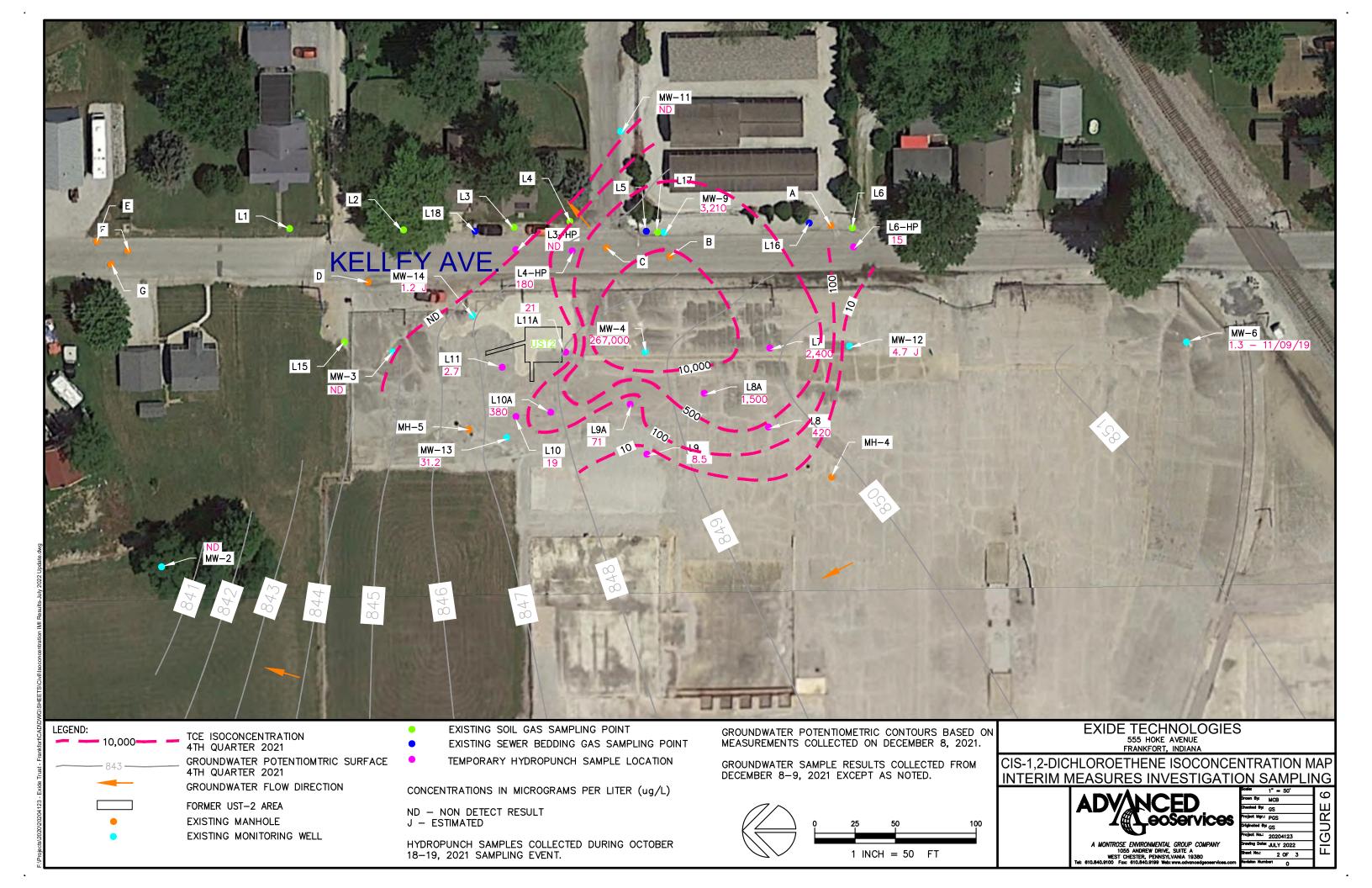


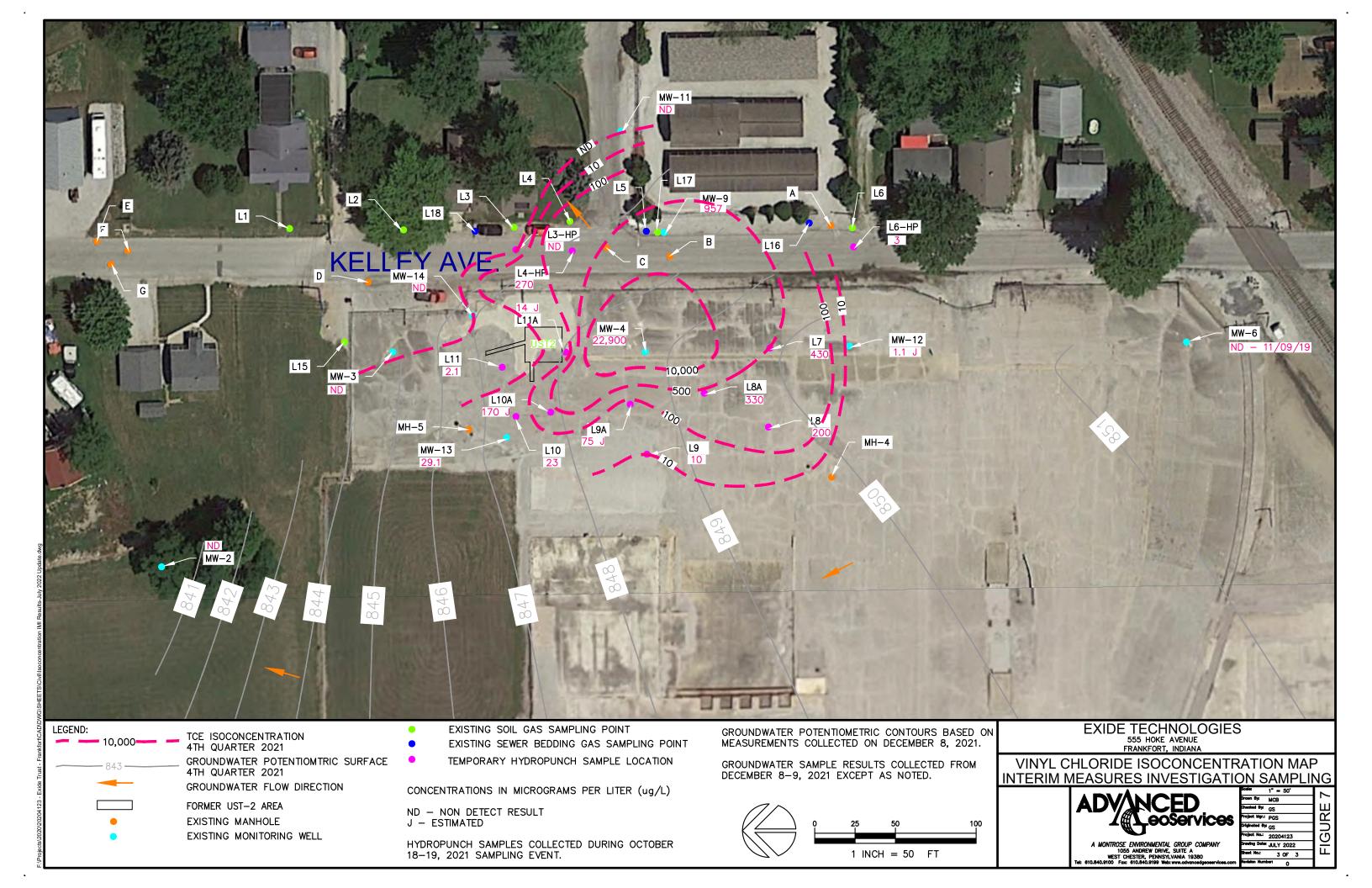


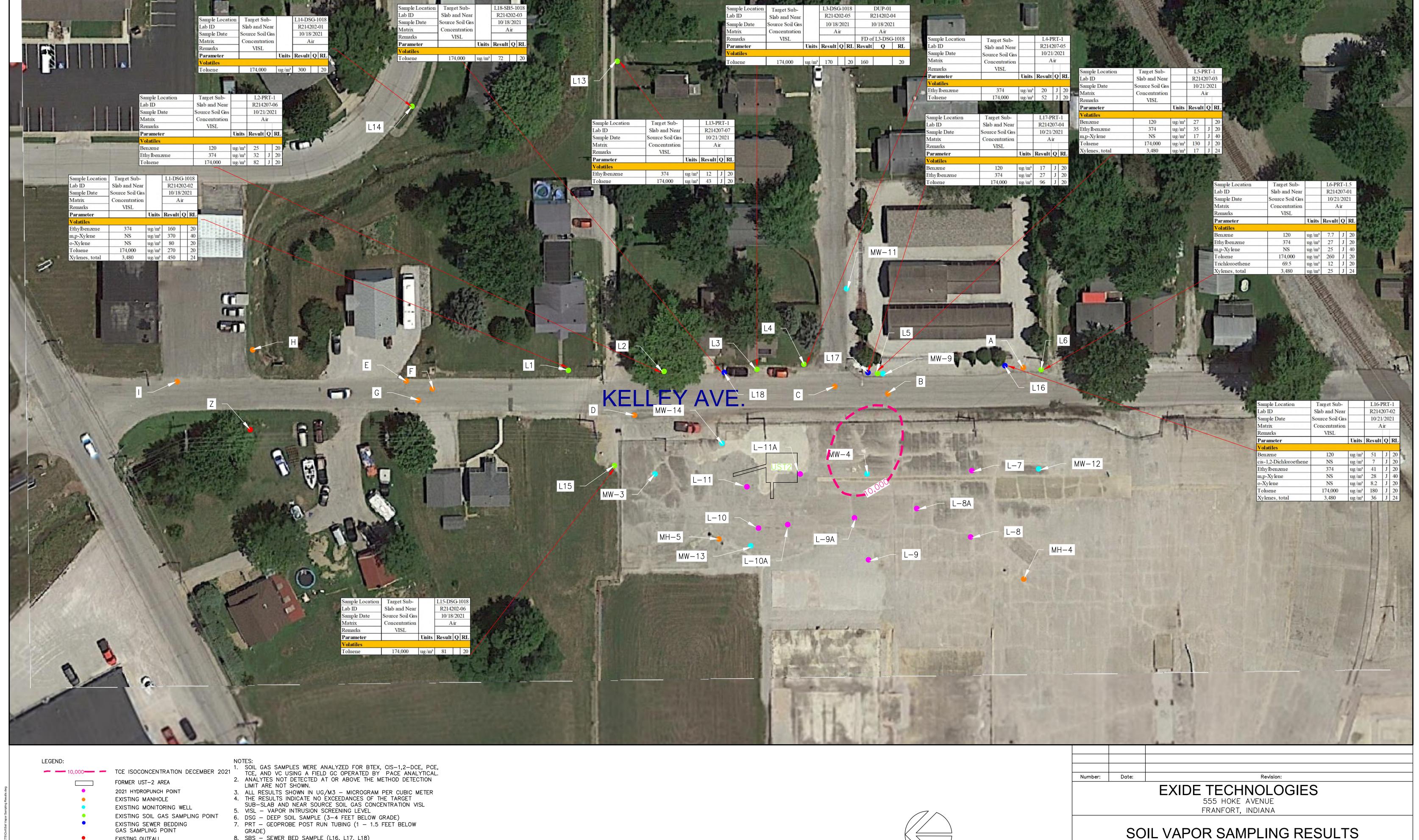
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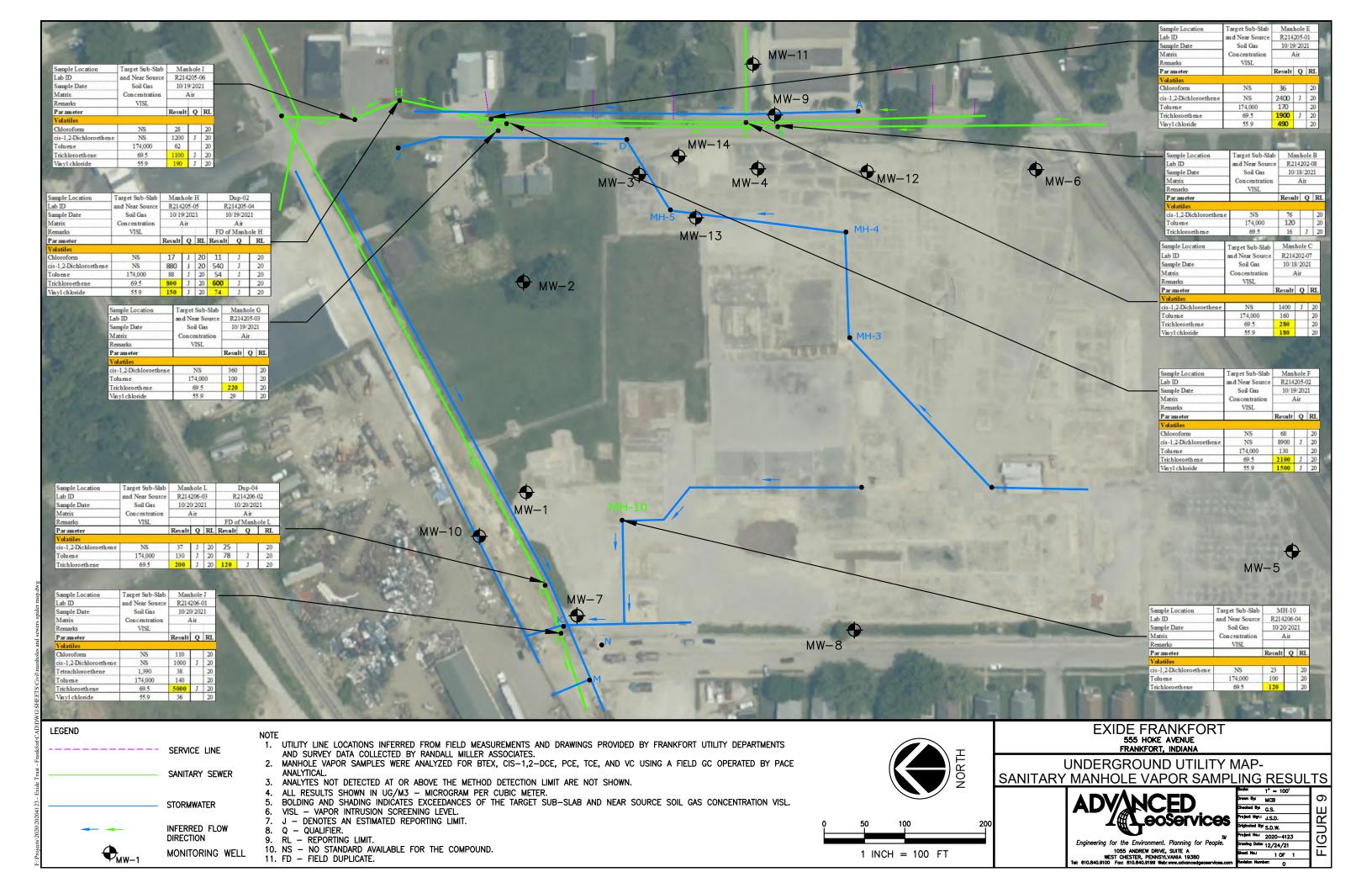
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EXISTING OUTFALL

8. SBS — SEWER BED SAMPLE (L16, L17, L18)
9. J — DENOTES AN ESTIMATED REPORTING LIMIT

11. RL — REPORTING LIMIT
12. NS — NO STANDARD AVAILABLE FOR THE COMPOUND
13. FD — FIELD DUPLICATE

10. Q - QUALIFIER





TABLES

Table 1 Sampling Plan Decision Tree Exide Technologies Frankfort, Indiana

Sample Location	Type of Sample	Decision
Location 1	Soil Gas/ Potential Groundwater	If results are above VISL values, Hydropunch groundwater sampling will be conducted at the location with the highest exceedance with TCE as primary deciding indicator. The
Location 2	Soil Gas/ Potential Groundwater	groundwater will be analyzed with the Field GC for TCE, PCE, cis-1,2-DCE, VC, and BTEX.
Location 2A	Groundwater	If soil gas results at Locations 1 and 2 are below the VISL values, a Hydropunch boring will be completed at Location 2A and groundwater will be field analyzed for TCE, PCE, cis-1,2-DCE, VC, and BTEX. If the Hydropunch groundwater result for TCE is >600 ug/L, then Hydropunch groundwater sampling adjacent to Locations 2 will be conducted.
Location 3	Soil Gas/ Groundwater	No decision - collect both soil gas and Hydropunch groundwater sample for field GC testing for TCE, PCE, cis-1,2-DCE, VC, and BTEX.
Location 4	Soil Gas/ Groundwater	No decision - collect both soil gas and Hydropunch groundwater sample for field GC testing for TCE, PCE, cis-1,2-DCE, VC, and BTEX.
Location 5	Soil Gas	No decision - collect only soil gas sample for field GC testing for TCE, PCE, cis-1,2-DCE, VC, and BTEX.
Location 6	Soil Gas/ Groundwater	No decision - collect both soil gas and Hydropunch groundwater sample for field GC testing for TCE, PCE, cis-1,2-DCE, VC, and BTEX.
Location 7	Groundwater	Collect groundwater sample for Field GC testing for TCE, PCE, cis-1,2-DCE, VC, and BTEX. If TCE is >600 ug/L, sample groundwater via Hydropunch at Location 7B. If TCE is <100 ug/L, Hydropunch sample at Location 7A. Groundwater samples analyzed using Field GC for TCE, PCE, cis-1,2-DCE, VC, and BTEX.
Location 7A	Groundwater	Only conduct Hydropunch sampling if TCE in groundwater at Location 7 is <100 ug/l
Location 7B	Groundwater	Only conduct Hydropunch sampling if TCE in groundwater at Location 7 is >600 ug/l
Location 8	Groundwater	Collect groundwater sample for Field GC testing for TCE, PCE, cis-1,2-DCE, VC, and BTEX. If TCE is >600 ug/L, sample groundwater via Hydropunch at Location 8B. If TCE is <100 ug/L, Hydropunch sample at Location 8A. Groundwater samples analyzed using Field GC for TCE, PCE, cis-1,2-DCE, VC, and BTEX.
Location 8A	Groundwater	Only conduct Hydropunch sampling if TCE in groundwater at Location 8 is <100 ug/l
Location 8B	Groundwater	Only conduct Hydropunch sampling if TCE in groundwater at Location 8 is >600 ug/l

Table 1 Sampling Plan Decision Tree Exide Technologies Frankfort, Indiana

Sample Location	Type of Sample	Decision
Location 9	Groundwater	Collect groundwater sample for Field GC testing for TCE, PCE, cis-1,2-DCE, VC, and BTEX. If TCE is >600 ug/L, sample groundwater via Hydropunch at Location 9B. If TCE is <100 ug/L, Hydropunch sample at Location 9A. Groundwater samples analyzed using Field GC for TCE, PCE, cis-1,2-DCE, VC, and BTEX.
Location 9A	Groundwater	Only conduct Hydropunch sampling if TCE in groundwater at Location 9 is <100 ug/l
Location 9B	Groundwater	Only conduct Hydropunch sampling if TCE in groundwater at Location 9 is >600 ug/l
Location 10	Groundwater	Collect groundwater sample for Field GC testing for TCE, PCE, cis-1,2-DCE, VC, and BTEX. If TCE is >600 ug/L, sample groundwater via Hydropunch at Location 10B. If TCE is <100 ug/L, Hydropunch sample at Location 10A. Groundwater samples analyzed using Field GC for TCE, PCE, cis-1,2-DCE, VC, and BTEX.
Location 10A	Groundwater	Only conduct Hydropunch sampling if TCE in groundwater at Location 10 is <100 ug/l
Location 10B	Groundwater	Only conduct Hydropunch sampling if TCE in groundwater at Location 10 is >600 ug/l
Location 11	Groundwater	Collect groundwater sample for Field GC testing for TCE, PCE, cis-1,2-DCE, VC, and BTEX. If TCE is <100 ug/L. Location 11 does not have an 11B, because groundwater well MW-3 will be utilized as a step out groundwater sampling location. EPA Comment 10 and 22
Location 11A	Groundwater	Only conduct "step-in" Hydropunch sampling if TCE in groundwater at Location 11 is <100 ug/l.
Location 12 (New MW-11)	Groundwater	Groundwater sampled along with all existing and new monitoring wells and analyzed for RFI Work Plan parameters
Location 13	Step Out Soil Gas	No decision - collect only soil gas sample for field GC testing for TCE, PCE, cis-1,2-DCE, VC, and BTEX.
Location 14	Step Out Soil Gas	No decision - collect only soil gas sample for field GC testing for TCE, PCE, cis-1,2-DCE, VC, and BTEX.
Location 15	Step Out Soil Gas	No decision - collect only soil gas sample for field GC testing for TCE, PCE, cis-1,2-DCE, VC, and BTEX.
Location 16 Sewer Bedding Sample 1	Soil Gas/ Groundwater	Target the granular backfill/bedding of the utilities for temporary soil gas probes and, if possible, groundwater sampling using Hydropunch or soft dig temporary well point. One step out ~200 feet away if VISLs are significantly exceeded.

Table 1 Sampling Plan Decision Tree Exide Technologies Frankfort, Indiana

Sample Location	Type of Sample	Decision
Location 17 Sewer Bedding Sample 2	Soil Gas/ Groundwater	Target the granular backfill/bedding of the utilities for temporary soil gas probes and, if possible, groundwater sampling using Hydropunch or soft dig temporary well point. One step out ~200 feet away if VISLs are significantly exceeded.
Location 18 Sewer Bedding Sample 3	Soil Gas/ Groundwater	Target the granular backfill/bedding of the utilities for temporary soil gas probes and, if possible, groundwater sampling using Hydropunch or soft dig temporary well point. One step out ~200 feet away if VISLs are significantly exceeded.

NOTE: Field GC testing for TCE, PCE, cis-1,2-DCE, VC, and BTEX

Table 2 Summary of Soil Gas Point Installation Observations Exide Technologies Frankfort, Indiana

Location	Soil	Observed	Depth to	PID	Remarks
	Gas	Depth to	bottom of 8-	Reading	
	Sample	Water on	inch screen	(ppm)	
		Installation	(feet)		
		(feet)			
		(loot)			
L1	DSG	3	2	<1.0	
L2	PRT	4.5	3.5	6.4 -	Initially
				18.4	appeared dry to
					5 feet.
L3	DSG	3.5	2.5	1 – 22.8	
L4	PRT	4.5	3.5		
L5	PRT	4.5	3.5	1.0 -5.0	
L6	PRT	3.5	2.5	<1.0	
L13	PRT	>6	5	<1.0	Initially
					appeared dry to
					6 feet
	D00	. 0	4.5	.4.0	
L14	DSG	>6	4.5	<1.0	
L15	DSG	5	4	<1.0	
L16	PRT	>3	3 (top of	<1.0	Sewer Bed
	(SBS)		pipe)		Sample
	(000)				
L17	PRT	>2.5	2.5 (top of	<1.0	Sewer Bed
	(SBS)		pipe)		Sample

Table 2 Summary of Soil Gas Point Installation Observations Exide Technologies Frankfort, Indiana

Location	Soil Gas Sample	Observed Depth to Water on Installation (feet)	Depth to bottom of 8- inch screen (feet)	PID Reading (ppm)	Remarks	
L18	DSG (SBS)	>3	3 (top of pipe)	<1.0	Sewer Bee Sample	d

DSG – Deep Soil Gas collected at depth shown on Column 4 after Helium Leak Test PRT - Soil gas collected by Geoprobe Post Run Tubing (PRT) Method at 1.5 feet depth SBS - Sewer Bed Sample

TABLE 3 SUMMARY OF CALCULATED K VALUES FROM SLUG TEST DATA Exide Environmental Response Trust Frankfort, Indiana

	Calculated K
Well	(ft/day)
MW-1	0.0088
MW-3	0.0078
MW-4	0.0033
MW-7	0.0004
MW-9	0.0015
MW-10	0.0012

Table 4 **Monitoring Well Construction Information Exide Environmental Response Trust** Frankfort, Indiana

			Inner Casing	Outer Casing Diameter		DTW (TOIC)	DTW (TOIC)	DTW (TOIC)	DTW (TOIC)	Ground Surface	TOIC Elevation	Screen Interval	
Well ID	Date Installed	Construction Type	Diameter (in.)	(in.)	(f.b.g.s.)	May 2018	July 2018	November 2019	December 2021	Elevation (ft.)	(ft.)	(ft.)	Comments
MW-1	5/2/2018	Hollow Stem Auger	2	6	18	8.08	7.97	8.15	7.49	849.27	851.26	10 (8-18 ft. bgs)	0.0 PID readings
MW-2	5/2/2018	Hollow Stem Auger	2	6	18	10.24	10.83	10.44	8.83	846.97	848.92	10 (8-18 ft. bgs)	0.0 PID readings
MW-3		Hollow Stem Auger	2	6	16	7.23	7.10	7.13	6.67	849.43	851.45	10 (6-16 ft. bgs)	0.0 PID readings
MW-4		Hollow Stem Auger	2	6	18	4.74	4.33	4.82	4.43	851.19	853.17	10 (8-18 ft. bgs)	1.2 PID reading
MW-5		Hollow Stem Auger	2	6	13	5.28	5.15	5.26	4.48	853.95	856.05	5 (8-13 ft. bgs)	0.0 PID readings
MW-6		Hollow Stem Auger	2	6	18	4.31	4.25	4.39	4.13	853.24	855.47	10 (8-18 ft. bgs)	0.0 PID readings
MW-7	5/1/2018	Hollow Stem Auger	2	6	22	7.16	6.97	5.17	4.61	845.86	847.70	10 (12-22 ft. bgs)	0.0 PID readings
MW-8		Hollow Stem Auger	2	6	18	6.12	6.13	6.18	5.59	851.76	853.73	10 (8-18 ft. bgs)	0.0 PID readings
MW-9		Hollow Stem Auger	2	*	18	NM	NM	4.20	1.89	849.60	849.00	10 (8-18 ft. bgs)	0.0 PID readings
MW-10		Hollow Stem Auger	2	*	18	NM	NM	3.20	2.42	846.00	842.81	10 (8-18 ft. bgs)	0.0 PID readings
MW-11	10/19/2021	Hollow Stem Auger	2	*	15	NM	NM	NM	3.56	850.40	849.90	10 (5-15 ft. bgs)	
MW-12	10/20/2021	Hollow Stem Auger	2	4	15	NM	NM	NM	3.82	852.54	854.51	10 (5-15 ft. bgs)	
MW-13		Hollow Stem Auger	2	4	15	NM	NM	NM	3.73	849.33	851.20	10 (5-15 ft. bgs)	
MW-14	10/21/2021	Hollow Stem Auger	2	4	15	NM	NM	NM	4.48	849.51	851.60	10 (5-15' ft. bgs)	

FBGS- Feet Below Ground Surface

TOIC- Top of Inner Casing NM- Not Measured

NS- Not Surveyed

*MW-9, MW-10, and MW-11 are flush mount wells

TABLE 5a WATER DISPOSAL SAMPLE RESULTS Exide Environmental Response Trust Frankfort, Indiana

	Sample Location	Water-1021				
Name	Lab ID Sample Date					
Parameter	Matrix		Groundwater			
Volatiles		Units	Result	0	RL	
1.1.2.2 Tethohorochane	Volatiles					
1.1.2-Trichlorothane						
1.1-Dichloroethane	1,1,2-Trichloroethane					
1.1-Dichlorochenee	1,1,2-Trichlorotrifluoroethane					
1.2.3-Trichlorobenzene						
1.2-Dibromo-3-chloropropane	1,2,3-Trichlorobenzene					
1.2-Diehloromechane (EDB)	1,2,4-Trichlorobenzene					
1.2-Dichlorochanzen						
1.2-Dichloropropame	1,2-Dichlorobenzene					
1.3-Dichlorobenzene	1,2-Dichloroethane					
1.4-Dichlorobenzene						
2-Hexanone	1,4-Dichlorobenzene	ug/L		U	5	
### A-Methyl-2-pentanone (MIBK) ### Acctone ### Ug/L ###						
Acetone						
Bromochloromethane	Acetone	ug/L	159			
Bromodichloromethane	Benzene Bromochloromethane					
Bromoform Ug/L U S	Bromodichloromethane			U	5	
Carbon disulfide	Bromoform	ug/L			5	
Carbon tetrachloride						
Chlorotethane	Carbon tetrachloride					
Chloroform	Chlorobenzene	ug/L			5	
Chloromethane						
cis-1,2-Dichloroethene ug/L V 5 cis-1,3-Dichloropropene ug/L U 5 Cyclohexane ug/L U 100 Dibromochloromethane ug/L U 5 Dichlorodifluoromethane ug/L U 5 Ethylbenzene ug/L U 5 Isopropylbenzene (Cumene) ug/L U 5 Methylescholexane ug/L U 50 Methyleche Chloride ug/L U 50 Methylene Chloride ug/L U 5 Total ug/L U 5 Total ug/L U 5 Total ug/L U 5 Total ug/L U 5 Trichlorophene ug/L U 5	Chloromethane					
Cyclobexane	cis-1,2-Dichloroethene	ug/L	7.8		5	
Dibromochloromethane						
Ethylbenzene	Dibromochloromethane			_		
Sepropylbenzene (Cumene)	Dichlorodifluoromethane	ug/L				
Methyleyclohexane ug/L U 50 Methyleyclohexane ug/L U 50 Methyler Chloride ug/L U 5 Methyl-tert-butyl ether ug/L U 4 Styrene ug/L U 5 Tetrachloroethene ug/L U 5 Toluene ug/L U 5 Trans-1,3-Dichloropthene ug/L U 5 Trichloroethene ug/L U 5 Trichloropthene ug/L U 5 Trichloropthene ug/L U 5 Trichloropthene ug/L U 5 Vinyl chloride ug/L U 5 Wylene (Total) ug/L U 10 Semivolatiles ug/L U 10 Z2-C-Oxybis (1-chloropropane) ug/L U 10 Z,3-4,6-Trichlorophenol ug/L U 10 Z,4,5-Trichlorophenol ug/L U 10 <td></td> <td></td> <td></td> <td></td> <td></td>						
Methyl-tert-butyl ether ug/L U 5 Methyl-tert-butyl ether ug/L U 4 Styrene ug/L U 5 Tetrachloroethene ug/L U 5 Toluene ug/L U 5 trans-1,3-Dichloropropene ug/L U 5 trans-1,3-Dichloropropene ug/L U 5 Trichlorofthene ug/L U 5 Trichlorofthene ug/L U 5 Trichlorofthene ug/L U 5 Winyl chloride ug/L U 5 Winyl chloride ug/L U 10 Semivolatiles ug/L U 10 Semivolatiles ug/L U 10 Septiciborofiel ug/L U 10 2,2-Pornativaltile ug/L U 10 2,4-Sintrolorophenol ug/L U 10 2,4-Dinintrolulene ug/L U 10	Methyl acetate					
Methyl-tert-butyl ether ug/L U 4 Styrene ug/L U 5 Tetrachloroethene ug/L U 5 Toluene ug/L U 5 Trans-1,2-Dichloroethene ug/L U 5 trans-1,3-Dichloropropene ug/L U 5 Trichlorothuoromethane ug/L U 5 Trichlorothuoromethane ug/L U 5 Vinyl chloride ug/L U 5 Vinyl chloride ug/L U 10 Semivolatiles U 10 2 Sylker (Total) ug/L U 10 Semivolatiles U 10 10 2 2,3-6-Frichlorophenol ug/L U 10 10 2 3-1 2 2 2-Naybis(1-chlorophenol 10 10 2,4-5-Trichlorophenol 10 10 2,4-5-Trichlorophenol 10 10 2,4-5-Trichlorophenol 10 10 2,4-5-Trichlo	Methylcyclohexane					
Styrene						
Toluene	Styrene					
Trans-1,2-Dichloroethene	Tetrachloroethene					
trans-1,3-Dichloropropene ug/L U 5 Trichoroethene ug/L 0.61 J 5 Trichlorofuboromethane ug/L U 5 Vinyl chloride ug/L U 1.9 J 2 Xylene (Total) ug/L U 10 2 Semivolatiles Semivolatiles V U 10 2 2,2-Oxybis (1-chlorophenol ug/L U 10 2 3,3-frichlorophenol ug/L U 10 10 2,4-Dichlorophenol ug/L U 10 10 2,4-Dichlorophenol ug/L U 10 10 2,4-Dinitrotoluene ug/L U 10 10 10 10 2,4-Dinitrotoluene ug/L U 10 10						
Trichlorofluoromethane	trans-1,3-Dichloropropene					
Vinyl chloride ug/L 1.9 J 2 Xylenc (Total) ug/L U 10 Semivolatiles Ug/L U 10 2,2*-Oxybis (1-chlorophenol) ug/L U 10 2,3*4,6*-Tetrachlorophenol ug/L U 10 2,4*5-Trichlorophenol ug/L U 10 2,4*5-Trichlorophenol ug/L U 10 2,4*Dinitrophenol ug/L U 10 2,4*Dinitrobluene ug/L U 10 2,4*Dinitrotoluene ug/L U 10 2,4*Dinitrotoluene ug/L U 10 2,4*Dinitrotoluene ug/L U 10 2,4*Dinitrotoluene ug/L U 10 2,6*Dinitrotoluene ug/L U 10 2,6*Dinitrotoluene ug/L U 10 2,6*Dinitrotoluene ug/L U 10 2,6*Dinitrotoluene ug/L U 10 2,0*Dinitr			0.61			
Xylene (Total)			1.9			
2,2'-Oxybis(1-chloropropane) ug/L U 10 2,3,4,6-Tetrachlorophenol ug/L U 10 2,4,5-Trichlorophenol ug/L U 10 2,4,5-Trichlorophenol ug/L U 10 2,4-Dintrolophenol ug/L U 10 2,4-Dinitrophenol ug/L U 10 2,4-Dinitrophenol ug/L U 10 2,4-Dinitrotoluene ug/L U 10 2,5-Dinitrotoluene ug/L U 10 2,6-Dinitrotoluene ug/L U 10 2-Chloroaphthalene ug/L U 10 2-Chlorophenol ug/L U 10 2-Methylphenol(o-Cresol) ug/L U 10 2-Nitroaniline ug/L U 10 2-Nitroaniline ug/L U 10 2-Nitrophenol ug/L U 10 3-Nitroaniline ug/L U 10 3-Nitroaniline ug/L	Xylene (Total)					
2,3,4,6-Tetrachlorophenol ug/L U 10 2,4,4,5-Trichlorophenol ug/L U 10 2,4,6-Trichlorophenol ug/L U 10 2,4-Dinitrophenol ug/L U 10 2,4-Dinitrophenol ug/L U 10 2,4-Dinitrotoluene ug/L U 10 2,4-Dinitrotoluene ug/L U 10 2,4-Dinitrotoluene ug/L U 10 2,4-Dinitrotoluene ug/L U 10 2,6-Dinitrotoluene ug/L U 10 2,6-Dinitrotoluene ug/L U 10 2,6-Dinitrotoluene ug/L U 10 2-Chlorophenol ug/L U 10 2-Chlorophenol ug/L U 10 2-Methylphenol(o-Cresol) ug/L U 10 2-Nitrophenol ug/L U 10 2-Nitrophenol ug/L U 10 3,3'-Dichlorobenzidine ug/L		σ.			10	
2,4,5-Trichlorophenol ug/L U 10 2,4,6-Trichlorophenol ug/L U 10 2,4-Dichlorophenol ug/L U 10 2,4-Dimethylphenol ug/L U 10 2,4-Dimitrophenol ug/L U 10 2,4-Dimitrotoluene ug/L U 10 2,6-Dimitrotoluene ug/L U 10 2,6-Dimitrotoluene ug/L U 10 2-Chlorophenol ug/L U 10 2-Chlorophenol ug/L U 10 2-Methylphenol(o-Cresol) ug/L U 10 2-Nitroaniline ug/L U 10 2-Nitrophenol ug/L U 10 3,3-Dichlorobenzidine ug/L U 10 3,3-Dichlorobenzidine ug/L U 10 4,6-Dinitro-2-methylphenol ug/L U 10 4,6-Dinitro-2-methylphenol ug/L U 10 4-Chloropanline ug	2,2'-Oxybis(1-chloropropane) 2,3,4,6-Tetrachlorophenol					
2,4-Dichlorophenol ug/L U 10 2,4-Dimitrophenol ug/L U 10 2,4-Dimitrophenol ug/L U 50 2,4-Dimitrotoluene ug/L U 10 2,6-Dimitrotoluene ug/L U 10 2-Chlorophenol ug/L U 10 2-Chlorophenol ug/L U 10 2-Methylphenol(o-Cresol) ug/L U 10 2-Nitrophenol ug/L U 10 2-Nitrophenol ug/L U 10 3&-Methylphenol(m&p Cresol) ug/L U 10 3&-Mitroaniline ug/L U 10 4-S-Methylphenol(m&p Cresol) ug/L U 10 3-Nitroaniline ug/L U 10 4-Bromophenylphenylether ug/L U 10 4-Chloro-3-methylphenol ug/L U 10 4-Chlorophenylphenyl ether ug/L U 10 4-Chlorophenylphenyl ether	2,4,5-Trichlorophenol				10	
2,4-Dimethylphenol ug/L U 10 2,4-Dimitrophenol ug/L U 50 2,4-Dimitrotoluene ug/L U 10 2,6-Dinitrotoluene ug/L U 10 2,6-Dinitrotoluene ug/L U 10 2-Chlorophenol ug/L U 10 2-Mitrophenol(o-Cresol) ug/L U 10 2-Nitroaniline ug/L U 10 2-Nitrophenol ug/L U 10 3,3'-Dichlorobenzidine ug/L U 10 3,3'-Dichlorobenzidine ug/L U 10 3,3'-Dichlorobenzidine ug/L U 10 4-Bromophenylphenol ug/L U 10 4-Bromophenylphenyl ether ug/L U 10 4-Chloro-3-methylphenol ug/L U 10 4-Chlorophenylphenylphenyl ether ug/L U 10 4-Chlorophenylphenylphenyl ether ug/L U 10 4-Nitro						
2,4-Dinitrophenol ug/L U 50 2,4-Dinitrotoluene ug/L U 10 2,4-Dinitrotoluene ug/L U 10 2,6-Dinitrotoluene ug/L U 10 2-Chlorophenol ug/L U 10 2-Chlorophenol ug/L U 10 2-Nitroaniline ug/L U 10 2-Nitrophenol ug/L U 10 3-Rethylphenol(m&p Cresol) ug/L U 10 3-Nitroaniline ug/L U 10 3-Nitroaniline ug/L U 10 4-Ghoritro-2-methylphenol ug/L U 10 4-Chloro-3-methylphenol ug/L U 10 4-Chloro-3-methylphenol ug/L U 10 4-Chloroaniline ug/L U 10 4-Chlorophenylphenyl ether ug/L U 10 4-Nitroaniline ug/L U 10 4-Nitrophenol ug/L <t< td=""><td>2,4-Dienlorophenol 2,4-Dimethylphenol</td><td></td><td></td><td></td><td></td></t<>	2,4-Dienlorophenol 2,4-Dimethylphenol					
2,6-Dinitrotoluene ug/L U 10 2-Chloropaphthalene ug/L U 10 2-Chlorophenol ug/L U 10 2-Methylphenol(o-Cresol) ug/L U 10 2-Nitrophenol ug/L U 10 2-Nitrophenol ug/L U 10 3&-Methylphenol(m&p Cresol) ug/L U 10 3,3'-Dichlorobenzidine ug/L U 10 3-Nitroaniline ug/L U 10 4-Bromophenylphenol ug/L U 10 4-Bromophenylphenyl ether ug/L U 10 4-Chloro-3-methylphenol ug/L U 10 4-Chlorophenylphenyl ether ug/L U 10 4-Chlorophenylphenyl ether ug/L U 10 4-Nitrophenol ug/L U 10 4-Nitrophenol ug/L U 10 Acetophenone ug/L U 10 Aretophenone ug/L </td <td>2,4-Dinitrophenol</td> <td>ug/L</td> <td></td> <td>U</td> <td>50</td>	2,4-Dinitrophenol	ug/L		U	50	
2-Chloronaphthalene						
2-Chlorophenol ug/L U 10 2-Methylphenol(o-Cresol) ug/L U 10 2-Nitronline ug/L U 10 2-Nitrophenol ug/L U 10 3&-Hethylphenol(m&p Cresol) ug/L U 10 3&-Nitroanline ug/L U 20 3-Nitroanline ug/L U 10 4-G-Dinitro-2-methylphenol ug/L U 10 4-Bromophenylphenyl ether ug/L U 10 4-Chloro-3-methylphenol ug/L U 10 4-Chloro-shenylphenyl ether ug/L U 10 4-Chlorophenylphenyl ether ug/L U 10 4-Nitroanline ug/L U 10 4-Nitrophenol ug/L U 10 4-Nitrophenol ug/L U 10 4-Nitrophenol ug/L U 10 Acetophenone ug/L U 10 Benzaldehyde ug/L <	2-Chloronaphthalene		1			
2-Nitroaniline ug/L U 10 2-Nitrophenol ug/L U 10 3-Nitrophenol(m&p Cresol) ug/L U 10 3,3'-Dichlorobenzidine ug/L U 20 3-Nitroaniline ug/L U 10 4-6-Dinitro-2-methylphenol ug/L U 10 4-Bromophenylphenyl ether ug/L U 10 4-Chloro-3-methylphenol ug/L U 10 4-Chlorophenylphenyl ether ug/L U 10 4-Chlorophenylphenyl ether ug/L U 10 4-Nitrophenol ug/L U 10 4-Nitrophenol ug/L U 10 Acetophenone ug/L U 10 Atrazine ug/L U 10 Benzaldehyde ug/L U 10 Biphenyl (Diphenyl) ug/L U 10 bis(2-Chloroethoxy)methane ug/L U 10 bis(2-Ethylhexyl)phthalate	2-Chlorophenol	ug/L		U	10	
2-Nitrophenol ug/L U 10 3&4-Methylphenol(m&p Cresol) ug/L U 10 3&4-Methylphenol(m&p Cresol) ug/L U 20 3-Nitroaniline ug/L U 10 4,6-Dinitro-2-methylphenol ug/L U 10 4-Bromophenylphenyl ether ug/L U 10 4-Chloro-3-methylphenol ug/L U 10 4-Chloro-alline ug/L U 10 4-Chlorophenylphenyl ether ug/L U 10 4-Nitroniline ug/L U 10 4-Nitrophenol ug/L U 10 4-Nitrophenol ug/L U 10 Acetophenone ug/L U 10 Acrazine ug/L U 10 Benzaldehyde ug/L U 10 Benzaldehyde ug/L U 10 Biphenyl (Diphenyl) ug/L U 10 bis(2-Chloroethoxy)methane ug/L						
3&4-Methylphenol(m&p Cresol) ug/L U 10	2-Nitrophenol					
3-Nitroaniline	3&4-Methylphenol(m&p Cresol)	ug/L			10	
4,6-Dinitro-2-methylphenol ug/L U 20 4-Bromophenylphenyl ether ug/L U 10 4-Chloro-3-methylphenol ug/L U 10 4-Chloro-aniline ug/L U 10 4-Chlorophenylphenyl ether ug/L U 10 4-Nitrophienol ug/L U 10 A-Nitrophenol ug/L U 50 Acetophenone ug/L U 10 Atrazine ug/L U 10 Benzaldehyde ug/L U 50 Benzaldehyde ug/L U 10 bis(2-Chloroethoye) ug/L U 10 bis(2-Chloroethoxy)methane ug/L U 10 bis(2-Chloroethyl) ether ug/L U 10 bis(2-Ethylhexyl)phthalate ug/L U 10 Butylbenzylphthalate ug/L U 10 Carbazole ug/L U 10 Dibenzofuram ug/L						
4-Bromophenylphenyl ether ug/L U 10 4-Chloro-3-methylphenol ug/L U 10 4-Chloro-almethylphenol ug/L U 10 4-Chlorophenylphenyl ether ug/L U 10 4-Chlorophenylphenyl ether ug/L U 10 4-Nitroaniline ug/L U 10 4-Nitroaniline ug/L U 10 4-Nitrophenol ug/L U 50 Acetophenone ug/L U 10 Bighenyl (Diphenyl) ug/L U 50 Benzaldehyde ug/L U 50 Benzaldehyde ug/L U 50 bis(2-Chloroethoxy)methane ug/L U 10 bis(2-Chloroethyl) ether ug/L U 10 bis(2-Ethylhexyl)phthalate ug/L U 10 Caprolactam ug/L U 10 Caprolactam ug/L U 10 Carbazole ug/L U 10 Dibenzofuran ug/L U 10	4,6-Dinitro-2-methylphenol					
4-Chloroaniline	4-Bromophenylphenyl ether	ug/L				
4-Chlorophenylphenyl ether ug/L U 10 4-Nitrophienol ug/L U 10 Acetophenone ug/L U 50 Acetophenone ug/L U 10 Atrazine ug/L U 10 Benzaldehyde ug/L U 50 Bisphenyl (Diphenyl) ug/L U 10 bis(2-Chloroethoxy)methane ug/L U 10 bis(2-Ethylnexyl)phthalate ug/L U 10 bis(2-Ethylhexyl)phthalate ug/L U 10 Eaprolactam ug/L U 10 Carbazole ug/L U 10 Dibenzofuran ug/L U 10 Dibenzofuran ug/L U 10 Dimentylphthalate ug/L U 10						
4-Nitrophenol ug/L U 10	4-Chlorophenylphenyl ether	ug/L		U	10	
Acetophenone ug/L U 10 Atrazine ug/L U 10 Benzaldehyde ug/L U 50 Biphenyl (Diphenyl) ug/L U 10 bis(2-Chloroethoxy)methane ug/L U 10 bis(2-Chloroethyl) ether ug/L U 10 Bis(2-Ethylhexyl)phthalate ug/L U 10 Butylbenzylphthalate ug/L U 10 Carbazole ug/L U 10 Carbazole ug/L U 10 Dibenzofuran ug/L U 10 Diibethylphthalate ug/L U 10 Dimethylphthalate ug/L U 10	4-Nitroaniline	ug/L				
Atrazine ug/L U 10 Benzaldehyde ug/L U 50 Beiphenyl (Diphenyl) ug/L U 10 bis(2-Chloroethoxy)methane ug/L U 10 bis(2-Chloroethyl) ether ug/L U 10 bis(2-Ethylhexyl)phthalate ug/L U 10 Butylbenzylphthalate ug/L U 10 Caprolactam ug/L U 10 Carbazole ug/L U 10 Dibenzofuram ug/L U 10 Diethylphthalate ug/L U 10 Dimethylphthalate ug/L U 10			-			
Biphenyl (Diphenyl) ug/L U 10	Atrazine			U	10	
bis(2-Chloroethoxy)methane ug/L U 10 bis(2-Chloroethyl) ether ug/L U 10 bis(2-Ethylhexyl)phthalate ug/L U 10 Butylbenzylphthalate ug/L U 10 Caprolactam ug/L U 10 Carbazole ug/L U 10 Dibenzofuran ug/L U 10 Dibethylphthalate ug/L U 10 Dimethylphthalate ug/L U 10	Benzaldehyde					
bis(2-Chloroethyl) ether ug/L U 10 bis(2-Ethylhexyl)phthalate ug/L U 10 bistylbenzylphthalate ug/L U 10 Caprolactam ug/L U 10 Carbazole ug/L U 10 Dibenzofuran ug/L U 10 Diethylphthalate ug/L U 10 Dimethylphthalate ug/L U 10			-			
Butylbenzylphthalate ug/L U 10 Caprolactam ug/L U 10 Carbazole ug/L U 10 Dibenzofuran ug/L U 10 Diethylphthalate ug/L U 10 Dimethylphthalate ug/L U 10	bis(2-Chloroethyl) ether	ug/L		U	10	
Caprolactam ug/L U 10 Carbazole ug/L U 10 Dibenzofuran ug/L U 10 Diethylphthalate ug/L U 10 Dimethylphthalate ug/L U 10	bis(2-Ethylhexyl)phthalate					
Carbazole ug/L U 10 Dibenzofuran ug/L U 10 Diethylphthalate ug/L U 10 Dimethylphthalate ug/L U 10			1			
Diethylphthalate ug/L U 10 Dimethylphthalate ug/L U 10	Carbazole			U	10	
Dimethylphthalate ug/L U 10	Dibenzofuran					
	Di-n-butylphthalate					

TABLE 5a WATER DISPOSAL SAMPLE RESULTS Exide Environmental Response Trust Frankfort, Indiana

Sample Location			Water-1021	1
Lab ID Sample Date		3	10/22/2021	1
Matrix		(Groundwate	r
Remarks				
Parameter	Units	Result	Q	RL
Di-n-octylphthalate	ug/L		U	10
Hexachloro-1,3-butadiene Hexachlorobenzene	ug/L ug/L		U	10 10
Hexachlorocyclopentadiene	ug/L ug/L		U	10
Hexachloroethane	ug/L		U	10
Isophorone	ug/L		U	10
Nitrobenzene	ug/L		U	10
N-Nitroso-di-n-propylamine	ug/L		U	50
N-Nitrosodiphenylamine Pentachlorophenol	ug/L ug/L		U	10 50
Phenol	ug/L	5.9	J	10
Semivolatiles SIMs	-8-			
2-Methylnaphthalene	ug/L		U	1
Acenaphthene	ug/L		U	1
Acenaphthylene	ug/L		U	1
Anthracene Ponza(a)anthracena	ug/L		U	0.1
Benzo(a)anthracene Benzo(a)pyrene	ug/L ug/L		U	0.1
Benzo(b)fluoranthene	ug/L ug/L		U	0.1
Benzo(g,h,i)perylene	ug/L		U	0.1
Benzo(k)fluoranthene	ug/L		U	0.1
Chrysene	ug/L		U	0.5
Dibenz(a,h)anthracene Fluoranthene	ug/L		U	0.1
Fluoranthene Fluorene	ug/L ug/L		U	1
Indeno(1,2,3-cd)pyrene	ug/L ug/L		U	0.1
Naphthalene	ug/L		U	1
Phenanthrene	ug/L		U	1
Pyrene	ug/L		U	1
Total Metals	-			
Arsenic	ug/L	3.7	J	10
Barium Cadmium	ug/L ug/L	74.1	U	10
Chromium	ug/L ug/L	28.8	U	10
Lead	ug/L	20.0	U	10
Mercury	ug/L		U	2
Selenium	ug/L		U	10
Silver	ug/L		U	10
Volatiles TCLP 1,1-Dichloroethene	ma/I		U	0.05
1,2-Dichloroethane	mg/L mg/L		U	0.05
2-Butanone (MEK)	mg/L		U	1
Benzene	mg/L		U	0.05
Carbon tetrachloride	mg/L		U	0.05
Chlorobenzene	mg/L		U	0.05
Chloroform	mg/L		U	0.05
Tetrachloroethene Trichloroethene	mg/L		U	0.05
Vinyl chloride	mg/L mg/L		U	0.03
Semivolatiles TCLP	mg/L			0.02
1,4-Dichlorobenzene	mg/L		U	0.1
2,4,5-Trichlorophenol	mg/L		U	0.5
2,4,6-Trichlorophenol	mg/L		U	0.1
2,4-Dinitrotoluene	mg/L		U	0.1
2-Methylphenol(o-Cresol) 3&4-Methylphenol(m&n Cresol)	mg/L mg/L		U	0.1
3&4-Methylphenol(m&p Cresol) Hexachloro-1,3-butadiene	mg/L mg/L		U	0.2
Hexachlorobenzene	mg/L		U	0.1
Hexachloroethane	mg/L		U	0.1
Nitrobenzene	mg/L		U	0.1
Pentachlorophenol	mg/L		U	0.5
Pyridine	mg/L		U	0.1
TCLP Metals Arsenic	ma/I		U	0.1
Barium	mg/L mg/L		U	5
Cadmium	mg/L		U	0.05
Chromium	mg/L		U	0.1
Lead	mg/L		U	0.1
Mercury	mg/L		U	0.002
Selenium	mg/L		U	0.1
Silver Conventionals	mg/L		U	0.1
Cyanide, Reactive	mg/kg		U	1
Flashpoint	deg F	>200		<u> </u>
pH at 25 Degrees C	Std. Units	8.7		0.1
Sulfide, Reactive	mg/kg		U	10

J - Denotes an estimated reporting limit U - Analyte was not detected at or above the method detection limit ug/L - micrograms per liter mg/L - milligrams per liter mg/kg - milligrams per kilogram Q - Qualifier RL - Reporting Limit

TABLE 5b SOIL DISPOSAL SAMPLE RESULTS Exide Environmental Response Trust Frankfort, Indiana

Sample Location			Soil-1021	12
Lab ID Sample Date			5030088800 10/22/2021	
Matrix Remarks			Soil	
Parameter	Units	Result	Q	RL
Volatiles 1,1,1-Trichloroethane	mg/kg		U	0.0048
1,1,2,2-Tetrachloroethane	mg/kg		U	0.0048
1,1,2-Trichloroethane 1,1,2-Trichlorotrifluoroethane	mg/kg mg/kg		U U	0.0048 0.0048
1,1-Dichloroethane	mg/kg		U	0.0048
1,1-Dichloroethene 1,2,3-Trichlorobenzene	mg/kg mg/kg		U	0.0048 0.0048
1,2,4-Trichlorobenzene	mg/kg		U	0.0048
1,2-Dibromo-3-chloropropane 1,2-Dibromoethane (EDB)	mg/kg mg/kg		U	0.0096
1,2-Dichlorobenzene	mg/kg		U	0.0048
1,2-Dichloroethane 1,2-Dichloropropane	mg/kg mg/kg		U	0.0048
1,3-Dichlorobenzene	mg/kg		U	0.0048
1,4-Dichlorobenzene 1,4-Dioxane (p-Dioxane)	mg/kg mg/kg		U	0.0048
2-Butanone (MEK)	mg/kg		U	0.024
2-Hexanone	mg/kg		U	0.096
4-Methyl-2-pentanone (MIBK) Acetone	mg/kg mg/kg	0.01	J	0.024
Benzene	mg/kg		U	0.0048
Bromochloromethane Bromodichloromethane	mg/kg mg/kg		U	0.0048
Bromoform	mg/kg		U	0.0048
Bromomethane Carbon disulfide	mg/kg mg/kg		U	0.0048
Carbon tetrachloride	mg/kg		U	0.0048
Chlorobenzene	mg/kg		U	0.0048
Chloroethane Chloroform	mg/kg mg/kg	0.00096	J	0.0048
Chloromethane	mg/kg		U	0.0048
cis-1,2-Dichloroethene cis-1,3-Dichloropropene	mg/kg mg/kg	0.0037	J U	0.0048
Cyclohexane	mg/kg		U	0.096
Dibromochloromethane Dichlorodifluoromethane	mg/kg mg/kg		U	0.0048
Ethylbenzene	mg/kg		U	0.0048
Isopropylbenzene (Cumene)	mg/kg		U	0.0048
Methyl acetate Methylcyclohexane	mg/kg mg/kg		U U	0.0048 0.0048
Methylene Chloride	mg/kg		U	0.019
Methyl-tert-butyl ether Styrene	mg/kg mg/kg		U	0.0048 0.0048
Tetrachloroethene	mg/kg		U	0.0048
Toluene trans-1,2-Dichloroethene	mg/kg mg/kg		U	0.0048
trans-1,3-Dichloropropene	mg/kg		U	0.0048
Trichloroethene Trichlorofluoromethane	mg/kg mg/kg	0.00092	J U	0.0048
Vinyl chloride	mg/kg	0.00064	J	0.0048
Xylene (Total) Total Semivolatiles	mg/kg		U	0.0096
1,2,4,5-Tetrachlorobenzene	mg/kg		U	0.42
2,2'-Oxybis(1-chloropropane)	mg/kg		U	0.42
2,3,4,6-Tetrachlorophenol 2,4,5-Trichlorophenol	mg/kg mg/kg		U U	0.42
2,4,6-Trichlorophenol	mg/kg		U	0.42
2,4-Dichlorophenol 2,4-Dimethylphenol	mg/kg mg/kg		U	0.42
2,4-Dinitrophenol	mg/kg		U	2.1
2,4-Dinitrotoluene 2,6-Dinitrotoluene	mg/kg mg/kg		U	0.42
2-Chloronaphthalene	mg/kg		U	0.42
2-Chlorophenol	mg/kg		U	0.42
2-Methylnaphthalene 2-Methylphenol(o-Cresol)	mg/kg mg/kg		U	0.42 0.42
2-Nitroaniline	mg/kg		U	0.42
2-Nitrophenol 3&4-Methylphenol(m&p Cresol)	mg/kg mg/kg		U	0.42 0.85
3,3'-Dichlorobenzidine	mg/kg		U	0.85
3-Nitroaniline 4,6-Dinitro-2-methylphenol	mg/kg mg/kg		U	0.42 0.85
4-Bromophenylphenyl ether	mg/kg		U	0.42
4-Chloro-3-methylphenol 4-Chloroaniline	mg/kg		U U	0.85 0.85
4-Chlorophenylphenyl ether	mg/kg mg/kg		U	0.83
4-Nitroaniline	mg/kg		U	0.42
4-Nitrophenol Acenaphthene	mg/kg mg/kg		U U	2.1 0.42
Acenaphthylene	mg/kg		U	0.42
Acetophenone	mg/kg	<u> </u>	U	0.42

TABLE 5b SOIL DISPOSAL SAMPLE RESULTS Exide Environmental Response Trust Frankfort, Indiana

Lab ID	Sample Location			Soil-1021	
Matrix					
Remarks					
Parameter				3011	
Arrazine		Units	Result	Q	RL
Benzalehyde					
Benzo(a)anthracene					
Benzo(a)pyrene					
Benzo(ghi)proranthene					
Benzotk/fluoranthene	Benzo(b)fluoranthene				
Bipheny (Dipheny)					
bis(2-Chloroethoxy)methane mg/kg U 0.42 bis(2-Chloroethyl) ether mg/kg U 0.42 bis(2-Chloroethyl) ether mg/kg U 0.42 Butylbenzylphthalate mg/kg U 0.42 Butylbenzylphthalate mg/kg U 0.42 Carbazole mg/kg U 0.42 Chrysene mg/kg U 0.42 Dibenz(a,h)anthracene mg/kg U 0.42 Dienozofuran mg/kg U 0.42 Biscophilatalate mg/kg U 0.42 Birocorne mg/kg U 0.42 Hewachloro-1,3-butadiene mg/kg U 0.42 Hexachloro-benzene <					
bis(2-Ethylnexyl)phthalate mg/kg U 0.42 Butylbenrylphthalate mg/kg U 0.42 Carpotactam mg/kg U 0.42 Carbazole mg/kg U 0.42 Chrysene mg/kg U 0.42 Dibenz(a,h)anthracene mg/kg U 0.42 Dibenzofuran mg/kg U 0.42 Dibenzofuran mg/kg U 0.42 Dibenzhylphthalate mg/kg U 0.42 Dirn-butylphthalate mg/kg U 0.42 Pin-roetylphthalate mg/kg U 0.42 Fluorene mg/kg U 0.42 Fluorene mg/kg U 0.42 Hexachloroely-loghedidiene mg/kg U 0.42 Hexachloroely-loghedidiene mg/kg U 0.42 Hexachloroely-loghedidiene mg/kg U 0.42 Hexachloroely-loghedidiene mg/kg U 0.42 Hexachloroe	bis(2-Chloroethoxy)methane				
Butylbenzylphthalate				U	
Caprolactam					
Carbazole					
Chrysene mg/kg U 0.42 Dibenzofuran mg/kg U 0.42 Dibenzofuran mg/kg U 0.42 Dibenzofuran mg/kg U 0.42 Dimethylphthalate mg/kg U 0.42 Din-n-butylphthalate mg/kg U 0.42 Di-n-butylphthalate mg/kg U 0.42 Fluoranthene mg/kg U 0.42 Fluorene mg/kg U 0.42 Hexachloro-1,3-butadiene mg/kg U 0.42 Hexachloroe-Lyclopentadiene mg/kg U 0.42 Hexachloroethane mg/kg U 0.42 Idexachloroethane mg/kg U 0.42 Idexachloroethane mg/kg U 0.42 Idexachloroethane mg/kg U 0.42 Idexachloroethane mg/kg U 0.42 Naphthalene mg/kg U 0.42 Naphthalene mg/kg					
Dibenzofuran					
Diethylphthalate	Dibenz(a,h)anthracene				0.42
Dimethylphthalate					
Di-n-butylphthalate					
Di-n-octylphthalate					
Fluorene					
Hexachloro-1,3-butadiene	Fluoranthene	mg/kg		U	0.42
Hexachloroeyclopentatiene					
Hexachlorocyclopentadiene					
Hexachloroethane					
Isophorone					
Naphthalene		mg/kg			
N-Nitroso-din-propylamine mg/kg	Isophorone				
N-Nitroso-di-n-propylamine mg/kg					
N-Nitrosodiphenylamine					
Phenanthrene					
Phenol mg/kg					
Pyrene					
Protal Metals					
Barium		mg/kg		Ü	0.12
Cadmium					
Chromium					
Lead mg/kg 47.7 1.2					
Mercury					
Silver				J	
The color of the					
1,1-Dichloroethene		mg/kg		U	0.6
1,2-Dichloroethane		mø/L	I	II	0.05
2-Butanone (MEK) mg/L U 1 Benzene mg/L U 0.05 Carbon tetrachloride mg/L U 0.05 Chlorobenzene mg/L U 0.05 Chloroform mg/L U 0.05 Tetrachloroethene mg/L U 0.05 Trichloroethene mg/L U 0.05 Vinyl chloride mg/L U 0.02 TCLP Semivolatiles 1,4-Dichlorobenzene mg/L U 0.02 TCLP Semivolatiles 1,4-Dichlorobenzene mg/L U 0.1 2,4,5-Trichlorophenol mg/L U 0.1 2,4,5-Trichlorophenol mg/L U 0.1 2,4-Dinitrotoluene mg/L U 0.1 2,4-Dinitrotoluene mg/L U 0.1 2-Methylphenol(m&p Cresol) mg/L U 0.1 3&4-Methylphenol(m&p Cresol) mg/L U 0.1 Hexachloro-1,3-bu	7				
Carbon tetrachloride					
Chlorobenzene mg/L U 0.05 Chloroform mg/L U 0.05 Tetrachloroethene mg/L U 0.05 Trichloroethene mg/L U 0.05 Vinyl chloride mg/L U 0.05 Vinyl chloride mg/L U 0.02 TCLP Semivolatiles 1,4-Dichlorobenzene mg/L U 0.1 2,4,5-Trichlorophenol mg/L U 0.5 2,4,6-Trichlorophenol mg/L U 0.1 2,4-Dinitrotoluene mg/L U 0.1 2,4-Binitrotoluene mg/L U 0.1 2,4-Binitrotoluene mg/L U 0.1 2,4-Binitrotoluene mg/L U 0.1					
Chloroform mg/L U 0.05 Tetrachloroethene mg/L U 0.05 Trichloroethene mg/L U 0.05 Vinyl chloride mg/L U 0.02 TCLP Semivolatiles 1,4-Dichlorobenzene mg/L U 0.1 2,4,5-Trichlorophenol mg/L U 0.5 2,4,5-Trichlorophenol mg/L U 0.1 2,4-Fonitirotoluene mg/L U 0.1 2,4-Dinitirotoluene mg/L U 0.1 3&4-Methylphenol(o-Cresol) mg/L U 0.1 Hexachlorolomene mg/L U 0.1 Hexachloroethane mg/L U 0.1 Nitrobenzene mg/L U 0.1 <td></td> <td></td> <td></td> <td></td> <td></td>					
Tetrachloroethene mg/L U 0.05 Trichloroethene mg/L U 0.05 Vinyl chloride mg/L U 0.02 TCLP Semivolatiles 1,4-Dichlorobenzene mg/L U 0.1 2,4,5-Trichlorophenol mg/L U 0.5 2,4,5-Trichlorophenol mg/L U 0.1 2,4-Dinitrotoluene mg/L U 0.1 3&4-Methylphenol(o-Cresol) mg/L U 0.1 Hexachlorolo(m&p Cresol) mg/L U 0.1 Hexachloroetnee mg/L U 0.1 Hexachloroetnee mg/L U 0.1 Pentachlorophenol mg/L U <					
Vinyl chloride					
TCLP Semivolatiles		mg/L			
1,4-Dichlorobenzene mg/L U 0.1 2,4,5-Trichlorophenol mg/L U 0.5 2,4,6-Trichlorophenol mg/L U 0.1 2,4-Pointrotoluene mg/L U 0.1 2-Methylphenol(o-Cresol) mg/L U 0.1 3&4-Methylphenol(m&p Cresol) mg/L U 0.2 Hexachloro-1,3-butadiene mg/L U 0.1 Hexachlorobenzene mg/L U 0.1 Hexachloroethane mg/L U 0.1 Nitrobenzene mg/L U 0.1 Pentachlorophenol mg/L U 0.5 Pyridine mg/L U 0.1 TCLP Metals T T U 0.1 Arsenic mg/L U 0.1 Barium mg/L U 0.05 Cadmium mg/L U 0.0 Chromium mg/L U 0.1 Lead mg/L U		mg/L		U	0.02
2,4,5-Trichlorophenol mg/L U 0.5 2,4,6-Trichlorophenol mg/L U 0.1 2,4-Dinitrotoluene mg/L U 0.1 2,4-Dinitrotoluene mg/L U 0.1 2,4-Dinitrotoluene mg/L U 0.1 3&4-Methylphenol(o-Cresol) mg/L U 0.2 Hexachlorol,3-butadiene mg/L U 0.1 Hexachlorobenzene mg/L U 0.1 Hexachlorobenzene mg/L U 0.1 Nitrobenzene mg/L U 0.1 Pentachlorophenol mg/L U 0.5 Pyridine mg/L U 0.1 TCLP Metals T U 0.1 Arsenic mg/L U 0.1 Barium mg/L U 0.05 Cadmium mg/L U 0.05 Chromium mg/L U 0.1 Lead mg/L U 0.00		ma/I		II	0.1
2,4,6-Trichlorophenol mg/L U 0.1 2,4-Dinitrotoluene mg/L U 0.1 2-Methylphenol(o-Cresol) mg/L U 0.1 3&4-Methylphenol(m&p Cresol) mg/L U 0.2 Hexachloro-1,3-butadiene mg/L U 0.1 Hexachlorobenzene mg/L U 0.1 Hexachlorotethane mg/L U 0.1 Nitrobenzene mg/L U 0.1 Pentachlorophenol mg/L U 0.5 Pyridine mg/L U 0.1 TCLP Metals T U 0.1 Arsenic mg/L U 0.1 Barium mg/L U 0.05 Cadmium mg/L U 0.05 Chromium mg/L U 0.1 Lead mg/L U 0.0 Mercury mg/L U 0.00 Selenium mg/L U 0.1					
2-Methylphenol(o-Cresol) mg/L U 0.1 3&4-Methylphenol(m&p Cresol) mg/L U 0.2 Hexachloro-1,3-butadiene mg/L U 0.1 Hexachlorobenzene mg/L U 0.1 Hexachloroethane mg/L U 0.1 Nitrobenzene mg/L U 0.1 Pentachlorophenol mg/L U 0.5 Pyridine mg/L U 0.1 TCLP Metals Arsenic mg/L U 0.1 Barium mg/L U 0.0 Cadmium mg/L U 0.05 Chromium mg/L U 0.1 Lead mg/L U 0.1 Mercury mg/L U 0.002 Selenium mg/L U 0.1					
3&4-Methylphenol(m&p Cresol) mg/L U 0.2 Hexachloro-1,3-butadiene mg/L U 0.1 Hexachlorobenzene mg/L U 0.1 Hexachlorothane mg/L U 0.1 Nitrobenzene mg/L U 0.1 Pentachlorophenol mg/L U 0.5 Pyridine mg/L U 0.1 TCLP Metals Arsenic mg/L U 0.1 Barium mg/L U 0.05 Cadmium mg/L U 0.05 Chromium mg/L U 0.1 Lead mg/L U 0.1 Mercury mg/L U 0.002 Selenium mg/L U 0.1	2,4-Dinitrotoluene	mg/L			
Hexachloro-1,3-butadiene mg/L U 0.1 Hexachlorobenzene mg/L U 0.1 Hexachlorobenzene mg/L U 0.1 Hexachloropthane mg/L U 0.1 Hexachloropthane mg/L U 0.1 Pentachlorophenol mg/L U 0.5 Pyridine mg/L U 0.1 Pentachlorophenol mg/L U 0.1 Pentachlorophenol mg/L U 0.1 Barium mg/L U 0.1 Barium mg/L U 0.05 Cadmium mg/L U 0.05 Chromium mg/L U 0.1 Lead mg/L U 0.1 Mercury mg/L U 0.002 Selenium mg/L U 0.01					
Hexachlorobenzene mg/L					
Hexachloroethane					
Nitrobenzene mg/L U 0.1 Pentachlorophenol mg/L U 0.5 Pyridine mg/L U 0.1 TCLP Metals Arsenic mg/L U 0.1 Barium mg/L U 0.5 Cadmium mg/L U 0.05 Chromium mg/L U 0.1 Lead mg/L U 0.002 Selenium mg/L U 0.002 Selenium mg/L U 0.1	Hexachloroethane			U	0.1
Pyridine mg/L U 0.1 TCLP Metals Arsenic mg/L U 0.1 Barium mg/L 0.82 J 5 Cadmium mg/L U 0.05 Chromium mg/L U 0.1 Lead mg/L U 0.1 Mercury mg/L U 0.002 Selenium mg/L U 0.1		mg/L			
TCLP Metals Arsenic mg/L U 0.1 Barium mg/L 0.82 J 5 Cadmium mg/L U 0.05 Chromium mg/L U 0.1 Lead mg/L U 0.1 Mercury mg/L U 0.002 Selenium mg/L U 0.1					
Arsenic mg/L U 0.1 Barium mg/L 0.82 J 5 Cadmium mg/L U 0.05 Chromium mg/L U 0.1 Lead mg/L U 0.1 Mercury mg/L U 0.002 Selenium mg/L U 0.1		mg/L		U	0.1
Barium mg/L 0.82 J 5 Cadmium mg/L U 0.05 Chromium mg/L U 0.1 Lead mg/L U 0.1 Mercury mg/L U 0.002 Selenium mg/L U 0.1		mg/L		U	0.1
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	Barium	mg/L	0.82		5
Mercury mg/L U 0.002 Selenium mg/L U 0.1					
Selenium mg/L U 0.1					
				U	
				U	

TABLE 5b SOIL DISPOSAL SAMPLE RESULTS Exide Environmental Response Trust Frankfort, Indiana

Sample Location			Soil-1021	
Lab ID			5030088800)2
Sample Date			10/22/2021	
Matrix			Soil	
Remarks				
Parameter	Units	Result	Q	RL
Conventionals				
Cyanide, Reactive	mg/kg		U	1.3
Ignitability, non-metallic	mm/sec	<2.2	U	2.2
Percent Moisture	%	22.2		0.1
pH at 25 Degrees C	Std. Units	6.8		0.1
Sulfide, Reactive	mg/kg		U	12.9

- J Denotes an estimated reporting limit
 U Analyte was not detected at or above the method detection limit
 mg/kg milligrams per kilogram
 mm/sec millimeters per second
 Q Qualifier
 RL Reporting Limit

TABLE 6 GROUNDWATER MONITORING WELL SAMPLE RESULTS

Exide Environmental Response Trust Frankfort, Indiana

G 1 7 2				X7 1				1 1	XX 2		633 7				t, Indiai		0) (IVI OF		3.637.10	3.037.11		TT 10		7.10	3.6337	1.1
Sample Location				W-1	0.6	MV			W-3		ИW-			W-7		MW		MW-9D		MW-10	MW-11		W-12		V-13	MW	
Lab ID	2021		503049			503049			1949003			9014		194900			19009	503049490		50304949002	50304949008		4949007	503049		503049	
Sample Date	IDEM		12/8/	/2021		12/8/	2021	12/8	3/2021	12	/9/20)21	12/8	3/2021	1	12/9/2	021	12/9/202	1	12/8/2021	12/9/2021	12/	9/2021	12/9/	2021	12/9/2	2021
Matrix	RCG GW		Groun	dwat	er	Ground	dwater	Grou	ndwater	Gro	undv	vater	Groun	ndwate	r Gı	round	water	Groundwa	ter	Groundwater	Groundwater	Grou	ındwater	Groun	dwater	Ground	lwater
Remarks	Tap Limit																	FD of MW	V-9								
Parameter		Units	Result	0	RL	Result	O R	L Result	O RL	Result	0	RL	Result	OF	RL Resu	ult C) RL	Result Q	RL	Result Q RI	Result Q R	L Resul	t Q RL	Result	Q RL	Result	O RL
Volatiles		0 111111		~			V		¥		1 ×												. &		¥		V
1,1,1-Trichloroethane	200	na/I	0.34	т	5		U :	: 1	U 5	1	II	2500		U	5	I	J 25	U	25	III 5	l lul:		UJ 5		11 5		UI 5
		ug/L	0.34	U	-					-	U				<i>5</i>	I		U	25	U 5					U 3		0 0
1,1,2,2-Tetrachloroethane	0.76	ug/L			5			5			U	2500			5							5			U 5		
1,1,2-Trichloroethane	5	ug/L		U	5			5	U 5		U	2500			5	Ţ		U	25	U 5		5	UJ 5		U 5		U 5
1,1,2-Trichlorotrifluoroethane	10000	ug/L		U	5			5	U 5		U	2500			5	Į		U	25	U 5	U :		UJ 5		U 5		U 5
1,1-Dichloroethane	28	ug/L	1.3	J	5		U :	5	U 5	795	J	2500		U	5 10.	8 J	25	11.1 J	25	U 5	U :	5 0.68	J 5	2.3	J 5		U 5
1,1-Dichloroethene	7	ug/L		U	5		U :	5	U 5	554	J	2500		U	5 7.4	4 J	25	7.7 J	25	U 5	U :	5	UJ 5		U 5		U 5
1,2,3-Trichlorobenzene	7	ug/L		U	5		U :	5	U 5		U	2500		U	5	J	J 25	U	25	U 5	U :	5 5	UJ 5		U 5		U 5
1,2,4-Trichlorobenzene	70	ug/L		U	5			5	U 5		U	2500		U	5	Į		U	25	U 5	U :	5 3.6	J 5		U 5		U 5
1,2-Dibromo-3-chloropropane	0.2	ug/L			10		U 1		U 10		IJ	5000			10	I		U	50	U 10			UJ 10		U 10		U 10
1,2-Dibromoethane (EDB)	0.05	ug/L		U	5			5	U 5		II	2500			5	ī		U	25	U 5	U :		UJ 5		U 5		U 5
1.2-Dichlorobenzene	600	ug/L ug/L		U	5		U :		U 5		II	2500	 		5	I		U	25	U 5		5 0.21	J 5	 	U 5		U 5
1,2-Dichloroethane	5	ì		U	5			5	U 5	1	II	2500			5	Ţ		U	25	U 5		5 0.21	UJ 5		U 5		U 5
		ug/L		_	-					1	U		1		-									1			
1,2-Dichloropropane	5	ug/L		U	5			5	U 5	1	U	2500	1		5	Į		U	25	U 5		5	UJ 5	1	0 3		
1,3-Dichlorobenzene	NC	ug/L		U	5		U :		U 5		U	2500			5	U		U	25	U 5		5	UJ 5		U 5		U 5
1,4-Dichlorobenzene	75	ug/L		U	5			5	U 5		U	2500			5	Ţ		U	25	U 5		5	UJ 5		U 5		U 5
2-Butanone (MEK)	5600	ug/L			25		U 2		U 25		U	12500		U 2		Ţ	_	U	125	U 25	U 2		UJ 25		U 25		U 25
2-Hexanone	38	ug/L			25		U 2		U 25		U	12500			25	U	125	U	125	U 25	U 2	.5	UJ 25		U 25		U 25
4-Methyl-2-pentanone (MIBK)	6300	ug/L		U	25		U 2	5	U 25		U	12500		U 2	25	J	125	U	125	U 25	U 2	.5	UJ 25		U 25		U 25
Acetone	14000	ug/L		U	100		UJ 10	00	UJ 100)	U	50000	12.9	J 1	00	Į	500	U	500	UJ 100	U 10	00	UJ 100		U 100		U 100
Benzene	5	ug/L		U	5		U :	5	U 5		U	2500		U	5 3.2	2 J	25	3 J	25	U 5	U :	5	UJ 5		U 5		U 5
Bromochloromethane	83	ug/L		U	5		U :	5	U 5		U	2500		U	5	I	J 25	U	25	U 5	U :	5	UJ 5		U 5		U 5
Bromodichloromethane	80	ug/L		U	5			5	U 5		II	2500			5	ī		U	25	U 5		5	UJ 5		U 5		U 5
Bromoform	80	ug/L		U	5			5	U 5		II	2500			5	ī		U	25	U 5		5	UJ 5		U 5		U 5
Bromomethane	7.5	ug/L		U	5			5	U 5		II	2500			5	ī		U	25	U 5	U		UJ 5		U 5		U 5
Carbon disulfide	810	ug/L			10			0	U 10		TII	5000			0	U		UJ	50	U 10			UJ 10		UJ 10		UJ 10
)		U					U 5	-	II					I				U 5							U 5
Carbon tetrachloride	5	ug/L			5			5			U	2500			5			U	25			5			0		
Chlorobenzene	100	ug/L		U	5			5	U 5		U	2500			5	Ţ		U	25	U 5		5	UJ 5		U 5		U 5
Chloroethane	8300	ug/L		U	5		U :		U 5		U	2500			5	J		U	25	U 5		5	UJ 5		U 5		U 5
Chloroform	80	ug/L		UJ	5			5	UJ 5	149	J	2500			5 25			25 U	25	UJ 5	UJ :		UJ 5		UJ 5		UJ 5
Chloromethane	190	ug/L		U	5			5	U 5		U	2500			5	U		U	25	U 5		5	UJ 5		U 5		U 5
cis-1,2-Dichloroethene	70	ug/L	9.8		5		U :	5	U 5	267000		25000	0.72	J	5 321	0	250	3290	250	U 5	U :	5 4.7	J 5	31.2	5	1.2	J 5
cis-1,3-Dichloropropene	4.7	ug/L		U	5		U :	5	U 5		U	2500		U	5	J	J 25	U	25	U 5	U	5	UJ 5		U 5		U 5
Cyclohexane	13000	ug/L		UJ	100		U 10	00	U 100)	UJ	50000		U 1	00	U	J 500	UJ	500	U 100	UJ 10	00	UJ 100		UJ 100		UJ 100
Dibromochloromethane	80	ug/L		U	5		U :	5	U 5		U	2500		U	5	Į	J 25	U	25	U 5	U :	5	UJ 5		U 5		U 5
Dichlorodifluoromethane	200	ug/L		U	5		U :		U 5		U	2500		U	5	Į		U	25	U 5	U :	5	UJ 5		U 5		U 5
Ethylbenzene	700	ug/L		U	5			5	U 5		II	2500	1		5	ī		U	25	U 5	U		UJ 5	1	U 5		U 5
Isopropylbenzene (Cumene)	450	ug/L		U	5			5	U 5		II	2500			5	ī		U	25	U 5		5	UJ 5		U 5		U 5
Methyl acetate	20000	ug/L ug/L		U	50		U 5		U 50		II	25000	1		50	I		III	250	U 50	U 5		UJ 50	1	U 50		U 50
Methylcyclohexane	NC	ug/L ug/L			50		U 5	-	U 50	1	TIT	25000	1		50	U		UJ	250	U 50			UJ 50	 	UJ 50		UJ 50
_ , ,		ì						-		1	UJ		-			I											
Methylene Chloride	5	ug/L		U	5			5		1	U	2500	1		5			U	25			5	UJ 5	1	U 5		
Methyl-tert-butyl ether	140	ug/L		U	4			1	U 4		U	2000			4	Į		U	20	U 4		4	UJ 4		U 4		U 4
Styrene	100	ug/L		U	5		U :		U 5		U	2500			5	U		U	25	U 5		5	UJ 5		U 5		U 5
Tetrachloroethene	5	ug/L		U	5			5	U 5		U	2500			5	Į		U	25	U 5	U :		UJ 5		U 5		U 5
Toluene	1000	ug/L		U	5			5	U 5	239	J	2500	<u> </u>		5	Ţ		U	25	U 5		5	UJ 5	<u> </u>	U 5		U 5
trans-1,2-Dichloroethene	100	ug/L	0.39	J	5			5	U 5	1700	J	2500			5 53	3	25	53.6	25	U 5		5 0.27	J 5	0.31	J 5		U 5
trans-1,3-Dichloropropene	4.7	ug/L		UJ	5		UJ :	5	UJ 5		UJ	2500		UJ	5	U	J 25	UJ	25	UJ 5	UJ :	5	UJ 5		UJ 5		UJ 5
Trichloroethene	5	ug/L	49.1		5			5	U 5	187000		25000		U	5 1.8	3 J	_	1.9 J	25	U 5	U :	5 0.46	J 5	0.43	J 5		U 5
Trichlorofluoromethane	5200	ug/L		U	5		U :		U 5		U	2500			5	Į		U	25	U 5		5	UJ 5		U 5		U 5
Vinyl chloride	2	ug/L		U	2		U 2		U 2	22900	Ť	1000	1		2 95		10	932	10	U 2	U		J 2	29.1	2		U 2
Xylene (Total)	10000	ug/L		U	10		U 1		U 10		11	5000	l		10	·		U	50	U 10			UJ 10	1	U 10		U 10
zigione (10mi)	10000	ug/L		U	10		0 1	V	0 10		U	5000	l	0 1		·	, 50	10	50	0 10	U	V	03 10		5 10		0 10

J - Denotes an estimated reporting limit ug/L - microgram per liter

 \mbox{U} - Analyte was not detected at or above the method detection limit RCG - Remediation Closure Guide

NA - Not Analyzed for this parameter

TABLE 6 GROUNDWATER MONITORING WELL SAMPLE RESULTS Exide Environmental Response Trust Frankfort, Indiana

Sample Location			EB-	01-120	0821	EB-01	-120	0821	TB-01-	110	919
Lab ID				04949		50304			50304	-	
Sample Date	2021			2/8/202		12/8			11/9		
Matrix	IDEM			queou		Aqu			Aqu		
Remarks	RCG GW Tap Limit			ment		Equipm			Trip		
Parameter	тар Еши	Units	Result		RL				Result		
Volatiles	l .	Cints	Result	V	KL	Result	V	KL	Result	V	KL
1,1,1-Trichloroethane	200	ug/L		U	5		U	5		U	5
1,1,2,2-Tetrachloroethane	0.76	ug/L ug/L		U	5		U	5		U	5
1,1,2-Trichloroethane	5	ug/L ug/L		U	5		U	5		U	5
1,1,2-Trichlorotrifluoroethane	10000	ug/L ug/L		U	5		U	5		U	5
1.1-Dichloroethane	28	ug/L ug/L		U	5		U	5		U	5
1.1-Dichloroethene	7	ug/L ug/L		U	5		U	5		U	5
1.2.3-Trichlorobenzene	7	ug/L ug/L		U	5		U	5		U	5
1,2,4-Trichlorobenzene	70	ug/L ug/L		U	5		U	5		U	5
1,2-Dibromo-3-chloropropane	0.2	ug/L ug/L		U	10		U	10		U	10
1,2-Dibromoethane (EDB)	0.05	ug/L ug/L		U	5		U	5		U	5
1.2-Dichlorobenzene	600	ug/L ug/L		U	5		U	5		U	5
1.2-Dichloroethane	5	ug/L ug/L		U	5		U	5		U	5
1,2-Dichloropropane	5	ug/L		U	5		U	5		U	5
1,3-Dichlorobenzene	NC	ug/L		U	5		U	5		U	5
1,4-Dichlorobenzene	75	ug/L		U	5		U	5		U	5
2-Butanone (MEK)	5600	ug/L		U	25		U	25		U	25
2-Hexanone	38	ug/L		U	25		U	25		U	25
4-Methyl-2-pentanone (MIBK)	6300	ug/L		U	25		U	25		U	25
Acetone	14000	ug/L		U	100		U	100		U	100
Benzene	5	ug/L		U	5		U	5		U	5
Bromochloromethane	83	ug/L		U	5		U	5		U	5
Bromodichloromethane	80	ug/L		U	5		U	5		U	5
Bromoform	80	ug/L		U	5		U	5		U	5
Bromomethane	7.5	ug/L		U	5		U	5		U	5
Carbon disulfide	810	ug/L		U	10		U	10		U	10
Carbon tetrachloride	5	ug/L		U	5		U	5		U	5
Chlorobenzene	100	ug/L		U	5		U	5		U	5
Chloroethane	8300	ug/L		U	5		U	5		U	5
Chloroform	80	ug/L	5	U	5	5	U	5	5	Ū	5
Chloromethane	190	ug/L		U	5		U	5		U	5
cis-1,2-Dichloroethene	70	ug/L		U	5		U	5		U	5
cis-1,3-Dichloropropene	4.7	ug/L		U	5		U	5		U	5
Cyclohexane	13000	ug/L		UJ	100		UJ	100		UJ	100
Dibromochloromethane	80	ug/L		U	5		U	5		U	5
Dichlorodifluoromethane	200	ug/L		U	5		U	5		U	5
Ethylbenzene	700	ug/L		U	5		U	5		U	5
Isopropylbenzene (Cumene)	450	ug/L		U	5		U	5		U	5
Methyl acetate	20000	ug/L		U	50		U	50		U	50
Methylcyclohexane	NC	ug/L		UJ	50		UJ	50		UJ	50
Methylene Chloride	5	ug/L		U	5	1	U	5		U	5
Methyl-tert-butyl ether	140	ug/L		U	4		U	4		U	4
Styrene	100	ug/L		U	5		U	5		U	5
Tetrachloroethene	5	ug/L		U	5		U	5		U	5
Toluene	1000	ug/L		U	5		U	5		U	5
trans-1,2-Dichloroethene	100	ug/L		U	5		U	5		U	5
trans-1,3-Dichloropropene	4.7	ug/L		U	5		U	5		U	5
Trichloroethene	5	ug/L		U	5		U	5		U	5
Trichlorofluoromethane	5200	ug/L		U	5		U	5		U	5
Vinyl chloride	2	ug/L		U	2		U	2		U	2
Xylene (Total)	10000	ug/L		U	10		U	10		U	10

J - Denotes an estimated reporting limit ug/L - microgram per liter

NA - Not Analyzed for this parameter

 \mbox{U} - Analyte was not detected at or above the method detection limit RCG - Remediation Closure Guide

TABLE 7 GROUNDWATER SAMPLING RESULTS - TEMPORARY PIEZOMETERS **Exide Environmental Response Truat**

Frankfort, Indiana

Sample Location			L6-H	P-9-	11	L4-H	P-9-	11	L3-H	P-9-	11	L7-HF	P-10-	11	L8-HI	P -10-	11	L9-HI	P-10-	-11
Lab ID			R214	201-	01	R214	201-	02	R2142	201-	03	R2142	201-	04	R214	201-	05	R214	201-	06
Sample Date	2021 IDEM		10/18	3/202	21	10/18	3/202	21	10/18	3/202	21	10/18	3/202	21	10/18	3/202	21	10/18	3/202	21
Matrix	RCG GW Tap		Groun	idwa	ter	Grour	idwa	iter	Groun	dwa	iter	Groun	dwa	ter	Groun	dwa	ter	Grour	ndwa	ter
Remarks	Limit																			
Parameter		Units	Result	Q	RL	Result	Q	RL	Result	Q	RL	Result	Q	RL	Result	Q	RL	Result	Q	RL
Volatiles																				
Benzene	5	ug/L	0.41	J	0.5	1.2	J	5		U	0.5		U	50		U	5	1.5		0.5
Chloroform	NS	ug/L		U	0.5		U	5		U	0.5		U	50		U	5		U	0.5
cis-1,2-Dichloroethene	70	ug/L	15		0.5	180		5		U	0.5	2400		50	420		5	8.5		0.5
Ethylbenzene	700	ug/L	0.19	J	0.5		U	5	0.15	J	0.5		U	50	2.3	J	5	0.19	J	0.5
m,p-Xylene	10,000	ug/L	0.42	J	1		U	10	0.16	J	1		U	100	8	J	10	0.39	J	1
o-Xylene	10,000	ug/L	0.17	J	0.5		U	5	0.11	J	0.5		U	50	3.5	J	5	0.2	J	0.5
Tetrachloroethene	5	ug/L		U	0.5		U	5		U	0.5		U	50		U	5		U	0.5
Toluene	1,000	ug/L	23		0.5	2.3	J	5	5.3		0.5		U	50	0.8	J	5	0.75		0.5
Trichloroethene	5	ug/L	6.7		0.5		U	5		U	0.5	280		50	66		5	0.2	J	0.5
Vinyl chloride	2	ug/L	3		0.5	270	J	5		U	0.5	430		50	200		5	10		0.5
Xylenes, total	10,000	ug/L	0.59	J	1.5		U	15	0.27	J	1.5		Ü	150	12	J	15	0.59	J	1.5

Sample Location			L10-H	P-10	-11	L11-H	P-10	-11	L8A-H	P-10)-11	L9A-H	P-1()-11	L10A-F	IP-10	0-11	L11A-F	IP-1	0-11
Lab ID	1		R214	201-	07	R2142	201-	08	R2142	203-	01	R2142	203-	02	R214	203-	03	R214	203-	04
Sample Date	2021 IDEM		10/18	3/202	21	10/18	3/202	21	10/19	9/202	21	10/19	0/202	21	10/19	9/202	21	10/19	9/202	21
Matrix	RCG GW Tap		Groun	idwa	ter	Groun	dwa	ter	Groun	ıdwa	ter	Groun	dwa	ıter	Groun	idwa	ter	Groun	ıdwa	ter
Remarks	Limit																			
Parameter		Units	Result	Q	RL	Result	Q	RL	Result	Q	RL	Result	Q	RL	Result	Q	RL	Result	Q	RL
Volatiles																				
Benzene	5	ug/L	0.18	J	0.5	0.36	J	0.5		U	50	0.44	J	1		U	10	0.24	J	0.5
Chloroform	NS	ug/L		U	0.5		U	0.5		U	50		U	1		U	10		U	0.5
cis-1,2-Dichloroethene	70	ug/L	19		0.5	2.7		0.5	1500		50	71		1	380		10	21		0.5
Ethylbenzene	700	ug/L	0.19	J	0.5	0.18	J	0.5		U	50	0.96	J	1	1.6	J	10	0.57		0.5
m,p-Xylene	10000	ug/L	0.41	J	1	0.37	J	1		U	100	3.1		2	4	J	20	1.5		1
o-Xylene	10000	ug/L	0.2	J	0.5	0.2	J	0.5		U	50	1.2		1	1.8	J	10	0.54		0.5
Tetrachloroethene	5	ug/L		U	0.5		U	0.5		U	50		U	1		U	10		U	0.5
Toluene	1,000	ug/L	0.83		0.5	1.6		0.5		U	50	0.42	J	1		U	10	3.3		0.5
Trichloroethene	5	ug/L	0.49	J	0.5		U	0.5		U	50		U	1	510		10	1.1		0.5
Vinyl chloride	2	ug/L	23		0.5	2.1		0.5	330	J	50	75	J	1	170	J	10	14	J	0.5
Xylenes, total	10000	ug/L	0.61	J	1.5	0.57	J	1.5		U	150	4.4		3	5.8	J	30	2		1.5

Notes:

- J Denotes an estimated reporting limit
 U Analyte was not detected at or above the method detection limit ug/L - microgram per liter Q - Qualifier
- RL Reporting Limit

TABLE 8 GROUNDWATER SAMPLING RESULTS - PERMANENT SOIL VAPOR PORT WATER SAMPLING **Exide Environmental Response Trust**

Frankfort, Indiana

Sample Location			L13-G	W-1	019	D	UP-0	3	L2-GV	V-10)19	L4-GV	V-10)19	L17-GW-	-10	19	L5-GV	V-10	19	L16-GW	7-10)19	L6-G	$W-1\overline{0}$	19
Lab ID			R214	204-	01	R21	4204	-02	R214	204-	03	R2142	204-	04	R214204	4-0	5	R2142	204-	06	R21420)4-()7	R214	1204-0	08
Sample Date	2021 IDEM		10/19	9/202	21	10/	19/20	21	10/19	9/202	21	10/19	0/202	21	10/19/20	02	1	10/19	9/202	21	10/19/2	202	.1	10/19	9/202	:1
Matrix	RCG GW Tap		Grour	ndwa	ter	Gro	undwa	ater	Groun	dwa	iter	Groun	dwa	ter	Groundy	vat	er	Groun	dwa	ter	Ground	wa	ter	Groui	ndwa	ter
Remarks	Limit					FD of L	13-GV	W-1019																		
Parameter		Units	Result	Q	RL	Result	Q	RL	Result	Q	RL	Result	Q	RL	Result (2	RL	Result	Q	RL	Result	Q	RL	Result	Q	RL
Volatiles			•					•																		
Benzene	5	ug/L		U	0.5		U	0.5		U	0.5		U	0.5	J	J	0.5		U	0.5		U	0.5		U	0.5
Chloroform	NS	ug/L		U	0.5		U	0.5		U	0.5		U	0.5	Ţ	J	0.5		U	0.5		U	0.5		U	0.5
cis-1,2-Dichloroethene	70	ug/L		U	0.5		U	0.5		U	0.5		U	0.5	J	J	0.5	5.5		0.5		U	0.5		U	0.5
Ethylbenzene	700	ug/L		U	0.5		U	0.5		U	0.5		U	0.5	Ţ	J	0.5		U	0.5		U	0.5		U	0.5
m,p-Xylene	10000	ug/L		U	1		U	1	0.1	J	1	0.14	J	1	J	J	1		U	1		U	1		U	1
o-Xylene	10000	ug/L		U	0.5		U	0.5		U	0.5		U	0.5	I	J	0.5		U	0.5		U	0.5		U	0.5
Tetrachloroethene	5	ug/L		U	0.5		U	0.5		U	0.5		U	0.5	Ţ	J	0.5		U	0.5		U	0.5		U	0.5
Toluene	1,000	ug/L	0.14	J	0.5		U	0.5		U	0.5		U	0.5	Ţ	J	0.5		U	0.5		U	0.5		U	0.5
Trichloroethene	5	ug/L		U	0.5		U	0.5		U	0.5		U	0.5	J	J	0.5		U	0.5		U	0.5		U	0.5
Vinyl chloride	2	ug/L		U	0.5		U	0.5		U	0.5		U	0.5	J	J	0.5	0.19	J	0.5		U	0.5		U	0.5
Xylenes, total	10000	ug/L		U	1.5		U	1.5	0.15	J	1.5	0.19	J	1.5	J	J	1.5		U	1.5		U	1.5		U	1.5

Notes:

- J Denotes an estimated reporting limit
- U Analyte was not detected at or above the method detection limit
- ug/L microgram per liter
 Q Qualifier
- RL Reporting Limit

TABLE 9 SOIL SAMPLE RESULTS - MONITORING WELL INSTALLATION Exide Environmental Response Trust Frankfort, Indiana

Sample Location	Ī			MW	₋₁₁	-2-3	MW	-12-	2-3	MW	-13-	-2-3	TB-01	-1019	21	TB-02	-102	021
Lab ID	2021 RCG Soil	2021 RCG Soil		5030			50300			50300			50300			50300		
Sample Date	Direct Contact	Direct Contact		10/1			10/2			10/2				9/202		10/20		
Matrix	Residential	Non-Residential			Soil			Soil	J21		Soil			ueous			ueou	
Remarks	Limit (mg/kg)	Limit (mg/kg)		,	3011	L		5 011			5 011		АЧ	ucous		Аці	ucou	5
Parameter	Limit (mg/kg)	Lillit (llig/kg)	Unita	Result	Ω	RL	Result	Q	RL	Result	Q	RL	Result	Q	RL	Result	Q	RL
Volatiles			Ullits	Result	V	KL	Kesuit	V	KL	Result	V	KL	Result	Ų	KL	Result	Ų	KL
1,1,1-Trichloroethane	640	640	m = /1. =		TT	0.0043		ΤŢ	0.0037	I	TI	0.0055		TT	-		TT	0.005
1,1,2,2-Tetrachloroethane	8.4		mg/kg		U	0.0043		U	0.0037		U	0.0055		U	5		U	
	2.1	27	mg/kg		_			_	0.0037		U	0.0055		_	5			0.005
1,1,2-Trichloroethane		6.3	mg/kg		U	0.0043		U			U			U			U	0.005
1,1,2-Trichlorotrifluoroethane	910	910	mg/kg		U	0.0043		U	0.0037		U	0.0055		U	5		U	0.005
1,1-Dichloroethane	50	160	mg/kg		U	0.0043		U	0.0037		U	0.0055		U	5		U	0.005
1,1-Dichloroethene	320	1000	mg/kg		U	0.0043		U	0.0037		U	0.0055		U	5		U	0.005
1,2,3-Trichlorobenzene	88	930	mg/kg		U	0.0043	0.00069	J	0.0037	0.00054	J	0.0055		U	5	0.00068	J	0.005
1,2,4-Trichlorobenzene	81	260	mg/kg		U	0.0043		U	0.0037		U	0.0055		U	5	0.0005	J	0.005
1,2-Dibromo-3-chloropropane	0.074	0.64	mg/kg		U	0.0086		U	0.0074		U	0.011		U	10		U	0.01
1,2-Dibromoethane (EDB)	0.5	1.6	mg/kg		U	0.0043		U	0.0037		U	0.0055		U	5		U	0.005
1,2-Dichlorobenzene	380	380	mg/kg		U	0.0043		U	0.0037		U	0.0055		U	5		U	0.005
1,2-Dichloroethane	6.4	20	mg/kg		U	0.0043		U	0.0037		U	0.0055		U	5		U	0.005
1,2-Dichloropropane	22	66	mg/kg		U	0.0043		U	0.0037		U	0.0055		U	5		U	0.005
1,3-Dichlorobenzene	NC	NC	mg/kg		U	0.0043		U	0.0037		U	0.0055		U	5		U	0.005
1,4-Dichlorobenzene	36	110	mg/kg		U	0.0043		U	0.0037		U	0.0055		U	5		U	0.005
1,4-Dioxane (p-Dioxane)	74	240	mg/kg		U	0.43		U	0.37		U	0.55		NA			U	0.5
2-Butanone (MEK)	28000	28000	mg/kg		U	0.021		U	0.018		U	0.028		U	25		U	0.025
2-Hexanone	280	1300	mg/kg		U	0.086		U	0.074		U	0.11		U	25		U	0.1
4-Methyl-2-pentanone (MIBK)	3400	3400	mg/kg		U	0.021		U	0.018		II	0.028		U	25		U	0.025
Acetone (VIIBIC)	85000	100000	mg/kg		U	0.086	0.0059	I	0.074	0.012	T	0.028		U	100		U	0.023
Benzene	17	51			U	0.0043	0.0039	U	0.0037	0.012	U	0.0055		U	5		U	0.005
Bromochloromethane	210	630	mg/kg		U	0.0043		U	0.0037		II	0.0055		U	5		U	0.005
			mg/kg			0.0043		U			U				5			
Bromodichloromethane	4.1	13	mg/kg		U			U	0.0037		U	0.0055		U			U	0.005
Bromoform	270	860	mg/kg		U	0.0043		U	0.0037		U	0.0055		U	5		U	0.005
Bromomethane	9.5	30	mg/kg		U	0.0043	0.0010	U	0.0037	0.004	U	0.0055		U	5		U	0.005
Carbon disulfide	740	740	mg/kg		U	0.0086	0.0013	J	0.0074	0.001	J	0.011		U	10		U	0.01
Carbon tetrachloride	9.1	29	mg/kg		U	0.0043		U	0.0037		U	0.0055		U	5		U	0.005
Chlorobenzene	390	760	mg/kg			0.0043		U	0.0037		U	0.0055		U	5		U	0.005
Chloroethane	2100	2100	mg/kg		U	0.0043		U	0.0037		U	0.0055		U	5		U	0.005
Chloroform	4.5	14	mg/kg		U	0.0043	0.00095	J	0.0037	0.0013	J	0.0055		U	5	0.0012	J	0.005
Chloromethane	150	460	mg/kg		U	0.0043		U	0.0037		U	0.0055		U	5		U	0.005
cis-1,2-Dichloroethene	220	2300	mg/kg		U	0.0043		U	0.0037		U	0.0055		U	5		U	0.005
cis-1,3-Dichloropropene	25	82	mg/kg		U	0.0043		U	0.0037		U	0.0055		U	5		U	0.005
Cyclohexane	120	120	mg/kg		U	0.086		U	0.074		U	0.11		U	100		U	0.1
Dibromochloromethane	120	390	mg/kg		U	0.0043		U	0.0037		U	0.0055		U	5		U	0.005
Dichlorodifluoromethane	120	370	mg/kg		U	0.0043		U	0.0037		U	0.0055		U	5		U	0.005
Ethylbenzene	81	250	mg/kg		U	0.0043		U	0.0037	0.00034	J	0.0055		U	5		U	0.005
Isopropylbenzene (Cumene)	270	270	mg/kg		U	0.0043		U	0.0037		U	0.0055		U	5		U	0.005
Methyl acetate	29000	29000	mg/kg		U	0.0043		U	0.0037		U	0.0055		U	50		U	0.005
Methylcyclohexane	NC	NC	mg/kg		U	0.0043		U	0.0037		Ū	0.0055		U	50		U	0.005
Methylene Chloride	490	3200	mg/kg		U	0.017		U	0.015		Ū	0.022		U	5		U	0.02
Methyl-tert-butyl ether	660	2100	mg/kg		U	0.0043		U	0.0037		U	0.0055		U	4		U	0.005
Styrene	870	870	mg/kg		U	0.0043		U	0.0037		ĪĪ	0.0055		U	5		U	0.005
Tetrachloroethene	110	170	mg/kg		U	0.0043		U	0.0037		IT	0.0055		U	5		U	0.005
Toluene	820	820	mg/kg		U	0.0043		U	0.0037		II	0.0055		U	5		U	0.005
trans-1,2-Dichloroethene	98	300	mg/kg		U	0.0043		U	0.0037		TT	0.0055		U	5		U	0.005
trans-1,3-Dichloropropene	25	82			U	0.0043		U	0.0037		TT	0.0055		U	5		U	0.005
Trichloroethene	5.7	19	mg/kg		U	0.0043		U	0.0037		TT	0.0055		U	5		U	0.005
	1200		mg/kg			0.0043		_	0.0037		TT	0.0055		U	5			0.005
Trichlorofluoromethane		1200	mg/kg		U		0.00052	U			U			_			U	
Vinyl chloride	0.83	17	mg/kg		U	0.0043	0.00062	J	0.0037	0.0000	U	0.0055		U	2		U	0.005
Xylene (Total)	260	260	mg/kg		U	0.0086	0.00063	J	0.0074	0.0022	L	0.011		U	10		U	0.01
Metals	100	200	,-	12.7			P.CO		4.5	211				37.1) T :	
Lead	400	800	mg/kg	12.5		1.1	560	<u> </u>	1.1	214	<u> </u>	1		NA			NA	
Conventionals	110	110		4		6.1	4= -		0.1			0.5						
Percent Moisture	NC	NC	%	15.2		0.1	17.3		0.1	5.5		0.1		NA			NA	
J - Denotes an estimated reporting	limit																	

J - Denotes an estimated reporting limit

Bolding indicates exceedances of IDEM 2021 RCG Soil Direct Contact Residential Limit

U - Analyte was not detected at or above the method detection limit

mg/kg - milligrams per kilogram

Q - Qualifier

RL - Reporting Limit

NC - No Criteria

NA - Not Analyzed

TABLE 10 OUTFALL Z SAMPLE RESULTS Exide Environmental Response Trust Frankfort, Indiana

Sample Location	T				Outfall	ZSV	V		Out	tfall Z	Sed
Lab ID		2021 IDEM RCG Soil	2021 IDEM RCG Soil		503006					00615	
Sample Date	2021 IDEM RCG	Direct Contact	Direct Contact Non-		10/19/					/19/20	
Matrix	GW Tap Limit	Residential Limit	Residential Limit		Surface		er			edime	
Remarks	(ug/L)	(mg/kg)	(mg/kg)								
Parameter	(0)	(2 5)	(2 2/	Units	Result	Q	RL	Units	Result	Q	RL
Volatiles											
1,1,1-Trichloroethane	200	640	640	ug/L		U	5	mg/kg		U	0.0062
1,1,2,2-Tetrachloroethane	0.76	8.4	27	ug/L		U	5	mg/kg		U	0.0062
1,1,2-Trichloroethane	5	2.1	6.3	ug/L		U	5	mg/kg		U	0.0062
1,1,2-Trichlorotrifluoroethane	10000	910	910	ug/L		U	5	mg/kg		U	0.0062
1,1-Dichloroethane	28	50	160	ug/L		U	5	mg/kg		U	0.0062
1,1-Dichloroethene	7	320	1000	ug/L		U	5	mg/kg		U	0.0062
1,2,3-Trichlorobenzene	7	88	930	ug/L		U	5	mg/kg		U	0.0062
1,2,4-Trichlorobenzene	70	81	260	ug/L		U	5	mg/kg		U	0.0062
1,2-Dibromo-3-chloropropane	0.2	0.074	0.64	ug/L		U	10	mg/kg		U	0.012
1,2-Dibromoethane (EDB)	0.05	0.5	1.6	ug/L		U	5	mg/kg		U	0.0062
1,2-Dichlorobenzene	600	380	380	ug/L		U	5	mg/kg		U	0.0062
1,2-Dichloroethane	5	6.4	20	ug/L		U	5	mg/kg		U	0.0062
1,2-Dichloropropane	5	22	66	ug/L		U	5	mg/kg		U	0.0062
1,3-Dichlorobenzene	NC	NC	NC	ug/L		U	5	mg/kg		U	0.0062
1,4-Dichlorobenzene	75	36	110	ug/L		U	5	mg/kg		U	0.0062
1,4-Dioxane (p-Dioxane)	7.00	74	240	ug/L		NA		mg/kg		U	0.57
2-Butanone (MEK)	5600	28000	28000	ug/L		U	25	mg/kg		U	0.031
2-Hexanone	38	280	1300	ug/L		U	25	mg/kg		U	0.12
4-Methyl-2-pentanone (MIBK)	6300	3400	3400	ug/L		U	25	mg/kg		U	0.031
Acetone	14000	85000	100000	ug/L		U	100	2 2		U	0.12
Benzene	5	17	51	ug/L		U	5	mg/kg		U	0.0062
Bromochloromethane	83	210	630	ug/L		U	5	mg/kg		U	0.0062
Bromodichloromethane	80	4.1	13	ug/L		U	5	mg/kg		U	0.0062
Bromoform Bromomethane	7.5	270	860 30	ug/L		U	5	mg/kg		U	0.0062 0.0062
Carbon disulfide	810	9.5 740	740	ug/L		U	10	mg/kg		U	0.0062
Carbon tetrachloride	5	9.1	29	ug/L ug/L		U	5	mg/kg mg/kg		U	0.0062
Chlorobenzene	100	390	760	ug/L ug/L		U	5	mg/kg		U	0.0062
Chloroethane	8300	2100	2100	ug/L ug/L		U	5	mg/kg		U	0.0062
Chloroform	80	4.5	14	ug/L ug/L		U	5	mg/kg		U	0.0062
Chloromethane	190	150	460	ug/L		U	5	mg/kg		U	0.0062
cis-1,2-Dichloroethene	70	220	2300	ug/L	5.3		5	mg/kg		U	0.0062
cis-1,3-Dichloropropene	4.7	25	82	ug/L		U	5	mg/kg		U	0.0062
Cyclohexane	13000	120	120	ug/L		U	100	mg/kg		U	0.12
Dibromochloromethane	80	120	390	ug/L		U	5	mg/kg		U	0.0062
Dichlorodifluoromethane	200	120	370	ug/L		U	5	mg/kg		U	0.0062
Ethylbenzene	700	81	250	ug/L		U	5	mg/kg		U	0.0062
Isopropylbenzene (Cumene)	450	270	270	ug/L		U	5	mg/kg		U	0.0062
Methyl acetate	20000	29000	29000	ug/L		U	50	mg/kg		U	0.0062
Methylcyclohexane	NC	NC	NC	ug/L		U	50	mg/kg		U	0.0062
Methylene Chloride	5	490	3200	ug/L		U	5	mg/kg		U	0.025
Methyl-tert-butyl ether	140	660	2100	ug/L		U	4	mg/kg		U	0.0062
Styrene	100	870	870	ug/L		U	5	mg/kg		U	0.0062
Tetrachloroethene	5	110	170	ug/L		U	5	mg/kg		U	0.0062
Toluene	1000	820	820	ug/L		U	5	mg/kg		U	0.0062
trans-1,2-Dichloroethene	100	98	300	ug/L		U	5	mg/kg		U	0.0062
trans-1,3-Dichloropropene	4.7	25	82	ug/L		U	5	mg/kg		U	0.0062
Trichloroethene	5	5.7	19	ug/L		U	5	mg/kg		U	0.0062
Trichlorofluoromethane	5200	1200	1200	ug/L	2.1	U	5	mg/kg		U	0.0062
Vinyl chloride	2	0.83	17	ug/L	2.1		2	mg/kg		U	0.0062
Xylene (Total)	10000	260	260	ug/L		U	10	mg/kg		U	0.012
Total Metals	1.5	400	900	/ T	2.4	т	10		104		1.2
Lead Conventionals	15	400	800	ug/L	3.4	J	10	mg/kg	104		1.2
Percent Moisture	NC	NC	NC	%		NA		%	22.6		0.1
1 Creent Wioisture	INC	INC	INC	70		INA	<u> </u>	70	22.0	Щ	0.1

Q - Qualifier

RL - Reporting Limit

J - Denotes an estimated reporting limit

U - Analyte was not detected at or above the method detection limit

ug/L - microgram per liter

mg/kg - milligrams per kilogram

NA - Not Analyzed for this parameter

NC - No Criteria

Bolding indicates exceedances of IDEM 2021 RCG GW Tap Limit or the RCG Soil Direct Contact Residential Limit depending on matrix.

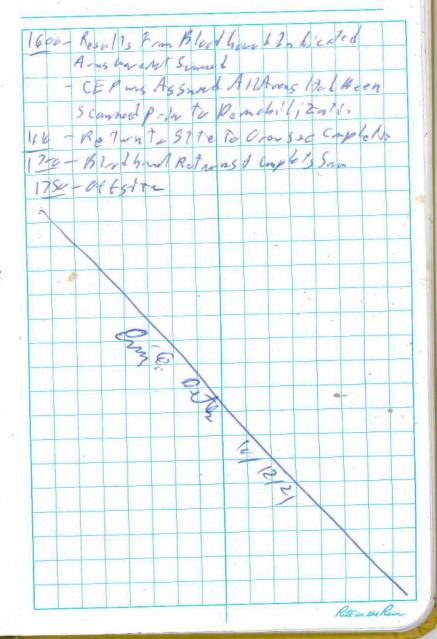


APPENDIX A

Field Notes

104 Location Fant	Sort Fr	7.0	Date 10	12/2/
Project / Client Ek	ile			
	,		Overcos	TES.
OTHE EVALL POTECCO	F. 0 - 460	1 (0,000)	Sn. 5/ De	
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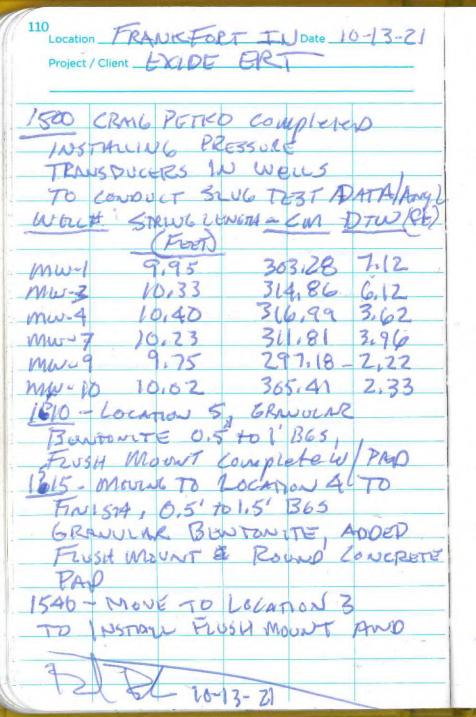


106 Location FRANKFORT DV. Date 10-13-21 Project / Client EXIDE ERT 0700 - B. BINCUE & CRAIG PETKO ON-SITE / ALSO CASCADE PURSONNIC JAMES ASHE, & TONY BREWER 9710 - ROMONDO HES PLAN, COMITING FOR DAYLIGHT 8800 - DISKUSSING FROTOLON FOR INSMELING SOIL PROBES, Permaent Arone Herry AVE CASCADE DOLS NOT HAVE TOWALAND BENTONITE FOR STALL ABOVE SCRIEN, ITUST HAVE PELLOTS 3/8" CASEMOE CANED SHOP TO CHECK ON POWDERED BENTON, TE. 5910 - CASCAGE HAVING GRANULAR BENTONITE TO BE DEZIVERED PU SITE FOR STALING SOLD UMPOR PROBES. 8915 MOBED TO LOCATION NO. I TO HAND ANGER FOR SON VAPOR PEOBE (SVP). 10-13-21

Project / Client EXIDE ERT

1930 - WET AT 4 BG5, 3.6 ppm, 3000 1 ppm 70 9940 - SITY OF FRUKFORT MUNIPER AUTHORING ALONG KOLYAVE, BRID - 765 421-3450. 9958 Jamon I word AT 31 1365, 30 mm6 work GRAVER FROM 2' TO 4', SET BOTTOM OF SCREEN AT 2'B65 WERL GRNED SED 1000 MOBED TO LOCATION 2 TO HAVE AVER 1020 - LOCATION 2' DRY AT 5 1365. NEED TO WANT FOR LONGER HOW ANGOR TO COME FROM EASCROE PLO - 6.4 TO 18.4 ppm 1025 - MOBED TO LOCATION 3 90 HALLO AUGER, PID-1-22.8 DOWN WATER AST 35365 10 = 3 - 2 | Rete in the Rain

Location FRANKFORS Date 10-13-2109 108 Location FRANKFORT Date 10-13-21 Project / Client ENDE ERT Project / Client EXIDE ERT LOCATION 3 - SETTING BOTTONS 1220 - moiso To Cocasion 5 OF SCROW @ 2.5' B68 TO HAND AVER SUP. FOR SUP, 6" tower GRAVER DOWN 5 B65, WARRE TO ABOUT SURCEN , ROMANG SEE IF WATER COMESIN PID-1300 - MOBED TO COCKTON 1,5 TO GRADE FOR GRAWURAR DID - < LPPM, 4'Z" DTB BUNDONTE 1110 - MOBOD TO LOCKTON 41 CHUK LOCATION 2 WATER unrer, SOTTING BOTTOM DF SUP SCREW AT \$ 2,51 Were Grover Set to 1.5' BGS AT 4.5' 60W6 TO SET SUP BUNDONTE GRUNIAL ADDOD 50000 @ 3,5' 365 NOW TO LOCATION 6 From 0,51 TO & come Fac want in Course 4, Lourse 4 1,5 thypano w/ with 1130 - BACK AT LOGAROW 4 BURAN DIGGING FOR FLUSH Mount, ADD ROUND CONG. PND 1430 AT LOCATION 5, 5 B65 WATER AT 45B65, SOTING 64 ware Serrus Borron OF SUP SCHOOL BOTTOM (D) SUP SCREEN PL 3,5', ADDING 3.5' BGS, WILL GAMES TO 251365 Well GRAVER TO 15 1865. DID- < I DOM, INSTALLATION OF FLOSH MOUNT & ROUND 1140 - CASCADE LUNCH BROME CRAIL HUNG UP 3 SITE SIGNS CONCRUTE PAND. INE ON EVEH OF 3 GATES 12 1 Rd 10-13-21 Oc 13-21 Retein the Rain



Project / Client Live BR To

Round concrete cours Pros 605 - INSTALLO FRUSH MOUNT & Rouse Concrete PAD AT 1640 - INSTELLED FLUSH MOVEY AND ROWED CONCRETE PAD AT COCATION CLEMING UP FOR THE DAY. 1700 - ALL OFF-SLIE 10-13-2 Rite in the Rei

112 Location FRANKFORT Date(0-14-2) Project / Client EXOE EZT 0745 - B. BETGLE & C. PUTKO ON-SITE, CASCADE ON-SITE JAMES ASHE & TONY BREWER, H& 5 METTING, 0815 - MOBED TO LOCATION 13 TO WETHLE SUP, HAND AUGURED TO 5 BGS, NO WATE, WILL WAST, POD- 2 LOOM 0835 - MOBED TO LOCKTION (4 TO WEIMIL SUP, HAND AVELRED TO 5' BGS, NO WATER, WILL WAST, PID- <1 ppm 0855 - MOBED TO LOCATION 15 TO WSTALL SUP, HAARD AUGERED TO 5' BGS, WET AT 5', 1.54 WATER WATER SETTING SCREEN BOTTOM AT 4 BGS ADD WITH GRAVER TO 3' 1365, 8,563' Bentonte GRANVIAL 1100 RID Completes FLUSH Mover AT LOCATION 19, HOLD AUGERED TO 6'BGS, SET BOTTOM OF SCREW @ 4,5

Project / Client LYCLDE 12KT Date 10-14-21 113

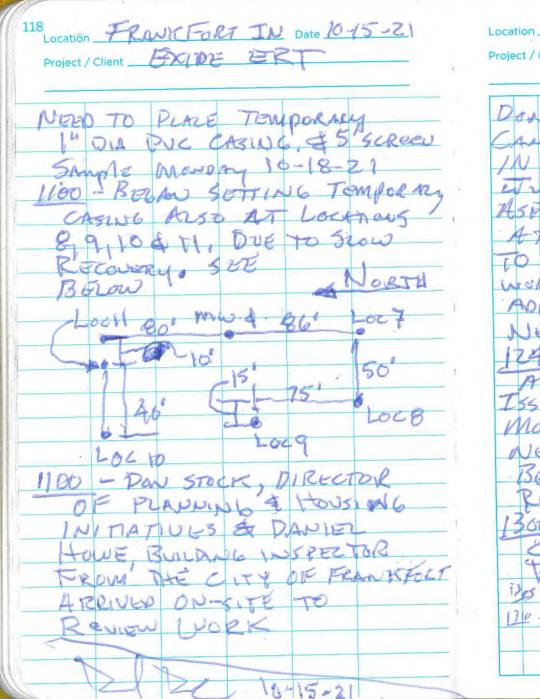
WELL GRAVES 35 TO 6 05-35 PAD AT LOCATION BELOW TUSTALLING BEDDING SUP. PID- < 100m 15 - AT LOCATION 17 MOND STORM SOUR, CORVER OF cercy AVE & E. M. CLURG Down 2,5 B65 A7 LOCATION 17 (BEDDING WHERING Appenes to BE WATEL NO COROUNTER OBSERVED, CU WILL SET SCREEN BOTTON 25 & weil GRAVER TO 0.5' BCS. PID- 41 ppm 1145 - MOBIO TO COCATION 16 100' SOUTH OF LOCATION 17 1200 - CASCADE LUNCIS 1230 - BACK AT LOCATION 16 1300 - DOWN 3' @ LOCATION

114 Location FRANKFORT IN Date 10-14-21 Project / Client EXPE EFT PID-21ppm CLAY & GRAVEZ, NO GROVED WOTER SUTTING SCREEN BUTTON @ 3'86 wow GRANT @ 1.5' 1365. Somul Frus H Mount & Rouns CONCRETE PAD, 0.5-1,5 BENTONITE 315 - AT COUNTON 13, 14AND AVENTO TO 6, DRY, TO SET BUTTOM OF SUP SCREEN @5', well Graves To 4' B65, 400 - AT LOCATION 18 TO SET SUP, PID-< Ippm HAND AUGUROS TO 31 B63. MATINE GANER & CLAY BETTOM OF SCREEN @ Were Games to 1.5', 0.5 10 1,5' BUS LEAN VINE BENTOWNE, HYDRATED. 1430 - CRAG POTKO Removes THE PRESSURE TRANSPUCERS FROM THE 6 WELLS THEY TOR SING TEST AVALLED.

Project / Client FORT DN Date 10-14-2 115

1430-1630, INSMED FRUSH Mounts sup Round ConcRETE PADS AT LOCATIONS 13, 15 WERE FRANKING WITH GRANULAE BRUTONTE # HYDRITED W/WHITER TO 30-BUTEN CLEMING UP FOR THE DAY 045 - ALL OFF-SITE

Location FRANFORT IN Date 015-2 1117 116 Location FRANKFORT Date 10-15-21 Project / Client LNDE FRT Project / Client BXIDE BRT 0740 - B. BUEGLE & C. PETKO NORTH 0145 CASCADE ON-BITE TODAY COLLECTIVE HYDR-ROAD PUNCH SAMPLE HELD HES MEETING 0905 - MOBED ON-SITE MY DOCATION 7, 86 SOUTH OF WORKING AROUND 650 PROBE. 0815 - MOBING TO LOCATION MW- 4 WATER ENCOUNTERED 3 TO HYDROPONCH, LIGHT RAIN NEW DUE TO OVERHEUD AT ABOUT Z', PENUTO COLLECT GROUPWATE Sample ELETRIC & UNDERGROUND Tom 10'-11' SIMULAR TO UTILITIES AT LOCATION 3 SAMPLING DEPTH AT MW-4 NEOD TO IN STREET 5 DISCUSSED W/ JAW, D. GREG S. AGREED. GROUNDWATER SIAM PLIEG TO & GREG SI OK, NOW TO OBIMN APPROVING FROM BE CONDUCTED W/ PER STALTIC THE CATY, CITY INSPECTOR Pump. 1015 - SERBON OPEN 2' From SCHEDULED TO ARRIVE 9- il', Slow Roccoursey ON-SITE AT 1000 TODAY 1030 - NO GROUNDWATER, TO DISCUSS 0-15-21 Rete in the Rai



Project / Client EXIDE EXT

DON \$ DAVIER STATED WE PUT HUDED PUNCHE. N ROAD - HERRY MVE AT 1115, AUSO THEY REGULSTED TO CALL STREET DEPT WHEN WORK TO GLOSE ROAD, NO Appropriate Poem of OR MONEY 1245 - Computer Prozometer AT LOCATION 5 7, 8 & 9 55US AT & 10. TO BE AUDRESS 02027 10-18-21 CASCADE 18703 TO LEAVE SLIE TO BE IN COMPLIANCE W/ DOT 1300 - Hanges Fire Book to ERAL PETKO FOR THE DAY 15-21-01 13 5 Promis Fours & Securing Equipment ansite 170- Alloftsite my & Pets 10/3/21 Rite in the Rais

120 Location Fruktat, In Date 147/21 Project / Client Exitofrat 1720 - orgita to reet ben Class CPace Annly Toul) To So Tong MapilaLink 1770 - Paco Setting up Lab To Ran craning 1+ 8755 - Pace Parished they gite Foy Copy - Will Serve Site upon Reportion 1800 - CBP Ottsite

Project / Client Experse? Date 10-18-2121

2730 - B. BEEGLE & C. PETKO BUN CLAAS FROM PAGE CU/MOSTLE LAB ON-SITE O CASCADE ON SITE Janes Af Tony B. Smart 10 PROBE 2810 - PAUS MOONE CAR. CALIBATING INSTRUMENT FOR WATER MARRY 515, 300 of there to conserve. 815-C. Perko Serra6 Up to court alongunt 1830 - CASCADE FILLING UP GOODTObe a / Free 9 OSES- MOSED TO LOCATION LO ON-SITE TO SET PRETOR STREETS DEPARTMENT 765-659 562-875-8530 70 2912 CLOSE KOLLY AVE FOR 10-18-21

122 Location FRANK TORT Date 10-18-21 Location FRANKFULT Date #0-18-2 123 Project / Client FODE ERT Project / Client ExpE EFT HYDROPUNCHES IN ROLD 1035 - MOBING TO LOCATION HYDROPUNCH ocations 3, 4 & 6. 1910-125 mins 14 DIA - BUP 6 LOCATON PHOROMETER AT LOCKTON 10 ASPINIT ROAD 1920 - MOBER TO LOCATION 1-6-48-9-11 HOLE COLD #11 ON-52 TE, Dry AND PATRICED W/TAR PIEZONETER AT LOCATION 10 15 SET AT 11' B65 W/ 1110 SUTTING HYDROPURKH 9'-5' SCREEN, Cocamon II PID 2 1ppm 11' B65, SONS POD - < Don 1125 - COLLETT 3-40m1 SON SMEZZES Viang OF GROWSOUNTER 0945 - SeTTING 1" DIA # L6-HP-9-11, 1125 PRESOMETER AT LOCATION 14 SET AT 11' BGS W/5' DELIVERED TO MOB 1/50-mois on to Location A SURBEN. 000 STREETS DEPT CAME 70 Hypsopunch PID-<1 ppm AND CLOSED ROAD ALONG KELLY AVE TO ASDROPUNCY L24-HP-9-1 CASCADE OUT TO GET KELLY AVE ASPHANT PATCH OF STARANT 10-18-21 10-18-71 Rete in the Rain

Location FRANK FORT Date 18-21 124 Location FRUK FORT Date 10-18-21 Project / Client Loans ERT Project / Client EXAPE LET 1515 - COULTRED 3-40ML 1240 COLLECTED 3-40 Mr VIAL 3 OF GROUDWATER, SAMPLE 7-APP-60-11 # L4-HP-9-11, Hout WAS ASPEARET COLD PATCHEN 6 ROUNDWATER 3 AVE CU/ TAR 258 CASCIDE CUNCIT 325 - MOBER TO LOCATION 3 TO HYDRODUNCH, LOCATION 3 # 19-419-10-1 - courceto 3-40 m MANS GROUDWATER SURAL L3-HP-# 418-HP-10-1 - COLLECTED 3-40-112 Vins GROUDENTER SAMPLE terry but 1645-BBOGFFSite 410 Consoro 3- 40 mc VIAS 1700 - Prillers Offsite # 13-HP-9-11 ASPAALT 1715 - CEP 08 55 --Ben (Mice) w Ulsecons Sity Mean Ass COLO DATEM W/ SEARANT Person ture Fallening Constator of Puntys 13 IN HOLE 1500 - OPELIED UP ROAD tra. Rote W1672 CALLED STREETS DEAT FOR THEN TO PICK UP CONES 7 10-18-21

Location Frank Yart IW Date (4/18/2) Project/Client Ebite ENT (EP Portone & Vapor Sampling ACTI vites Ag F. Ilong 0240- At Locatia 14 Parsil Ray Sumply CASO COCATONIY PASSED HOLDON Scooning (0,0%) - Sine miter founded Puring Snip (by Regulary Ros tyT 1000-CNoct 5 my 2 L14-DSG-1018 1040 - AT LAUSTER 1 PORP SUIT Gas Per 1045 - Holling Tost Porssed (00%) 1050- Callect Smple L1-056-1018 NOC - At Low Mon 2 Pox Sos (Gas Port 1136 - unia Produced At Location 2 Counting & 1134-AT Locata 13 Dogo Soil Gas Port Cannot Suple due to unter Production 1140 - AT Lounton 18 Sewer Bedding Cozation 1143 - Prosed Hellow Test (0,0%) 1148 - Collect Smple 1-18- 5BS-1018 1155 - At Low Ston 3 Deep S. J | GastenT 1204 - PBS Hellmtnt (0.096) 1215-Congot Smple L3- PSG-1018 - Cole of Puplicate Sayle Pup-61 Word 1155 220, - At Lunny Doep Sol Gas Port = Con 12 Mot 5 mp & Mine to Water Production Mr- AT Locatins Doep Soll Gas Port - Cannot Sompleduc to water Production Comes hants 16/19/21

Location F-nktat IN Date 10/18/21 127

Project/Client Exide ERT

1246 - AT Location 17 Sever Medding Soule - Cald not Suple On to water Broductive 1700 - AT Location & Somer Rodding Sayle - Cald P- 1 sayor Dre To with Rudi IN 1310 - At Location C Deep Soil Cus Port - Corld not say lednete mira Produte 132 - ATLANS 15 DOEPS, (GASPENT 173 - Pass Helm Sevenin (O. O/L) 1925 - Colect Single LIS OSG-1018 1440-Conect Manhole (Samo de Gas No Vapor Read 13500 M. 171-Gas May 12 1455- 1-16 of Munholo A Somple of Gas The Upper Readings on Multi Metor Use- Magin Agton plins to Pargo myton For Myalale - Pesty Proping Lown 1/ Apra 5 1548-STOP Rempi - Rongred = P. 75 Gal Cristat Flow 16 CK - 137 th Krupos y levent ton 2 1615 + Stop Pompso Rome L.W. 25 Wal, Royalar Fla 18 25- ATTIMET LOCATED IS CHOW LEEDING 163 - Stop rumpso - Romac & Ign/ on STAN Fra eng & poten (40/18/1) Rite in the Rais

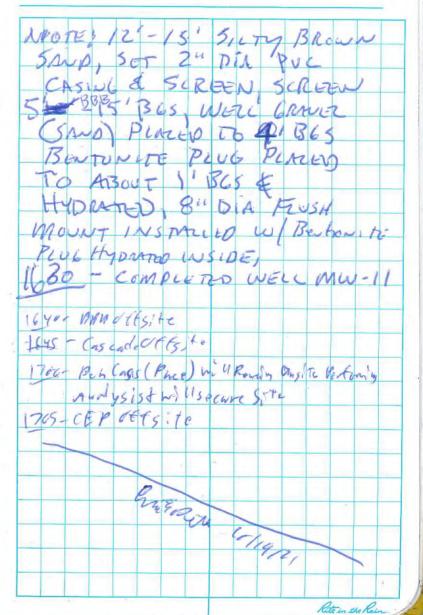
128 Location FRANKFORT PW Date 10-19-21 Project / Client ENDE ERT 0730 B. BIEGLE & C. PUTKO ON-SITE, RENEWED HYDRO-DUNCH DATA, NEWD TO AYDRO PUNCIA BA GA, IOA 11 A Locanous, Vin MOBILE BEER CASCARE HARIVED AT SUTE, JAMES & TONY \$ 5 METETING CONDUCTED 1838 - MOBED TO SET 1" DIA PIE ZOMETER, SET AT 11 PIE ZOMETER, SET TO 11 B6550000 0930 - MOBERS TO CATTON FOMETER, SUT TO 11' B65 000 mo Box TO LO GATION PLEZOMETER SET TO IV B65 W/ 51 Scleen 21 124 10-19-21

Project / Client EXAPE ERT Date 10-19-2199

COLLECTED 3- 45 mc VIA 1145 MOBER TO LOCATION 9A SAMPLE WATER (GROWANDER 9A-HP-10-11, COLECTED 3-40 ML VANS, AT 1155 1200 - MOBER TO LOCATION IOA TO COLUCK GROOND WATER Constitut 3-40 mm VIALS

130 Location FRANKFORT INDate 10-19-21 Project / Client EXDE ERT 1215-MOBER TO LOCATION HO. 11A to Court GROWN -3-40 mi VIAIS, COLLECTED AT 1230, 1240-1310 - CASCADE LUNCH 1315 - MOBED TO LOCATION 12 TO INSTAU NEW WELL MW-11, 0-8"-1" A3PHIT & BROWN CLAY TRACE GRAVEL WET AT 3' BGS, COLLECTED Son SAMPLE From 2'-31 # MW-11-2-3@1416, 5-5' WAS HAND AUGGED TO CLUBE UTLUTIES 430 - USING 414" ASA'S TO 151 500 - AT 12' BGS, BROWN CLAY TRACE GRAVES, 1510 - DOWN 151 B65, BEGAN SETTING WELL, 10-19-21

Project / Client BODE ERT



132 Location Lyide Fre KENT Project/Client Exide ERT - CEP Completed Supply As Fullows 0810 - Contino Sever Man holesmotis 0845 - Clect Manhole E Tor Voc's agran care of manhale F For Vocas cals - collect Munhales for Vacis 0470 - Chect muhale A for Vac; - Collect Pap-02 at Gazs For Manhole H 1000 - Collect punhole I For Vot's 1045 - Discuss Vapor Ports With Project Personnell - Will Supple water from Vapor Ports 1210- Colect words, Suple -13-CW-1014 For VOC'S - Collect Pup-02 For This Point at 1215 1216 - Colect mater Suple L2-64-1019 For VOCi 1246 - collect watersmole Ly-Gw-619 Par VOCS 1709 - Colle it water Suple (17-6W-1019 For Vac) 1320- Come of with a Sumple LS-6 W-1019 Far Voci 1734-Collect pota simple L16-6 willing for Volij 1348 - Callett intersimple LE-GW-1-19 Fer VOC) 1926- collect out tall 2-5w surtice witasmole 1725 - Conect duter 12-5-1 gedinal 9 miple Rus C. Rits 4/19/11

Project / Client EM DE ERT Date 10-20-21 133

730 - B. BUTGLE & CRALL PETRO ON-SITE, ALSO BEN FROM PACES MOBILE LAB 2800 CASCADE ON-SITE JAMES AND TONY CABCADE SETTING UP TO DO DEKON OF AVERS! HES METRAG CONPUCTED CRAIG PETRO SETTING UD TO SAMPLE MANHOLES, 0845 MOBED TO MW-12 Were LOCATION CLOCATE 78), ASPINIT Z" THICK ASPHAT, 54-121365 Sarry BLACK GRAVEZ WET 2' NOT ABLE TO Court symple DUE TORED BRICKEGRAVEZ, NOTE : HAND AUGURED TO & BOS URBAN FRU BLACK BRICK TO A! WILL ADVANCE 444 OLA HSA FROM 41 BGS

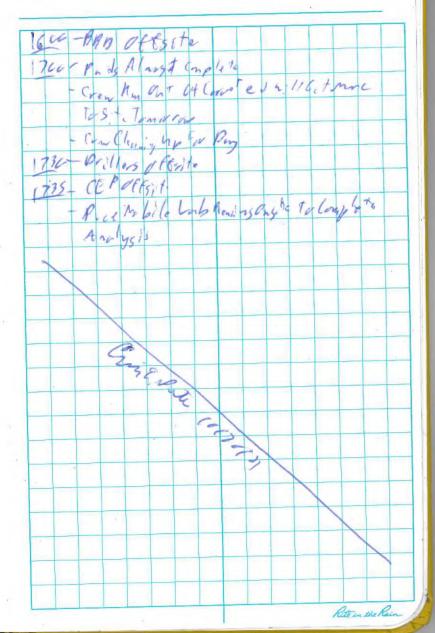
Location FRANKFORT IN Date 10-20-21 Project / Client EXIDE ERT 0950 - invecto sample of AUGERS AT 2'13' # MW-12-2-3, For VOC 1000 - HSA'S DOWN 5.5' TRANSITIONED INTO SOFTER 812 5.5' - ORBAN TYCE, 6 RANGEZ ROD BRICK, 4" PIECES OF METAL BLACK, WET 5-9' BLECK, VERY WET, SILTY, BLAYEY TUE SAVO. 91-1511865 BLACK SLOPPY SOUD SILT W/ TRACE FINE SAND - BLACK SILY WIFWE SAND. 1045, - HSA'S DOWN TO 15' BGS WILL BEGIN TO SET WELL @ 15' Z" DIA PUC CASING & SCRUEN, SCREEN 5'+15' Well beauty save To 4' > bu 10020-21

Project / Client LOOPE LETT Date 16-20-2 135

1100 - CASCADE DROPPED OFF PRT SOU UMPOR SAMPUNG 1140 - SAUD PAIK TO 4' B65 BUNTONITE HOLE PLUE TO 64 B65 WILL PLACED PAD & STICKUP CAPER, 1195 - CASCADE DERUNNO HSAS 1323 MOBIO TO MW-13 TO INSTALL WELL CAMB PETKO 15515TING 6" TO 8" GRAVEL BASE, DK BROWN CLAYEY SAID, SOME GRAVER, LOAD Prices, Souple MW-13-2-3 Dey WS/MSD SAMPLE COLLECTED BY CRAG PETEO HARD MULING 3709" URBAN FLL, BED BRICK materiar, 1315-1330 CASCAPE LUNGH 12-22-0

136 Location FRANK FORT IN Date 10-20-21 Project / Client 6 DE ERT FILL WITH RED BRILL & MOTH (4") OBJECTS, 41/2 SMATURE TO GET INTO BLACK CLAY MATTERAS, 1345 - WET AT 71, NO 1415 - DOWN 15 BAS TO SET WELL AT 15' B65 7'-15' Soupy BLACK CLAY MATERIAS, WET, 1500 - completes were mw-13 ZHDIA PUC CASING & SCRIEN SCREEN 5'-15' BGS, 5000 TO 4' BENTON TE HOLE PLUG 1 Hypratro TO 6" 1365, 540 pr mw-12, sau CUTTING PANEMENT TO SET CONCRETE PAPE STUCUP CLOSURE,

Project / Client 6x10 E GR 7



138 Location Frankfort/IN Date 14/29/21 Project / Client Exide ERP - (EP Completed Supty A, Follows Tody
0836- Chatterne Somer Muchole Sumpling of Napores : C535 - Auntile NIS Could To be A Reitingalow your Sternornite - Did NIT Smyle 0850 - theit Supple Montrole J Far VV6 (0,0 PM) 0720 Collect Suple Manhole L Forvor's (0,3 PM) - Crivet Pup-04 A+ 0910 OASU-Cament Somple MH-10 For Vocis (0.3 PPM) - Significal moto Prosent In MHI-100

Location MANKFURT IN Date 10-21-21

Project / Client LXIDE ERT

0740 - B, BUEZLE & CRAIGP. PACE MOBILE LAB. 0800 CASCADE ON SINE, ITAMES Coursete Pro AT MW-13 & DERON AUGURS 5 MES BIBUEGLE GOING TO SAMPLE PRT 3VP W/ CRM6 P. That TI & Smy mans HELD. 0830 - AT LOCATION 6 TO FOLLECT PRT- SV SAMPLE CUTILIZENS PRT 545 FEM Smu De # LO-PRT-1-5 DEPTH 1.5', TIME 0844. 0855 AT LOCATION 16 TO COLLECT PRIT-SV SAMPLE SAMPLE # LIG-PRIT-1 DEPTH 1", TIME - 0858 0905 AT LOCATION 5 TO COLLECT PET-SV SAMPLE 12/ 10-21-21

Location TRANKFORT IN Date 10-21-21 Project / Client ______ EXIDE ERT SAMPLE # L5-PRT-1 DEPTH 1', TIME - 6910. 0920 AT LOCATION 17 TO COURT PRI-SV SAMPLE # LIT-PRT-1, DEPTHI TME - 0922 0930- AT LOCATION 4 TO COLLECT PRT-SV SAMPLE # 14-PRT-1, DEPTH 11 TIME -0944 0955-AT LOCATION 2 TO COLLECT PRT-SV SAMPLE # LZ-PRT-1, DOPTH 1010 AT LOCATION 13 TO COLLECT PRT-SV SAMPLE # 413-PRT-1, DEPTH 11 TIME-1018 030 - RETURNING TO SITE TO FINSH DECONNING AUGURS 15-15-01

Project / Client EXIDE ERT Date 10-21-21

1100 mosep To MW-14 tourner to instructure 115- HAVO AVGERED TO 21. ward AT 6"1365, 1145 - 5 maro movercorco A'A" DIA HEA DUE TO REFUSAR AT Z' W/ HAND 1205 - Opun a' NO CUTTURES Romano to GROUD, NO SAMPLE COLLECTED. 1230 Down 10°, CUMNE BROWN VERY WET SOURY SUT, TRACE FINE SAND 0-3" ASPHART 3"-10" ERMER BASE 745 Down 151 BCS TO SES well, 10 - 5 VRBA File NO Bricks OR Morns OBSERVED 35 -15 BROWN VORY WET Soury sury w/ Frue SAUP 10-21 - 2 Pete in the Rein 142 Location FRSUKFORT IN Date 10-21-21 Project / Client EXIDE ERT SETULG 24 DIA DIE CASING & SCROWN, SCROW 5'-10' BGB, SAND (Well GRAVER) SET TO 4"1365, REMAINDER TO 6" BGS W/ BOWTONITE CHIPS Hyprotop 1325 - who mw-14 completes BEGINNE CLEANUID OF CUTTINGS TO SET CONCRETE PAN & STICKUD, 345 - PACE RELOCATED GENERATOR OUTSIDE FENCE, CRAIG P. PLAZED CHAN & LOCKS, RENTAL COMPANY TO PICK UP LATER PACE HOOKING Up MOBILE LAB TRANTE TO TRUCK, GENERATOR TO BE PUCKED UP TOMORROW, 400 - PACE MOBILE LAIS & TRUCK LEAVE SITE 10-21-21

Project / Client Date & FRT 143

1430 - COMPLETED CONCRETE PADE STICKUP AT MW-14, CASCADE CLEANNE UP. LONDING GEOPROBE ON TRAILER. CLEANNG UP AUGER DEZON AREA 1510 - moiso 70 mw-11 TO Deveras WELL, PUMPED 10 GALLOWS TO DRY, LOST 1 GALLON STARTING TO BE CLEAR WILL WAT 15 MIN TO TO RUCHARGE 1535 - Pumper ANOTHER 16 CARLONS TO DRY, MUCH CHARTE WILL LET BEZHARGE AGAIN, CASCADE ADDED MORE ASPERT PATCH TO 3 LOCATIONS ON KELL AVE ROADWAY, 1600 Pumpers ANDROR 8 GALLONS, ROBATION CUERR WILL COME BACK TOMBEROW

Location FRANKFORT JN Date 10-71-2) Project / Client EXIDE ERT 615- AT MW-12 TO DEVELOP 636 PUMPOD 30 GALLOUS AT MW-12, STARTING TO CLEAR UP, WHI COME BACK TOMORROW, 1635, AT MW-13 TO DEVELOP WELL. 1650 - Pump 30 6AZLONS AT WW-13, STARTING TO CLERR A LITTLE, WILL COME BACK TOMORROW, 1655 AT MW-14 TO DEVELOP 70 - Pumpro 17 6Azianz AT MW-14, mospy went DRY, will COME BACK TOMORROW, 1704 - BAb Cttsite - Orillers Security Sico 1720- Orillers Offgite - CE POFTSite Print 10/21/21

Project / Client EXIOF ERT Date 70-21-2 145

0740-BUDGICE & C, PETRO 0755 - CASCROE ON-5175 JAM63 & 700 HES MITTING 0815 Bean To Conney = DOVECOPING & NEW WELLS mw-11 - coupy, Net sung Pumpero 6 GALONS TILL DE WATTUG 10 MINUTES ADDITIONE A GALLONS PUMPED, RELATIVE 0845 AT MW-12 TO CONTINUE DOVELDING WELL, SILTY THE FRST /2 GALLON, THEN CLOUDY, Pumper 30 Grans, Romanier SUMR 0915 - AT MW-13 TO CONTINUE DENSIDENS, PUMPED 30 CHIONS warm sury, we want 60 MINUUZZ 15-22-01

146 Location FRANKFORT IN Date 70-22-21 Project / Client EXIDE ERT ATMW-13, PUM DED ANOTHER 10 GALLONS, WATER WAS 10 MINUTES DEVELOBMENT 1048 - Driles Stagin Darn Drines + Shiter Drings ts. te Todelise Disposal Suptist las - Oillas Loading Equipunt Generator Is Styed ontside Gate For Pick ap te Science upen Deporture

The manufacturers of *Rite* in the *Rain* all-weather writing products are grateful to the numerous environmental experts who have contributed to the development of this book. Should you have any additions, improvements or corrections for future publications of this field book or have suggestions for other environmental field book formats, we welcome your input.

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- RE Recording Error
- E Calculation Erro
- TE Transcription Error
- SE Spelling Error
- CL Changed for Clarity
- DC Original Sample Description Changed After Further Evaluation
- WO Write Over
- NI Not Initialed and Dated at Time of Entry
- OB Not Recorded at the Time of Initial Observation

Note: Error code should be circled, dated, and initialed when recorded.

Hazard Classifications

- Class 1 Explosives
- Class 2 Gas
- Class 3 Flammable Liquid
- Class 4 Flammable Solids (Potential spontaneous combustion, or emission of flammable gases when in contact with water)
- Class 5 Oxidizing Substances and Organic Peroxides
- Class 6 Toxic (poisonous) and infectious substances
- Class 7 Radioactive material
- Class 8 Corrosives
- Class 9 Miscellaneous dangerous goods

Container type abbreviations (for sampling guidelines)

BR - Boston Round • ABR - Amber Boston Round • AJ - Amber Jug • AWM - Amber Wide Mouth • Poly - Polyethylene Bottles • BOD - Bottle • CWM - Clear Wide Mouth

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Exite Fraktri, IN Date 12/8/21 Project / Client Exide Aw-Z D73 well 14 16 My - 11 3-86 17.68 NW 1722 17.28 Mar 14 MV-Y 19-89 Round of with Couli Combete Food To MW-2 to By in Small. 0948 - AT MW-Z BOSIN Knyl DUN DIB SPE 1000 7 29 -166 1446 9776 13.93 0736 9.793 09.4C 5.47 0.800 118 9 6.46 9.6 0944 16.89 7.23 109.2 4.67 -10.17 7.23 0948 3.62 0.74y 104.4 2.46 7.23 100.7 4164.10 00152 4 36 0.784 4.92 0.790 944 278-7.23 1000 -4,20 0.705 7.23 9.68 244-1601 - Suple MW-2 For Vac Marnid my 161 First born of 10° - Herder Munio En & Pora (2/8/21

Location Trim ton, Ir Date 12/8/27								
Project / Client Enile								
mw =0/mw-3								
1036 - A-1 Mn - 16 Be 3/2 Propo (m)								
-DTW 2 42 161(7)F								
DTB Pay 57 at 13								
Tim Jano SPC DC pH GAR Turb WIL								
1978 4.25 10863 14.24 7.12 75.8 778 -								
K42 5.93 1:618 6.64 7.03 74.3 772								
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1950 5.65 1.711 5.46 7.61 71.9 114 -								
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Location Frank Part Date 12/8/21 Project / Client Exide MW-3/MW-7 1212 982 6674 244 7.22 660 548-1216 19-77 0,067 1.82 7,23 63.7 75-5 122011-23 0,666 443 2236 65 25.46.99 1224 11.11 9666 1-24 7.22 57.5 21-0 -1228 139 0661 Loy 7.23 54.8 149 -132 11-69 0661 091 7,23 51-9 15.1 6.9 y 1236 1464 ECS8 0,84 723 50,5 1884-1246 1188 0619 0,83 7.23 48,5 14.19 1244 163 0 Gry 0,79 723 463 1008901 1245 Finished Suplamw -3 For Vac - Collect ms/ ms & Hero - Finished At mw-3 Renned 2 2901 Fin DTW = 7.01 - Head to MW-7 (214 - A+ MN-7, Besis Parping 1001(2") PTO pomyo SotA) 17 Time Tomp Sec pe pt and Tomb bits 1322 9.22 1135 4.36 6.67 56.4 567 -1326 9.90 1242280666 55.1 5269 -FlowRate Them//my DTW2 5.18 1330 18.18 1304 2.41 6.65 + 76.6 10.59 -Gril Peter 12/8/21

Location Erak for THV	Date 12/8/21 7
Project / Client Ext de	
MW-7/EB-01/MW-1	7
to top SPC Da	pH ORD ING DIS
1340 1005 1772 1.63	6.66-722 11.6 4 48
1344 9.91 1.736 1.53	6.66 72.2 12.1 -
1318 9-69 1-327 1-40	6.66 -71.8 12.4 -
1352 980 1-298 138	668-71.4 137 -
1356 1655 1,230 129	6.71 -697 132 764
1357 - Sugle MV-7 Fox	Vacis
- Figushed At M	-7
Rouved is 1501	
(410 - Correct the was a	+ plant FB-C1-120821
By Pouring Cub-Su	poplied on in over
a Dogar mi truted	19 kild co
1415 - Hord to Mar 1	
127 - Athw-1 hegin	Pung, ng
Div 7.49	14-1(27)
DIB	May 5,70 (3
Time Town SPE pa	pit opp Ind Dis
1430 16.27 1.437 11.28	7.32 72.3 18.0 -
1434 1273 1.479 3.15	7.22 38.5 129 -
fluperte= 120	m1/min Mr = 5.4)
1438 16.63 1.464 290	7.22 - 4.7 8:35 8.41
1446 9.63 1.484 2.75	7.22 -11.6 5.32 -
(450 9.46 1.485 258	722 -68 471 836
amtip to	(28/21 Rete in the Rain

Location Frankfat, IN Date 12/8/21 Project / Client Exide Tim Temo SPC DE pt par Time DTL 145 y 9.5 y 1488 241 221 -3 3 214 -1458 964 1496 220 221 7-1 2.82 -1459 - Suple Mar-1 For Vice's - Pinishd At Mar- 1 Rayre 121-25 gal Fial DT we 8.46 1515 - Hand Te Shiply Aren To Decor & Roty 12 Supplies
1000 - S Enging Proged animalistant Completion

Proper worth 1639 - Offgittets Aztol

Location From Kton, FN	Date (2/9/2)	9
Project / Client Eride		

Oper-CEP (Months e) Leaves For	1110
OBO-CEP Casile & Cather Ch	Somety E wat
- City brating 457 556 MP.	3
5 Tulant Nota R	
p A 7.CL 7.64	700
pH400 4.02	2 4.00
SPC(1813) 1.479	
Unp(240) 233 8	
. Po(%) (3377	
- CulibaTing Latelte 20	c 20 me
Staded 1-100	
10.00Th 9.91	4 10.00
- Had To NW-12	
080 - ATMW-12 Bigin P	2011.18
DTW	(va(Q2)
DID	12 m Se7 at 12
Tim Tap 5 16 PG 105 10 1-85 6-761 6-27 7	2H OKP ING DIE
0816 4-85 6-761 0.27 7	20 117.3 142 -
0372 5.51 9-872 9.54 7.	22 16 64
Flow Meta = 18 cm 1/min	
0826 565 0-502 672 7	23 7.4 72 -
0830 9 06 5.825 0.45 7	24 -8.8 68 -
Co 34 9 18 F. 528 F. 49 7	
0810 951 4.537 0,34 7	7.23 -542 796 -
Eris E. Pole	2/9/2 Rete in the Rain

Location Frankfart IV Date 12/972) Project / Client Exide MW-12/MW-11 time trup SPC PO pot GRP Tank DIM 0844 932 0835 0,28 7,23 73.8 74.9 3.89 0848 970 0576 545 7.22 360733 -0852 9.80 0,840 6,43 721 -72-3 705 0856 971 6841 0.42 7.21 -74.3 68.7 0857 - Saple MW-12 For Voe's - Finished AT MW-12 Fix1074=389 - Ronly 6 = 2 agal - Decon Eyniand - Hed Tome-n cayy - At mm-17, Byis Pmpis 1r=1(2") DTW 7.56 10 mp SOT DI ton Time soc be ple our Tous DIL 0957 592 6901 279 7.28 -239 455 -0958 697 6918 1.82 7.73 -19.4 32.4-Flankate = 110 ml/min pth = 3.72 > 1002 783 G924 1. 43 725 88 23.1 -106 8 43 0.938 1.19 7.20 36 148 2.48 1616 853 6-946 1-07 7.14 118 134 -1614 843 6,948 0,96 7.18 200 9.95 -8 48 6,746 0,88 7.17 262 8.28 3.51 1018 1022 8 80 0,444 0,82 7.17 334 7.57 -1026 8,54 0,944 0,77 7.16 367 7.23 7.54 Pring Vette (2/9/4)

Location Pom ktot IN	Date 2/9/2/ 11
Project / Client Exide	
NW-11/mw-g	1
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- Finished Atmm.	- 11
- Nonexda 1501	
- Pecon	
- Hard To Mus 9	
196 - AT MW-7 Regis Page	وبناه
pn 1.89	(V=(0))
PIB	Aupsoint 3
The trap SK Da !	PA ORP IND Alw
11/10 7.41 6.700 0.07	7.17 79.7 19.4 -
1114 9.28 0, Day 0,04.	7.69 875 188 -
12/02 Rato = (30.1)	Im = DTW = 2.98 V
1118 9.65 0,707 0,04	7.97 829 15.8 -
1122 9 61 0.708 0,03	7.47 820 160 372
1126 9.69 0,7070.04	
1732 9.70 0.708 0.02	
(136 9.77 0.767 6.03	7.46 821 19.3
1440 9.820.708 0.03	7.04 81.8 13.2
1941 - Sourle Mr- 4 For 1	100 5
-coloc+ Field Dap	1/1c / e my 90 at 121010
Pring Attac	11/1/2
They's love	14/9/2)
3, 1	Rita Ha Rain

Location Frm ktar) IN Date 12/9/21 Project / Client Exide MW-13 - Font had At mung Removed = 25g 41 Final PT 5.67 - Decon Hondtehw13 1220 - AT MY 13, Ray , Pay 13 DTV 373 1va((2") DTB Pap Set ut () Time tamp SMC DE pH CRP 7500 DTL 1226 7.92 02743 626 7.20 879 123 1230 9.46 1.85 0.05 7.04 66. 4 193 Flunkator NEMIASDY w = 7.89 -> 1234 897 0793 003 7.03 FEO 141 1238 858 0.794 0.05 7.03 593 188 1242 827 0794 405 7.63 543 147 3.84 1246 8080,7930,037,03 54.4 104 1250 795 0792 GOY 704 528 862 1254 798 0.790 6,04 7,04 52,1 55,73,84 1258 805 0.790 Cay 7.04 49.5 86.2 -1302 5.06 6.760 0.03 7.04 - 8.6 520 :-1306 807 0.740 0.04 7.04 536 51.7 3.54 1307 - Smale MW 13 ForVais =1745hod AT AV-13 Romand 20075501, A 10Pm 389 - Decm am & dets (2/9/27

Location from Etar 9, FN	Date (2/9/2) 13							
Project / Client Exide								
aw-14/mh-7	9							
+ Head to nu-14								
1366 ATMW-14 B-15	in Pungoling							
BTW /	W-10212							
070	Page Satar 13'							
Time Top SPC Da	PH PRA Jud DIL							
	24 42.4 (5.6 -							
1342 9.16 0.596 6,06 7	725 78.9 603 -							
Flurates 100	11/2 DTu= 4.78							
1746 9.620,596 0,04 7.	1.22 38.5 54.0 -							
	22 374 500 4.88							
1354 10,81 0,599 0,02 7.	122 14.5 11.8 -							
1358 1,03 0,500 0,03 7	22 31.5 34.1 -							
1402 1125 4,601 4.03 7	22 28.9 26.5 4.97							
1406 1150 9.601 902 7.	22 28 y 28 1 -							
1400 1136 96020,02 7.	2221.3 25.6							
MATT Supe WATTA DE	-V0G5							
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T+ Hend to May-4								
1502- A TMV- Y PC 512								
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piB	Punge Setat 13							
0 00								
Com a That	(2/9/2)							
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Location Fruktot, 2N. Date 12/10/14 14 Location Frak for JIN ... Date (2/9/2) Project / Client & xide Project / Client Ext No. Overcas 7 50. Time Temp SPC De pt ORP Temb DIN 1510 16:91 (-651 541 8:78 425 786 _ OZCC - CERCHOTAGO LOUPS For Sito To Doplay Motopsoplars 0784- CE PORSITO, Organ zig Ushicle (5/4 1/54 1-720 3.31 8.77 396 94.3 cod - Crecking Status of Soul War Locations FlowRate - Dollah 10Ths S. Oy. LI - can Be 5 mp 1,51 1.764 2.78 €77 34.9 96.5 -1518 L2 - Fallothete nell continuor har-1522 1220 1794 2,23 6.76 24.8 84.6 -13 - Can the Saple E 1227 1805 204 6.76 27.5 982 5.49 Ly-Full of hoter 1.876 1.76 6.75 24.8 64.2 -12.33 1.816 1.65 8.75 23.2 58.2 -1534 12 38 LG - Fall of hatpa 1538 1244 1.523 1.50 6.75 19.5 51.8 5.54 Odys - Prepare T- Peper Balay Saplas 1542 1261 1.824 1.33 6.75 16.4 51.2 -41 mm-1, m-1 + ma-9 1546 1264 1-828 1-32 8-75 146 50,7 -0920- Bio Trap Replaced AT MW-1 1550 12.60 1.830 1.30 6.75 126 456 546 1551 - Suple MW- Y For VOC's - Py cuss Regulariant - Finished AT Mu- 4 0945 - Resme Deployment of Bis Traps A emored algallan FireIDIV=5.54 Color Birtray At Mr. - Y Peployod - Peconning & Staging nuterials iese - Buton A+ Mr-9 Deployer 1645 - Materials Styes 1230-12 Test Sorrey Corplet - Simply Protex - CEP Frantary is hell's 1245 = All wells Secure Em C. Prete - Site Secure (2/2/2) am hirits 12/10/21 Rite in the Rain

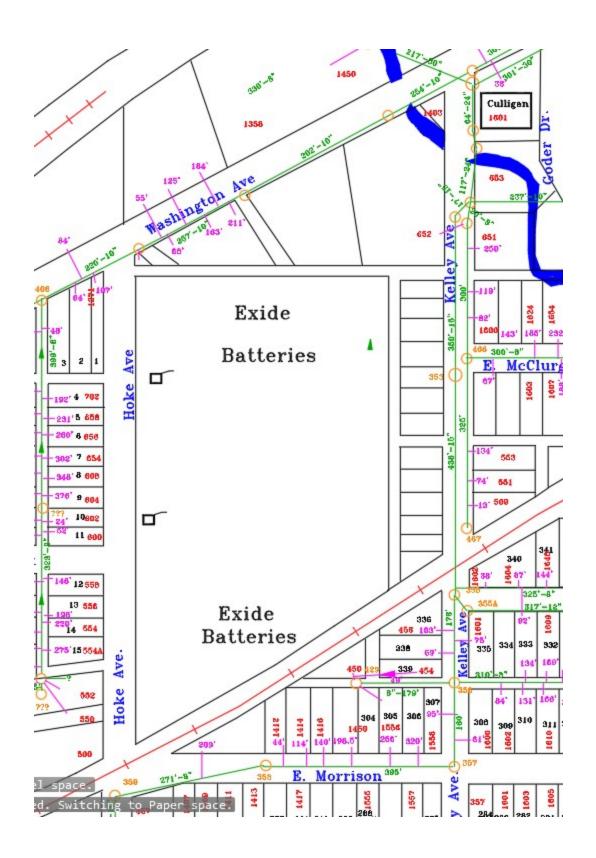
16 Location From Hard IN Date 12/13/21 Project / Client Ettle Project / Client Sanny Vo 0936-(EP(Montrose) Laures For Site To Meet with USEPA+ FDEM Represultations 1112 - Peter Rammansku d Tim De huser (USEPA)

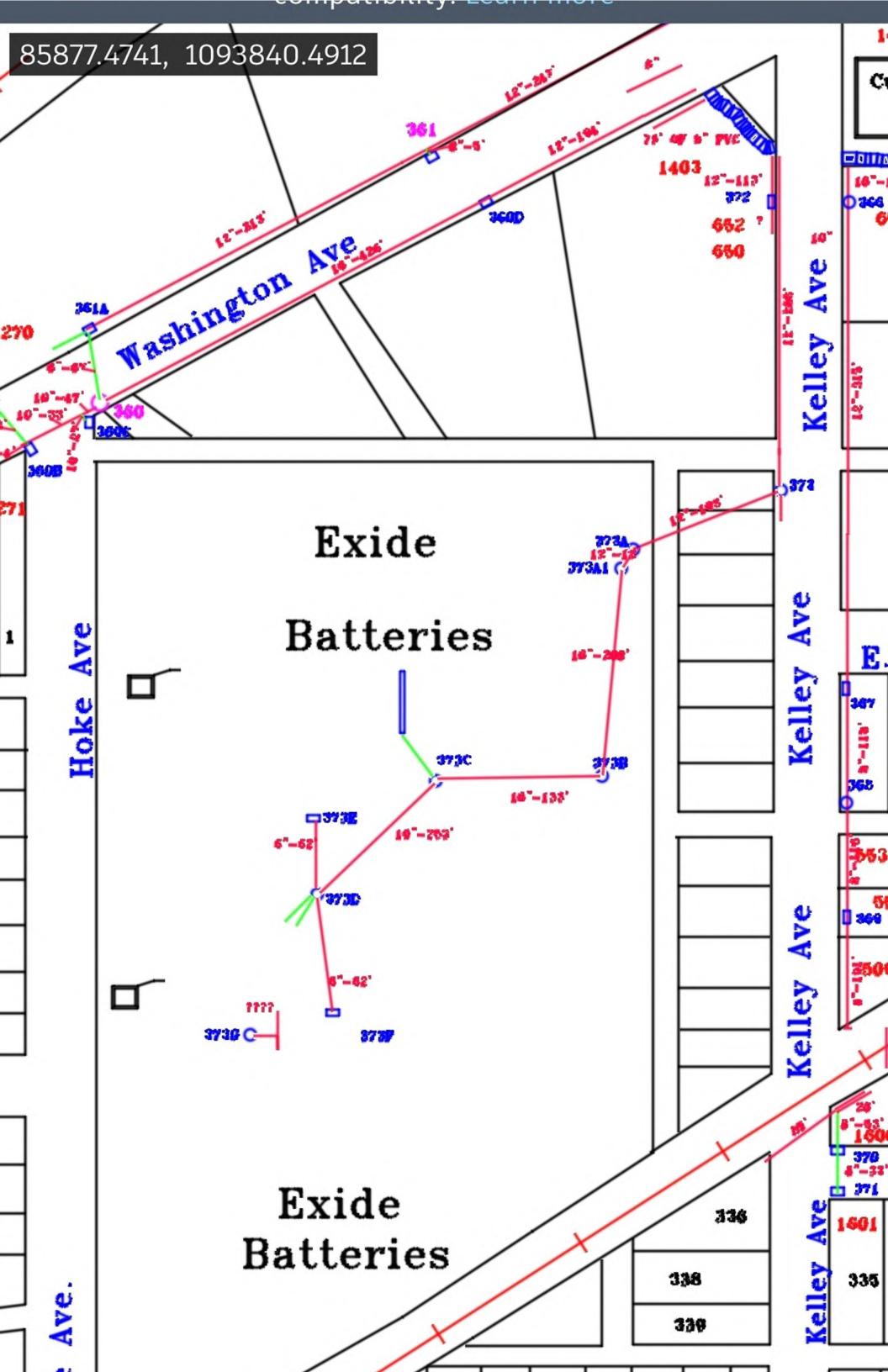
TOEM) Ons! te 115 - Perform Grand orans 1 g St Survey of - SI to & Recat Sings to Low trans
- USERA Crunted Additional Remoderation was 1200 - USEPA + DDEM oftglie - CE Pundates JSD (Marrose) On Prosper. Ing 12/13/2/ Rite in the Rain

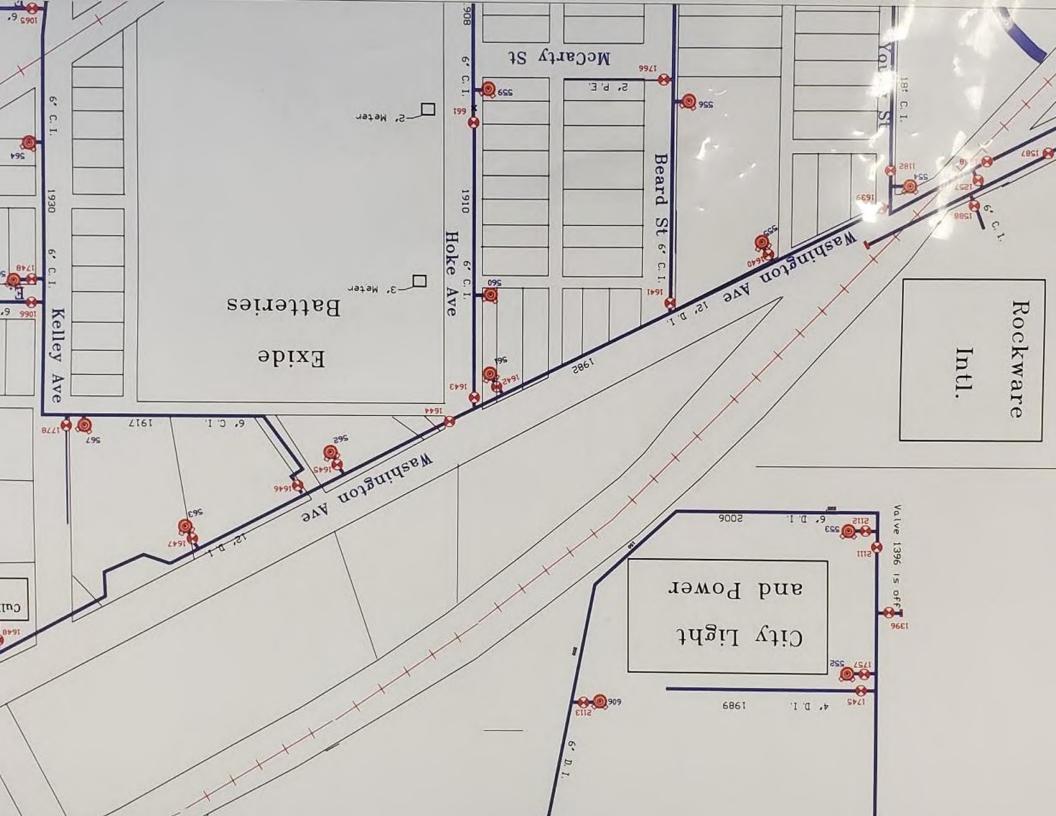


APPENDIX B

Underground Utility Information







MANHOLES SURROUNDING FRANKFORT EXIDE PROPERTY

Notes:

The manholes were assigned identifying letters "A" through "N" for this report, and picture of "Z" outfall to the creek was added; The red letters on the attached figure show these locations.

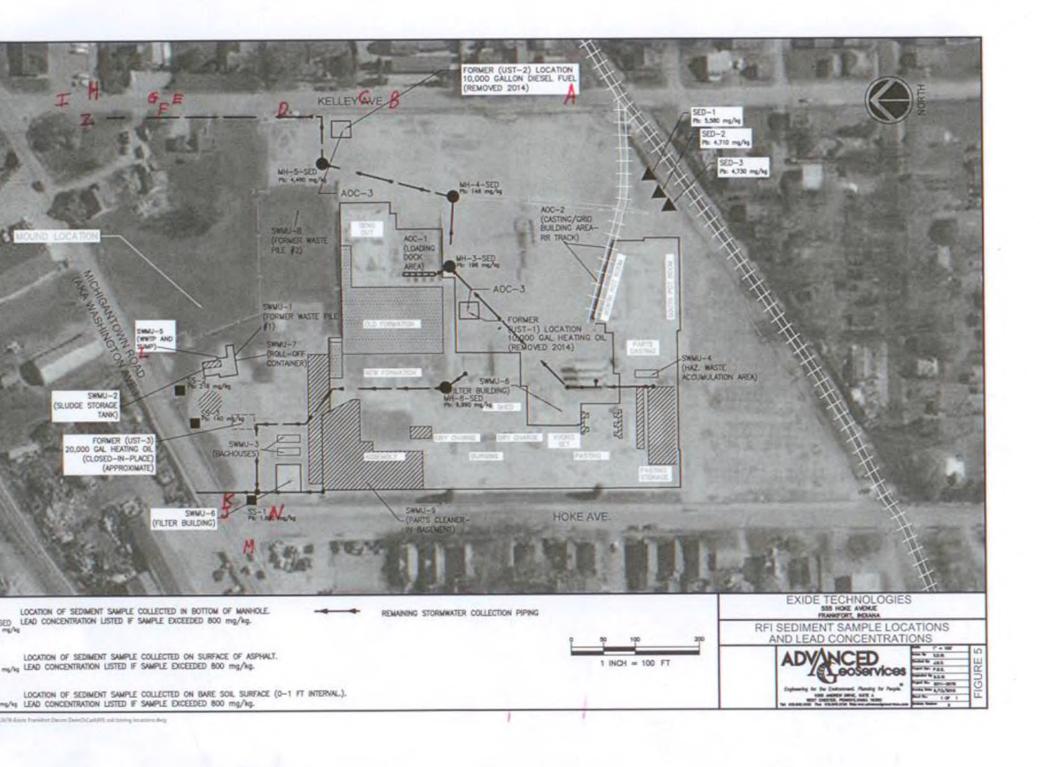
"Distance From Edge Of Street" is always from the Exide direction side of street; the negative numbers represent distance before the street edge (manhole is not in the street)

The start point for "Distance From Start Point" is the railroad tracks for Kelley Avenue manholes; and the inside corner at Hoke and Washington for the Washington and Hoke Avenue manholes

		DIRECTION		OUTLET &		DISTANCE	DISTANCE		
MANHO	LE AVENUE	PICTURES	TYPE	FLOW	MANHOLE INLETS	FROM EDGE	FROM	NOTES	5
IDENTIFI	CATION	ARE FACING		DIRECTION		OF STREET	START POINT	Γ	
Α	Kelley	West	Storm	North	South	14	. 7	'3	1
В	Kelley	East	Sanitary	North	South	11	. 36	55	
C*	Kelley	East	Storm	North	South & East	16	40)4	
D	Kelley	West	Storm	North	South West & South & South East	-3	45	51	2
Е	Kelley	West	Sanitary	North	South & South West	18	60	1	3
F	Kelley	North East East	Sanitary	North East	South	g	61	.1	4
G	Kelley	South	Sanitary	North East	South West West & South West & (2) East	22	62	1	5
Н	Kelley	West	Sanitary	North West	South West	44	. 73	4 6, 7	
1	Kelley	West	Sanitary	North	South East	22	79	1	
J	Washington	North	Sanitary	West	East	3	. 2	25	
K	Washington	South East East	Storm	West	South West, & (2) South South West, & North East	-5	. 3	32	8
L	Washington	North	Sanitary	West	North East & (2) South South West	8	28	34	9
M	Hoke	North East	Storm	East	North West	-1	,	5	
N	Hoke	North	Storm	South East East	North West West	44	. 3	35	10
Z	Kelley	South	Storm	North	South	-14	73	34	11

^{*} Montrose inspection indicates C is a sanitary sewer line Notes:

- 1 No inlet pipe was actually visible in manhole "A", it seemed to be covered with sediment, just a very little flow was seeping from the South side of the manhole.
- 2 The "D" manhole South West inlet comes from the Exide property, and previous imaging revealed this empties into the creek (picture "Z")
- 3 The "E" manhole South West inlet comes from the direction of the 652 Kelley Avenue house.
- 4 The "F" manhole North East outlet pipe is flowing toward the "G" manhole.
- 5 The "G" manhole North East outlet pipe is flowing toward the "H" manhole.
- 6 The "H" manhole is beyond the East side of Kelley Avenue, just North of the creek, beside the Culligan driveway.
- 7 The "H" manhole North West outlet pipe is flowing toward the "I" manhole.
- 8 Exide property stormwater empties into this "K" manhole from the South.
- 9 The Exide Waste Water Treatment Plant previously emptied into this "L" manhole from the South, but these two pipes are now grouted closed.
- 10 Previous imaging of the "N" South East East pipe revealed its making a corner toward the North and then emptying into the "K" manhole.
- 11 Added a photo "Z" of the storm sewer outfall into the creek just West of Kelley Avenue, where Exide stormwater flows from manhole "D".



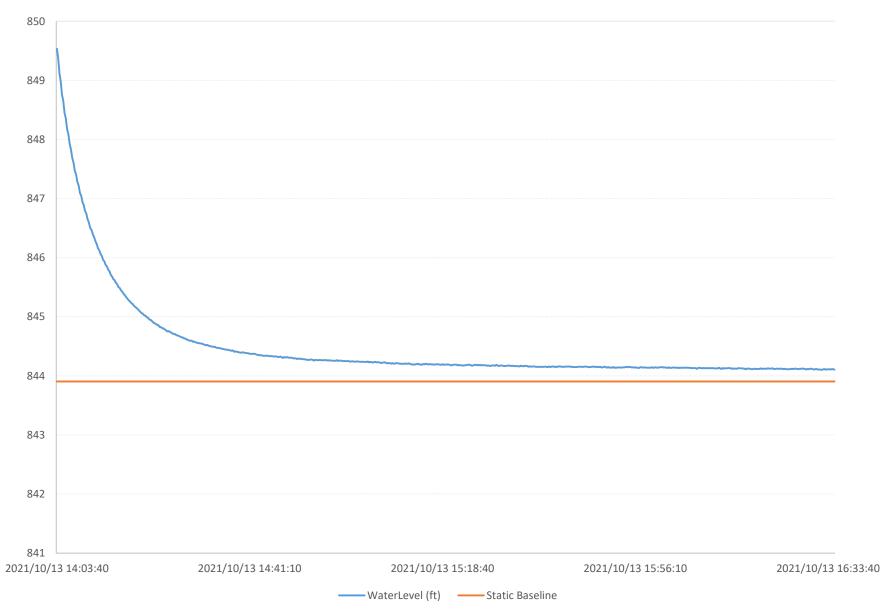


APPENDIX C

Slug Test Data Analysis

WELL ID: EXIDE FRANKFORT Reduced Data Local ID: MW-1 Water Time, **INPUT** Date: 10/13/2021 Entry Hr:Min:Sec Level Construction: Time: 14:04 14:04:20.0 849.53 1 2 Inch 2 Casing dia. (d_c) 14:07:40.0 847.61 Annulus dia. (dw) 8.25 Inch 3 14:10:20.0 846.50 Screen Length (L) 10 Feet 4 14:13:40.0 845.85 5 14:17:20.0 845.39 Depths to: 6 14:20:40.0 845.08 water level (DTW) 9.95 Feet 7 14:23:20.0 844.86 top of screen (TOS) 8 Feet 8 14:26:40.0 844.71 Base of Aquifer (DTB) 14:30:20.0 9 844.59 500 Feet 10 14:33:40.0 844.53 844.45 Annular Fill: 11 14:36:20.0 Base of Aquifer across screen -- Coarse Sand 12 14:39:40.0 844.39 above screen -- Bentonite 13 14:43:20.0 844.35 Adjust slope of line to estimate K 844.33 14 14:46:40.0 1.00 (Aquifer Material -- Till 15 14:49:20.0 844.31 16 14:52:40.0 844.27 COMPUTED 17 14:56:20.0 844.27 8.05 Feet 18 14:59:40.0 844.26 L_{wetted} D= 490.05 Feet 19 15:02:20.0 844.24 H = 8.05 Feet 20 15:05:40.0 844.24 $L/r_w =$ 23.42 21 15:09:20.0 844.20 8.22 Feet 22 15:12:40.0 844.20 $y_{0-DISPLACEMENT} =$ 9.19 Feet 23 844.19 15:15:20.0 $y_{0-SLUG} =$ y/y From look-up table using L/r_w 24 844.19 15:18:40.0 Partial penetrate A = 2.316 25 15:22:20.0 844.18 B = 0.372 26 15:25:40.0 844.19 27 15:28:20.0 844.17 In(Re/rw) = 1 841 28 15:31:40.0 844.17 Re= 2.17 Feet 29 15:35:20.0 844.17 30 844.15 15:38:40.0 Slope = 5.6E-05 log₁₀/sec 31 15:41:20.0 844.16 t_{90%} recovery = 17843 sec 32 15:44:40.0 844.15 15:48:20.0 844.15 Input is consistent. 33 34 15:51:40.0 844.15 35 15:54:20.0 844.14 K = 0.0088 Feet/Day 0.10 00:00 12:00 24:00 36:00 36 15:57:40.0 844.14 37 TIME, Minute: Second 16:01:20.0 844.14 38 16:04:40.0 844.14 39 16:07:20.0 844.13 REMARKS: Bouwer and Rice analysis of slug test, WRR 1976 40 16:10:40.0 844.13 844.13 41 16:14:20.0 Slug test was conducted in surficial aquifer, central Indiana, which is mostly glacial till. 42 16:17:40.0 844.13 43 16:20:20.0 844.12 844.12 44 16:23:40.0 45 16:27:20.0 844.12

FIGURE ___ SLUG TEST DATA MW-1 Exide Technologies Frankfort, Indiana



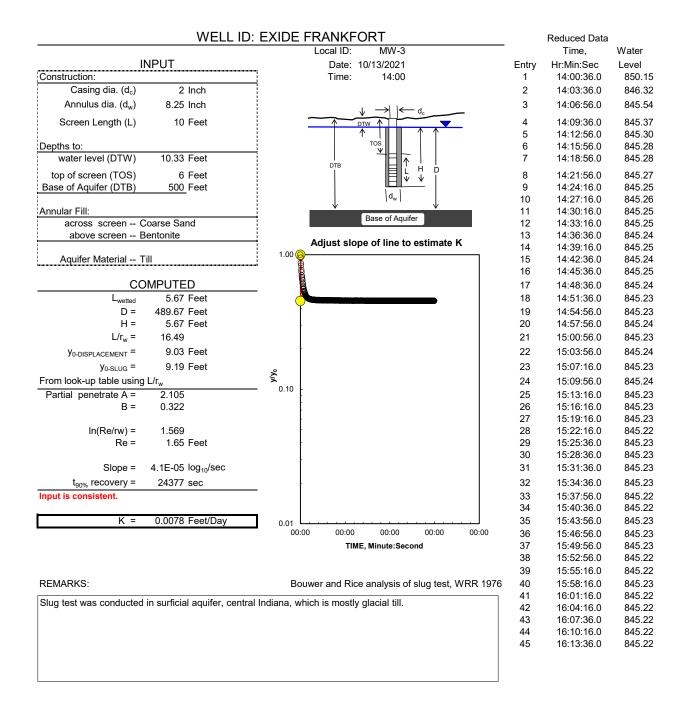
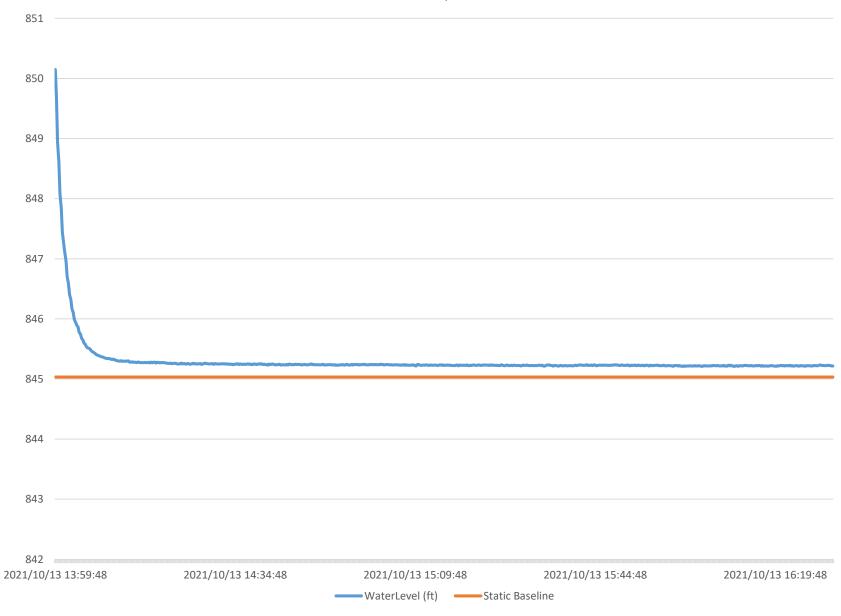


FIGURE ___ SLUG TEST DATA MW-3 Exide Technologies Frankfort, Indiana



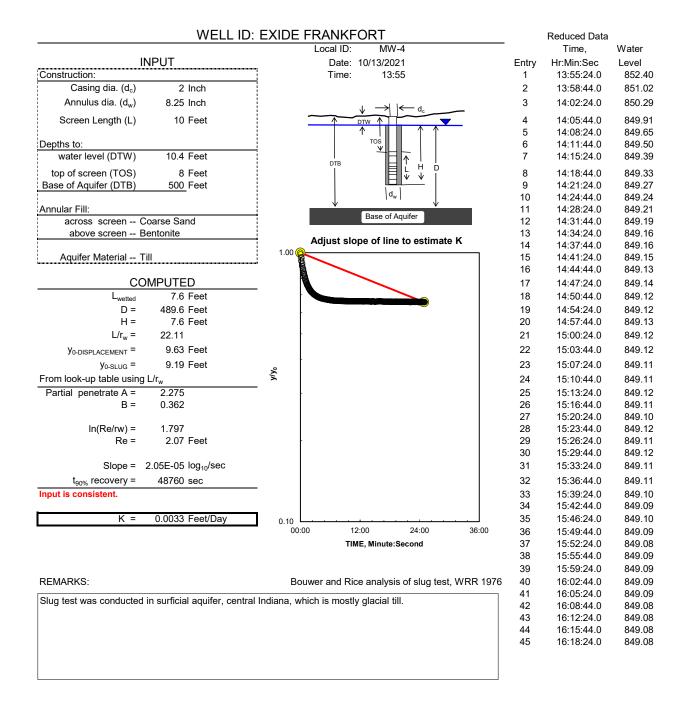
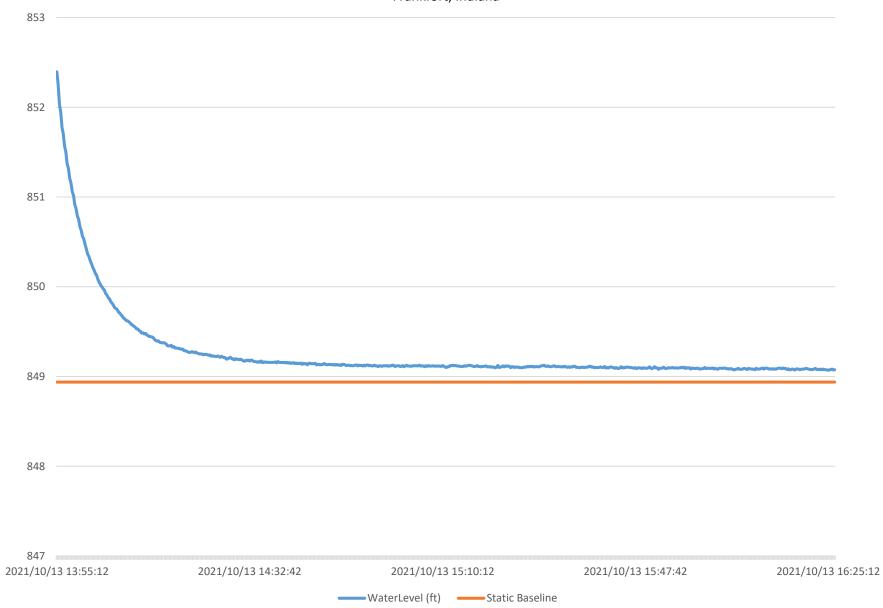


FIGURE ___ SLUG TEST DATA MW-4 Exide Technologies Frankfort, Indiana



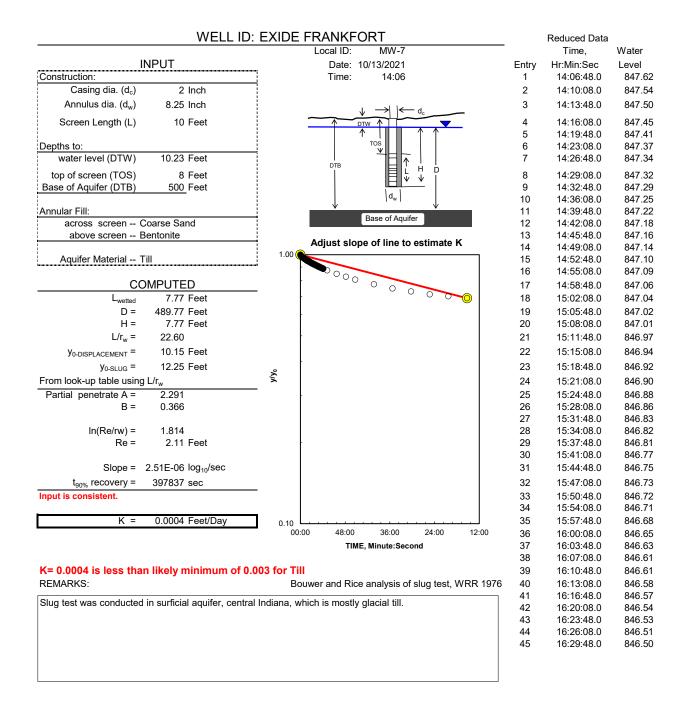
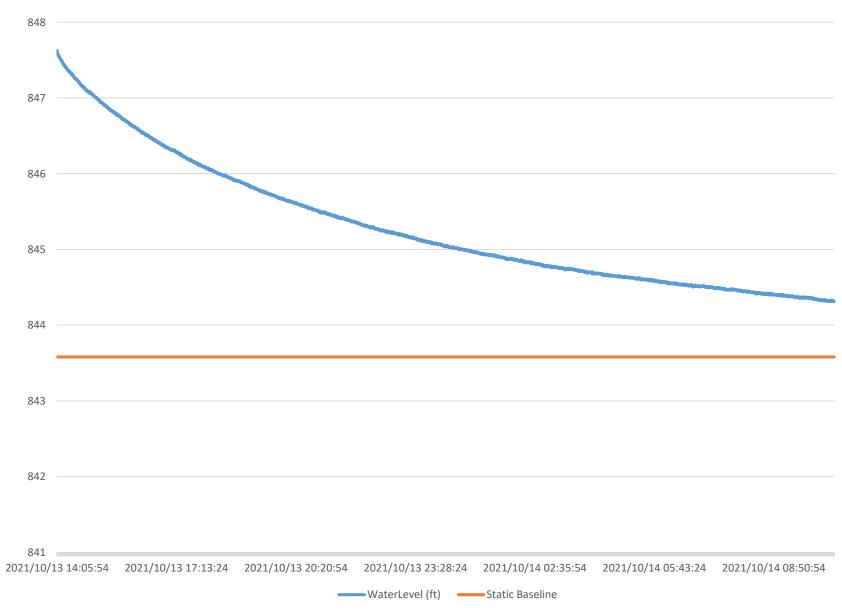


FIGURE ___ SLUG TEST DATA MW-7 Exide Technologies Frankfort, Indiana



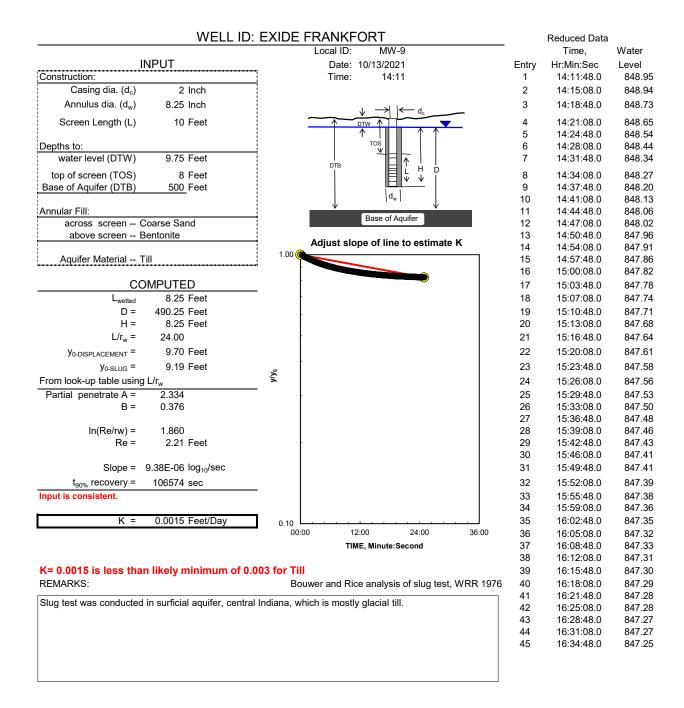
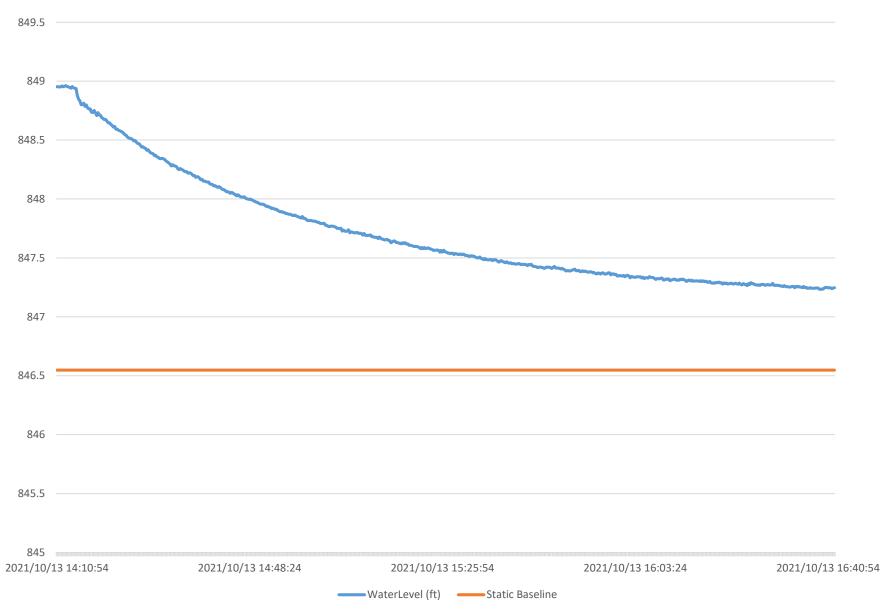


FIGURE ___ SLUG TEST DATA MW-9 Exide Technologies Frankfort, Indiana



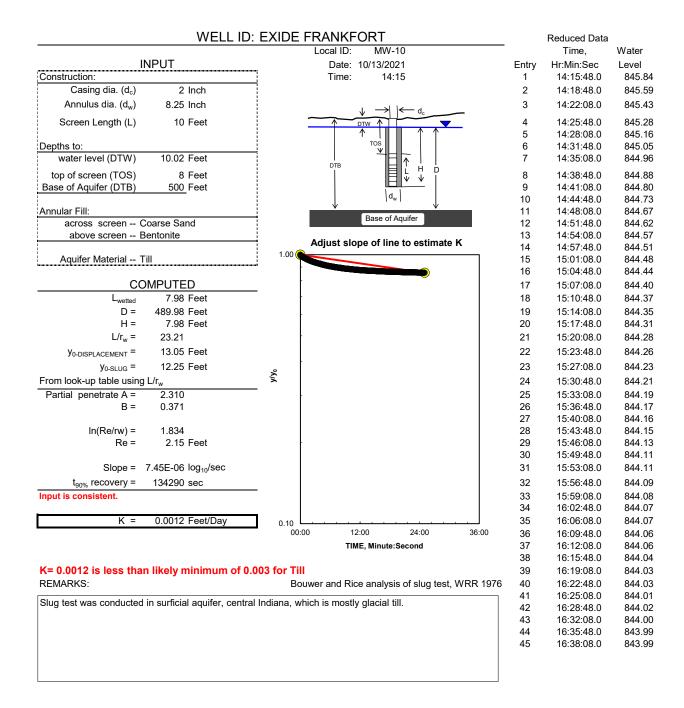
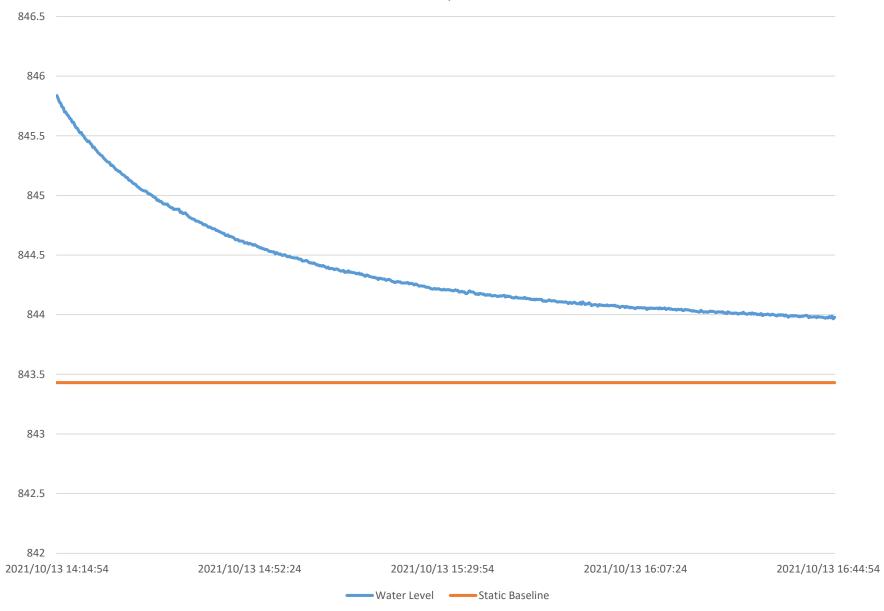


FIGURE ___ SLUG TEST DATA MW-10 Exide Technologies Frankfort, Indiana





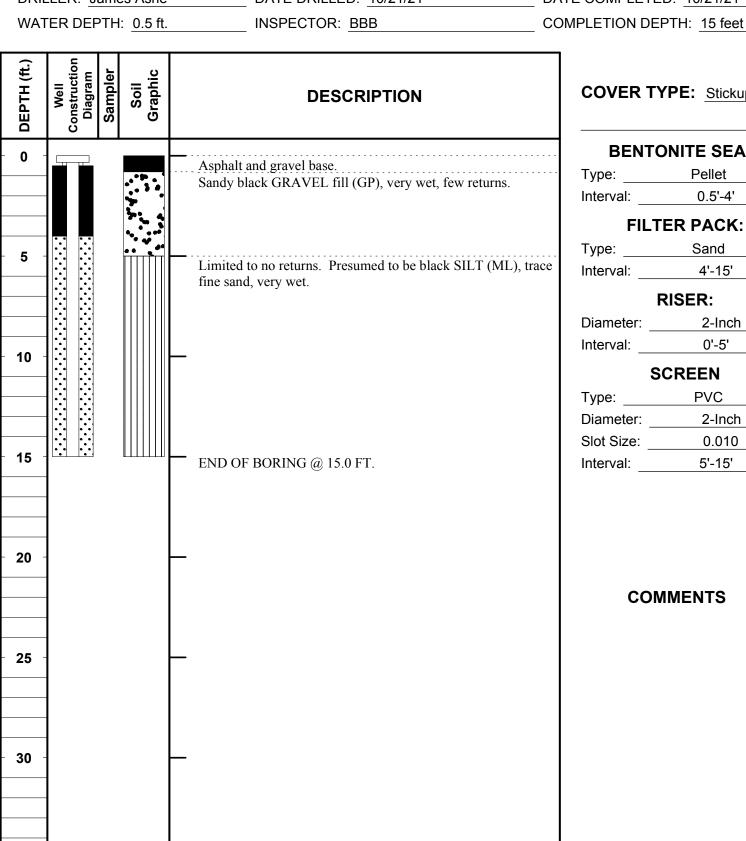
APPENDIX D

New Monitoring Well Logs

MONITORING WELL LOG

WELL NO. MW-14

PROJECT: Exide Trust - Frankfort		PROJECT NO.: 2020-4123
LOCATION: Frankfort, IN		TOIC ELEVATION:
DRILLER: James Ashe	DATE DRILLED: <u>10/21/21</u>	DATE COMPLETED: 10/21/21
WATER DEPTH: 0.5 ft.	INSPECTOR: BBB	COMPLETION DEPTH: 15 feet



COVER TYPE: Stickup

BENTONITE SEAL:

Diameter: 2-Inch

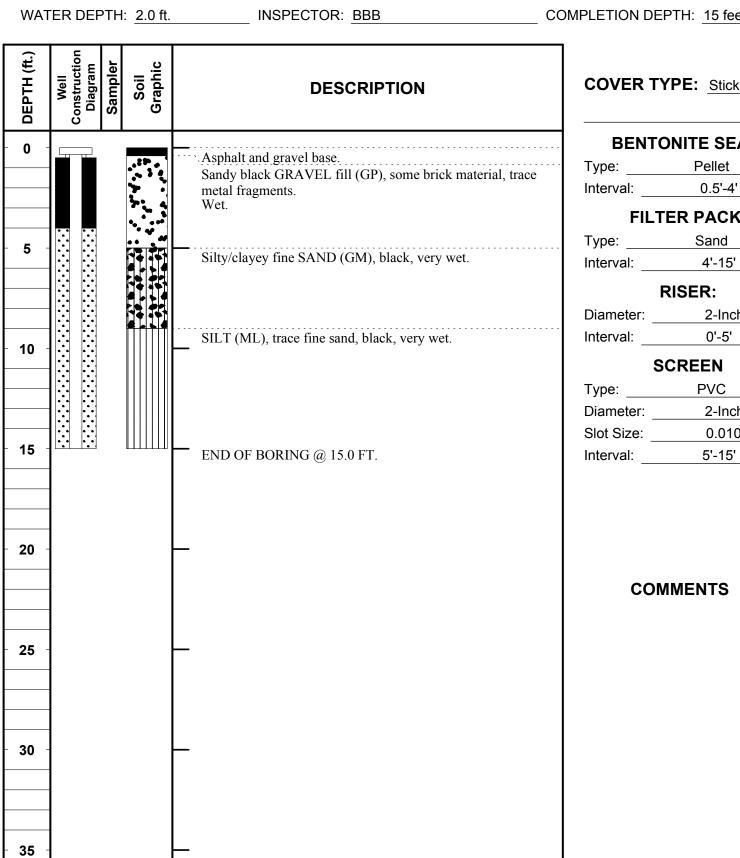
Diameter: 2-Inch Slot Size: 0.010

35

MONITORING WELL LOG

WELL NO. MW-12

PROJECT: Exide Trust - Frankfort		PROJECT NO.: 2020-4123
LOCATION: Frankfort, IN		TOIC ELEVATION:
DRILLER: James Ashe	DATE DRILLED: 10/20/21	DATE COMPLETED: 10/20/21
WATER DEPTH: 2.0 ft.	INSPECTOR: BBB	COMPLETION DEPTH: 15 feet



COVER TYPE: Stickup

BENTONITE SEAL:

Type: Pellet Interval: 0.5'-4'

FILTER PACK:

Type: Sand Interval: 4'-15'

RISER:

Diameter: 2-Inch Interval: 0'-5'

SCREEN

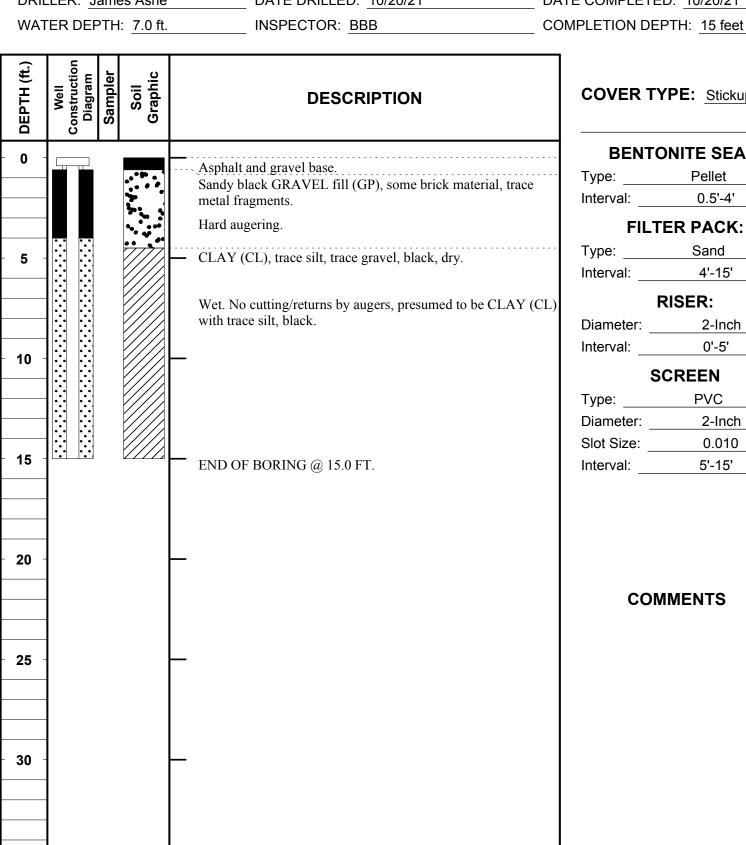
Type: PVC Diameter: 2-Inch Slot Size: 0.010

COMMENTS

MONITORING WELL LOG

WELL NO. MW-13

PROJECT: Exide Trust - Frankfort		PROJECT NO.: 2020-4123
LOCATION: Frankfort, IN		TOIC ELEVATION:
DRILLER: James Ashe	DATE DRILLED: 10/20/21	DATE COMPLETED: 10/20/21
WATER DEPTH: 7.0 ft.	INSPECTOR: BBB	COMPLETION DEPTH: 15 feet



COVER TYPE: Stickup

BENTONITE SEAL:

Diameter: 2-Inch

Diameter: 2-Inch Slot Size: 0.010

35



APPENDIX E

Investigation Derived Waste Records





November 05, 2021

Adam Doubleday Advanced GeoServices Corporation 1055 Andrew Drive, Suite A West Chester, PA 19380

RE: Project: Exide ERT Frankfort Site

Pace Project No.: 50300888

Dear Adam Doubleday:

Enclosed are the analytical results for sample(s) received by the laboratory on October 22, 2021. The results relate only to the samples included in this report. Results reported herein conform to the applicable TNI/NELAC Standards and the laboratory's Quality Manual, where applicable, unless otherwise noted in the body of the report.

The test results provided in this final report were generated by each of the following laboratories within the Pace Network:

- Pace Analytical Services Indianapolis
- Pace Analytical Services Greensburg

If you have any questions concerning this report, please feel free to contact me.

Sincerely,

Brian Hall

brian.hall@pacelabs.com

in Hel

(616)975-4500

Project Manager

Enclosures

cc: Amy Graham, Advanced GeoServices Corporation





CERTIFICATIONS

Project: Exide ERT Frankfort Site

Pace Project No.: 50300888

Pace Analytical Services Pennsylvania

1638 Roseytown Rd Suites 2,3&4, Greensburg, PA 15601

ANAB DOD-ELAP Rad Accreditation #: L2417

Alabama Certification #: 41590 Arizona Certification #: AZ0734

Arkansas Certification

California Certification #: 04222CA Colorado Certification #: PA01547 Connecticut Certification #: PH-0694

Delaware Certification EPA Region 4 DW Rad

Florida/TNI Certification #: E87683 Georgia Certification #: C040 Florida: Cert E871149 SEKS WET

Guam Certification Hawaii Certification Idaho Certification Illinois Certification Indiana Certification Iowa Certification #: 391

Kansas/TNI Certification #: E-10358 Kentucky Certification #: KY90133 KY WW Permit #: KY0098221 KY WW Permit #: KY0000221

Louisiana DHH/TNI Certification #: LA180012 Louisiana DEQ/TNI Certification #: 4086

Maine Certification #: 2017020 Maryland Certification #: 308

Massachusetts Certification #: M-PA1457 Michigan/PADEP Certification #: 9991 Missouri Certification #: 235

Montana Certification #: Cert0082

Nebraska Certification #: NE-OS-29-14 Nevada Certification #: PA014572018-1 New Hampshire/TNI Certification #: 297617 New Jersey/TNI Certification #: PA051 New Mexico Certification #: PA01457 New York/TNI Certification #: 10888 North Carolina Certification #: 42706

Ohio EPA Rad Approval: #41249 Oregon/TNI Certification #: PA200002-010 Pennsylvania/TNI Certification #: 65-00282 Puerto Rico Certification #: PA01457 Rhode Island Certification #: 65-00282

South Dakota Certification
Tennessee Certification #: 02867

North Dakota Certification #: R-190

Texas/TNI Certification #: T104704188-17-3
Utah/TNI Certification #: PA014572017-9
USDA Soil Permit #: P330-17-00091
Vermont Dept. of Health: ID# VT-0282
Virgin Island/PADEP Certification
Virginia/VELAP Certification #: 9526
Washington Certification #: C868
West Virginia DEP Certification #: 143
West Virginia DHHR Certification #: 9964C

Wisconsin Approve List for Rad Wyoming Certification #: 8TMS-L

Pace Analytical Services Indianapolis

7726 Moller Road, Indianapolis, IN 46268

Illinois Accreditation #: 200074

Indiana Drinking Water Laboratory #: C-49-06

Kansas/TNI Certification #: E-10177 Kentucky UST Agency Interest #: 80226 Kentucky WW Laboratory ID #: 98019 Michigan Drinking Water Laboratory #9050

Ohio VAP Certified Laboratory #: CL0065

Oklahoma Laboratory #: 9204 Texas Certification #: T104704355 Wisconsin Laboratory #: 999788130 USDA Soil Permit #: P330-19-00257





SAMPLE SUMMARY

Project: Exide ERT Frankfort Site

Pace Project No.: 50300888

Lab ID	Sample ID	Matrix	Date Collected	Date Received
50300888001	Water-1021	Water	10/22/21 08:20	10/22/21 11:40
50300888002	Soil-1021	Solid	10/22/21 08:50	10/22/21 11:40



SAMPLE ANALYTE COUNT

Project: Exide ERT Frankfort Site

Pace Project No.: 50300888

Lab ID	Sample ID	Method	Analysts	Analytes Reported	Laboratory
50300888001	Water-1021	EPA 6010	JPK	7	PASI-I
		EPA 6010	RAM	7	PASI-I
		EPA 7470	ILP	1	PASI-I
		EPA 7470	DDA	1	PASI-I
		EPA 8270	JCM	18	PASI-I
		EPA 8270 by SIM	GRM	19	PASI-I
		EPA 8270	GRM	53	PASI-I
		EPA 5030/8260	TLS1	13	PASI-I
		EPA 8260	TLS1	53	PASI-I
		EPA 1020B	SWJ	1	PASI-I
		SM 4500-H+B	TKG	1	PASI-I
		EPA 9014	NAH	1	PASI-PA
		SM 4500-S2-F-2011	NAH	1	PASI-PA
50300888002	Soil-1021	EPA 6010	JDG	7	PASI-I
		EPA 6010	JPK	7	PASI-I
		EPA 7470	ILP	1	PASI-I
		EPA 7471	ILP	1	PASI-I
		EPA 8270	JCM	73	PASI-I
		EPA 8270	JCM	18	PASI-I
		EPA 5030/8260	TLS1	13	PASI-I
		EPA 8260	AEP	54	PASI-I
		SM 2540G	ADT	1	PASI-I
		1030	SWJ	1	PASI-I
		EPA 9045	SWJ	1	PASI-I
		EPA 9014	NAH	1	PASI-PA
		SM 4500-S2-F-2011	NAH	1	PASI-PA

PASI-I = Pace Analytical Services - Indianapolis PASI-PA = Pace Analytical Services - Greensburg



SUMMARY OF DETECTION

Project: Exide ERT Frankfort Site

Pace Project No.: 50300888

Lab Sample ID	Client Sample ID					
Method	Parameters	Result	Units	Report Limit	Analyzed	Qualifiers
0300888001	Water-1021					
EPA 6010	Arsenic	3.7J	ug/L	10.0	10/27/21 17:26	
EPA 6010	Barium	74.1	ug/L	10.0	10/27/21 17:26	
EPA 6010	Chromium	28.8	ug/L	10.0	10/27/21 17:26	
EPA 8270	Phenol	5.9J	ug/L	10.0	11/01/21 20:37	
EPA 8260	Acetone	159	ug/L	100	11/01/21 14:54	
EPA 8260	cis-1,2-Dichloroethene	7.8	ug/L	5.0	11/01/21 14:54	
EPA 8260	Trichloroethene	0.61J	ug/L	5.0	11/01/21 14:54	
EPA 8260	Vinyl chloride	1.9J	ug/L	2.0	11/01/21 14:54	
EPA 1020B	Flashpoint	>200.0	deg F		10/26/21 13:17	N2
SM 4500-H+B	pH at 25 Degrees C	8.7	Std. Units	0.10	10/30/21 09:37	H3
0300888002	Soil-1021					
EPA 6010	Arsenic	7.6	mg/kg	1.2	10/27/21 12:52	
EPA 6010	Barium	76.5	mg/kg	1.2	10/27/21 12:52	
EPA 6010	Cadmium	0.62	mg/kg	0.60	10/27/21 12:52	
EPA 6010	Chromium	14.4	mg/kg	1.2	10/27/21 12:52	
EPA 6010	Lead	47.7	mg/kg	1.2	10/27/21 12:52	
EPA 6010	Barium	0.82J	mg/L	5.0	11/04/21 00:14	
EPA 7471	Mercury	0.037J	mg/kg	0.26	11/01/21 10:00	
EPA 8260	Acetone	0.010J	mg/kg	0.096	11/04/21 21:48	
EPA 8260	Chloroform	0.00096J	mg/kg	0.0048	11/04/21 21:48	В
EPA 8260	cis-1,2-Dichloroethene	0.0037J	mg/kg	0.0048	11/04/21 21:48	
EPA 8260	Trichloroethene	0.00092J	mg/kg	0.0048	11/04/21 21:48	
EPA 8260	Vinyl chloride	0.00064J	mg/kg	0.0048	11/04/21 21:48	
SM 2540G	Percent Moisture	22.2	%	0.10	10/25/21 11:52	N2
030	Ignitability, non-metallic	<2.2 mm/sec	mm/sec	2.2	10/27/21 10:05	N2
EPA 9045	pH at 25 Degrees C	6.8	Std. Units	0.10	10/29/21 13:07	H3



Project: Exide ERT Frankfort Site

Pace Project No.: 50300888

Method: EPA 6010
Description: 6010 MET ICP

Client: Advanced GeoServices Corporation

Date: November 05, 2021

General Information:

2 samples were analyzed for EPA 6010 by Pace Analytical Services Indianapolis. All samples were received in acceptable condition with any exceptions noted below or on the chain-of custody and/or the sample condition upon receipt form (SCUR) attached at the end of this report.

Hold Time:

The samples were analyzed within the method required hold times with any exceptions noted below.

Sample Preparation:

The samples were prepared in accordance with EPA 3010 with any exceptions noted below.

The samples were prepared in accordance with EPA 3050 with any exceptions noted below.

Initial Calibrations (including MS Tune as applicable):

All criteria were within method requirements with any exceptions noted below.

Continuing Calibration:

All criteria were within method requirements with any exceptions noted below.

Method Blank:

All analytes were below the report limit in the method blank, where applicable, with any exceptions noted below.

Laboratory Control Spike:

All laboratory control spike compounds were within QC limits with any exceptions noted below.

Matrix Spikes:

All percent recoveries and relative percent differences (RPDs) were within acceptance criteria with any exceptions noted below.



Project: Exide ERT Frankfort Site

Pace Project No.: 50300888

Method: EPA 6010

Description: 6010 MET ICP, TCLP

Client: Advanced GeoServices Corporation

Date: November 05, 2021

General Information:

2 samples were analyzed for EPA 6010 by Pace Analytical Services Indianapolis. All samples were received in acceptable condition with any exceptions noted below or on the chain-of custody and/or the sample condition upon receipt form (SCUR) attached at the end of this report.

Hold Time:

The samples were analyzed within the method required hold times with any exceptions noted below.

Sample Preparation:

The samples were prepared in accordance with EPA 3010 with any exceptions noted below.

Initial Calibrations (including MS Tune as applicable):

All criteria were within method requirements with any exceptions noted below.

Continuing Calibration:

All criteria were within method requirements with any exceptions noted below.

Method Blank:

All analytes were below the report limit in the method blank, where applicable, with any exceptions noted below.

Laboratory Control Spike:

All laboratory control spike compounds were within QC limits with any exceptions noted below.

Matrix Spikes:

All percent recoveries and relative percent differences (RPDs) were within acceptance criteria with any exceptions noted below.



Project: Exide ERT Frankfort Site

Pace Project No.: 50300888

Method: EPA 7470

Description: 7470 Mercury, TCLP

Client: Advanced GeoServices Corporation

Date: November 05, 2021

General Information:

2 samples were analyzed for EPA 7470 by Pace Analytical Services Indianapolis. All samples were received in acceptable condition with any exceptions noted below or on the chain-of custody and/or the sample condition upon receipt form (SCUR) attached at the end of this report.

Hold Time:

The samples were analyzed within the method required hold times with any exceptions noted below.

Sample Preparation:

The samples were prepared in accordance with EPA 7470 with any exceptions noted below.

Initial Calibrations (including MS Tune as applicable):

All criteria were within method requirements with any exceptions noted below.

Continuing Calibration:

All criteria were within method requirements with any exceptions noted below.

Method Blank:

All analytes were below the report limit in the method blank, where applicable, with any exceptions noted below.

Laboratory Control Spike:

All laboratory control spike compounds were within QC limits with any exceptions noted below.

Matrix Spikes:

All percent recoveries and relative percent differences (RPDs) were within acceptance criteria with any exceptions noted below.

QC Batch: 648297

A matrix spike and/or matrix spike duplicate (MS/MSD) were performed on the following sample(s): 50298356057,50300491001,50300517005,50300888002,50300927001,50301209001,50301220001,50301317001,50301362001,50301380001,50301536002

M0: Matrix spike recovery and/or matrix spike duplicate recovery was outside laboratory control limits.

- MS (Lab ID: 2986883)
 - Mercury

Additional Comments:



Project: Exide ERT Frankfort Site

Pace Project No.: 50300888

Method: EPA 7470
Description: 7470 Mercury

Client: Advanced GeoServices Corporation

Date: November 05, 2021

General Information:

1 sample was analyzed for EPA 7470 by Pace Analytical Services Indianapolis. All samples were received in acceptable condition with any exceptions noted below or on the chain-of custody and/or the sample condition upon receipt form (SCUR) attached at the end of this report.

Hold Time:

The samples were analyzed within the method required hold times with any exceptions noted below.

Sample Preparation:

The samples were prepared in accordance with EPA 7470 with any exceptions noted below.

Initial Calibrations (including MS Tune as applicable):

All criteria were within method requirements with any exceptions noted below.

Continuing Calibration:

All criteria were within method requirements with any exceptions noted below.

Method Blank:

All analytes were below the report limit in the method blank, where applicable, with any exceptions noted below.

Laboratory Control Spike:

All laboratory control spike compounds were within QC limits with any exceptions noted below.

Matrix Spikes:

All percent recoveries and relative percent differences (RPDs) were within acceptance criteria with any exceptions noted below.



Project: Exide ERT Frankfort Site

Pace Project No.: 50300888

Method: EPA 7471
Description: 7471 Mercury

Client: Advanced GeoServices Corporation

Date: November 05, 2021

General Information:

1 sample was analyzed for EPA 7471 by Pace Analytical Services Indianapolis. All samples were received in acceptable condition with any exceptions noted below or on the chain-of custody and/or the sample condition upon receipt form (SCUR) attached at the end of this report.

Hold Time:

The samples were analyzed within the method required hold times with any exceptions noted below.

Sample Preparation:

The samples were prepared in accordance with EPA 7471 with any exceptions noted below.

Initial Calibrations (including MS Tune as applicable):

All criteria were within method requirements with any exceptions noted below.

Continuing Calibration:

All criteria were within method requirements with any exceptions noted below.

Method Blank:

All analytes were below the report limit in the method blank, where applicable, with any exceptions noted below.

Laboratory Control Spike:

All laboratory control spike compounds were within QC limits with any exceptions noted below.

Matrix Spikes:

All percent recoveries and relative percent differences (RPDs) were within acceptance criteria with any exceptions noted below.



Project: Exide ERT Frankfort Site

Pace Project No.: 50300888

Method: EPA 8270

Description: 8270 SVOC SS Soil

Client: Advanced GeoServices Corporation

Date: November 05, 2021

General Information:

1 sample was analyzed for EPA 8270 by Pace Analytical Services Indianapolis. All samples were received in acceptable condition with any exceptions noted below or on the chain-of custody and/or the sample condition upon receipt form (SCUR) attached at the end of this report.

Hold Time:

The samples were analyzed within the method required hold times with any exceptions noted below.

Sample Preparation:

The samples were prepared in accordance with EPA 3546 with any exceptions noted below.

Initial Calibrations (including MS Tune as applicable):

All criteria were within method requirements with any exceptions noted below.

Continuing Calibration:

All criteria were within method requirements with any exceptions noted below.

Internal Standards:

All internal standards were within QC limits with any exceptions noted below.

Surrogates:

All surrogates were within QC limits with any exceptions noted below.

Method Blank:

All analytes were below the report limit in the method blank, where applicable, with any exceptions noted below.

Laboratory Control Spike:

All laboratory control spike compounds were within QC limits with any exceptions noted below.

Matrix Spikes:

All percent recoveries and relative percent differences (RPDs) were within acceptance criteria with any exceptions noted below.

Additional Comments:

Analyte Comments:

QC Batch: 647839

N2: The lab does not hold NELAC/TNI accreditation for this parameter but other accreditations/certifications may apply. A complete list of accreditations/certifications is available upon request.

- BLANK (Lab ID: 2985029)
 - Atrazine
 - Biphenyl (Diphenyl)
 - Benzaldehyde
 - Caprolactam



Project: Exide ERT Frankfort Site

Pace Project No.: 50300888

Method: EPA 8270

Description: 8270 SVOC SS Soil

Client: Advanced GeoServices Corporation

Date: November 05, 2021

Analyte Comments: QC Batch: 647839

N2: The lab does not hold NELAC/TNI accreditation for this parameter but other accreditations/certifications may apply. A complete list of accreditations/certifications is available upon request.

- Soil-1021 (Lab ID: 50300888002)
 - Atrazine
 - Biphenyl (Diphenyl)
 - Benzaldehyde
 - Caprolactam



Project: Exide ERT Frankfort Site

Pace Project No.: 50300888

Method: EPA 8270

Description: 8270 MSSV TCLP Sep Funnel **Client:** Advanced GeoServices Corporation

Date: November 05, 2021

General Information:

2 samples were analyzed for EPA 8270 by Pace Analytical Services Indianapolis. All samples were received in acceptable condition with any exceptions noted below or on the chain-of custody and/or the sample condition upon receipt form (SCUR) attached at the end of this report.

Hold Time:

The samples were analyzed within the method required hold times with any exceptions noted below.

Sample Preparation:

The samples were prepared in accordance with EPA 3510 with any exceptions noted below.

Initial Calibrations (including MS Tune as applicable):

All criteria were within method requirements with any exceptions noted below.

Continuing Calibration:

All criteria were within method requirements with any exceptions noted below.

Internal Standards:

All internal standards were within QC limits with any exceptions noted below.

Surrogates:

All surrogates were within QC limits with any exceptions noted below.

Method Blank:

All analytes were below the report limit in the method blank, where applicable, with any exceptions noted below.

Laboratory Control Spike:

All laboratory control spike compounds were within QC limits with any exceptions noted below.

Matrix Spikes:

All percent recoveries and relative percent differences (RPDs) were within acceptance criteria with any exceptions noted below.



Project: Exide ERT Frankfort Site

Pace Project No.: 50300888

Method: EPA 8270 by SIM

Description: 8270 100mL Combo RV

Client: Advanced GeoServices Corporation

Date: November 05, 2021

General Information:

1 sample was analyzed for EPA 8270 by SIM by Pace Analytical Services Indianapolis. All samples were received in acceptable condition with any exceptions noted below or on the chain-of custody and/or the sample condition upon receipt form (SCUR) attached at the end of this report.

Hold Time:

The samples were analyzed within the method required hold times with any exceptions noted below.

H2: Extraction or preparation conducted outside EPA method holding time.

• Water-1021 (Lab ID: 50300888001)

H3: Sample was received or analysis requested beyond the recognized method holding time.

• Water-1021 (Lab ID: 50300888001)

Sample Preparation:

The samples were prepared in accordance with EPA 3510 with any exceptions noted below.

Initial Calibrations (including MS Tune as applicable):

All criteria were within method requirements with any exceptions noted below.

Continuing Calibration:

All criteria were within method requirements with any exceptions noted below.

Internal Standards:

All internal standards were within QC limits with any exceptions noted below.

Surrogates:

All surrogates were within QC limits with any exceptions noted below.

Method Blank:

All analytes were below the report limit in the method blank, where applicable, with any exceptions noted below.

Laboratory Control Spike:

All laboratory control spike compounds were within QC limits with any exceptions noted below.

QC Batch: 647400

L2: Analyte recovery in the laboratory control sample (LCS) was below QC limits. Results for this analyte in associated samples may be biased low.

- LCS (Lab ID: 2982638)
 - 2-Methylnaphthalene
 - Acenaphthene
 - Acenaphthylene
 - Fluorene
 - Naphthalene
 - Phenanthrene



Project: Exide ERT Frankfort Site

Pace Project No.: 50300888

Method: EPA 8270 by SIM

Description: 8270 100mL Combo RV

Client: Advanced GeoServices Corporation

Date: November 05, 2021

Matrix Spikes:

All percent recoveries and relative percent differences (RPDs) were within acceptance criteria with any exceptions noted below.



Project: Exide ERT Frankfort Site

Pace Project No.: 50300888

Method: EPA 8270

Description: 8270 SVOC Combo Water

Client: Advanced GeoServices Corporation

Date: November 05, 2021

General Information:

1 sample was analyzed for EPA 8270 by Pace Analytical Services Indianapolis. All samples were received in acceptable condition with any exceptions noted below or on the chain-of custody and/or the sample condition upon receipt form (SCUR) attached at the end of this report.

Hold Time:

The samples were analyzed within the method required hold times with any exceptions noted below.

Sample Preparation:

The samples were prepared in accordance with EPA 3510 with any exceptions noted below.

Initial Calibrations (including MS Tune as applicable):

All criteria were within method requirements with any exceptions noted below.

Continuing Calibration:

All criteria were within method requirements with any exceptions noted below.

Internal Standards:

All internal standards were within QC limits with any exceptions noted below.

Surrogates:

All surrogates were within QC limits with any exceptions noted below.

Method Blank:

All analytes were below the report limit in the method blank, where applicable, with any exceptions noted below.

Laboratory Control Spike:

All laboratory control spike compounds were within QC limits with any exceptions noted below.

Matrix Spikes:

All percent recoveries and relative percent differences (RPDs) were within acceptance criteria with any exceptions noted below.



Project: Exide ERT Frankfort Site

Pace Project No.: 50300888

Method: EPA 5030/8260
Description: 8260 MSV TCLP

Client: Advanced GeoServices Corporation

Date: November 05, 2021

General Information:

2 samples were analyzed for EPA 5030/8260 by Pace Analytical Services Indianapolis. All samples were received in acceptable condition with any exceptions noted below or on the chain-of custody and/or the sample condition upon receipt form (SCUR) attached at the end of this report.

Hold Time:

The samples were analyzed within the method required hold times with any exceptions noted below.

Initial Calibrations (including MS Tune as applicable):

All criteria were within method requirements with any exceptions noted below.

Continuing Calibration:

All criteria were within method requirements with any exceptions noted below.

Internal Standards:

All internal standards were within QC limits with any exceptions noted below.

Surrogates:

All surrogates were within QC limits with any exceptions noted below.

Method Blank:

All analytes were below the report limit in the method blank, where applicable, with any exceptions noted below.

Laboratory Control Spike:

All laboratory control spike compounds were within QC limits with any exceptions noted below.

Matrix Spikes:

All percent recoveries and relative percent differences (RPDs) were within acceptance criteria with any exceptions noted below.



Project: Exide ERT Frankfort Site

Pace Project No.: 50300888

Method: EPA 8260 Description: 8260/5030 MSV

Client: Advanced GeoServices Corporation

Date: November 05, 2021

General Information:

1 sample was analyzed for EPA 8260 by Pace Analytical Services Indianapolis. All samples were received in acceptable condition with any exceptions noted below or on the chain-of custody and/or the sample condition upon receipt form (SCUR) attached at the end of this report.

Hold Time:

The samples were analyzed within the method required hold times with any exceptions noted below.

Initial Calibrations (including MS Tune as applicable):

All criteria were within method requirements with any exceptions noted below.

Continuing Calibration:

All criteria were within method requirements with any exceptions noted below.

Internal Standards:

All internal standards were within QC limits with any exceptions noted below.

Surrogates:

All surrogates were within QC limits with any exceptions noted below.

Method Blank:

All analytes were below the report limit in the method blank, where applicable, with any exceptions noted below.

Laboratory Control Spike:

All laboratory control spike compounds were within QC limits with any exceptions noted below.

Matrix Spikes:

All percent recoveries and relative percent differences (RPDs) were within acceptance criteria with any exceptions noted below.



Project: Exide ERT Frankfort Site

Pace Project No.: 50300888

Method: EPA 8260

Description: 8260 MSV 5035A VOA

Client: Advanced GeoServices Corporation

Date: November 05, 2021

General Information:

1 sample was analyzed for EPA 8260 by Pace Analytical Services Indianapolis. All samples were received in acceptable condition with any exceptions noted below or on the chain-of custody and/or the sample condition upon receipt form (SCUR) attached at the end of this report.

Hold Time:

The samples were analyzed within the method required hold times with any exceptions noted below.

Initial Calibrations (including MS Tune as applicable):

All criteria were within method requirements with any exceptions noted below.

Continuing Calibration:

All criteria were within method requirements with any exceptions noted below.

Internal Standards:

All internal standards were within QC limits with any exceptions noted below.

Surrogates:

All surrogates were within QC limits with any exceptions noted below.

Method Blank:

All analytes were below the report limit in the method blank, where applicable, with any exceptions noted below.

QC Batch: 648672

- B: Analyte was detected in the associated method blank.
 - BLANK for HBN 648672 [MSV/1554 (Lab ID: 2988853)
 - Chloroform

Laboratory Control Spike:

All laboratory control spike compounds were within QC limits with any exceptions noted below.

Matrix Spikes:

All percent recoveries and relative percent differences (RPDs) were within acceptance criteria with any exceptions noted below.

Additional Comments:

Analyte Comments:

QC Batch: 648672

N2: The lab does not hold NELAC/TNI accreditation for this parameter but other accreditations/certifications may apply. A complete list of accreditations/certifications is available upon request.

- BLANK (Lab ID: 2988853)
 - Cyclohexane
 - Methyl acetate
 - Methylcyclohexane



Project: Exide ERT Frankfort Site

Pace Project No.: 50300888

Method: EPA 8260

Description: 8260 MSV 5035A VOA

Client: Advanced GeoServices Corporation

Date: November 05, 2021

Analyte Comments: QC Batch: 648672

N2: The lab does not hold NELAC/TNI accreditation for this parameter but other accreditations/certifications may apply. A complete list of accreditations/certifications is available upon request.

- LCS (Lab ID: 2988854)
 - Cyclohexane
 - Methyl acetate
 - Methylcyclohexane
- Soil-1021 (Lab ID: 50300888002)
 - Cyclohexane
 - Methyl acetate
 - Methylcyclohexane



Project: Exide ERT Frankfort Site

Pace Project No.: 50300888

Method: EPA 1020B

Description: 1020 Flashpoint, Closed Cup **Client:** Advanced GeoServices Corporation

Date: November 05, 2021

General Information:

1 sample was analyzed for EPA 1020B by Pace Analytical Services Indianapolis. All samples were received in acceptable condition with any exceptions noted below or on the chain-of custody and/or the sample condition upon receipt form (SCUR) attached at the end of this report.

Hold Time:

The samples were analyzed within the method required hold times with any exceptions noted below.

Method Blank:

All analytes were below the report limit in the method blank, where applicable, with any exceptions noted below.

Laboratory Control Spike:

All laboratory control spike compounds were within QC limits with any exceptions noted below.

Matrix Spikes:

All percent recoveries and relative percent differences (RPDs) were within acceptance criteria with any exceptions noted below.

Additional Comments:

Analyte Comments:

QC Batch: 646840

N2: The lab does not hold NELAC/TNI accreditation for this parameter but other accreditations/certifications may apply. A complete list of accreditations/certifications is available upon request.

- Water-1021 (Lab ID: 50300888001)
 - Flashpoint



Project: Exide ERT Frankfort Site

Pace Project No.: 50300888

Method: 1030

Description: 1030 Ignitability of Solids

Client: Advanced GeoServices Corporation

Date: November 05, 2021

General Information:

1 sample was analyzed for 1030 by Pace Analytical Services Indianapolis. All samples were received in acceptable condition with any exceptions noted below or on the chain-of custody and/or the sample condition upon receipt form (SCUR) attached at the end of this report.

Hold Time:

The samples were analyzed within the method required hold times with any exceptions noted below.

Method Blank:

All analytes were below the report limit in the method blank, where applicable, with any exceptions noted below.

Laboratory Control Spike:

All laboratory control spike compounds were within QC limits with any exceptions noted below.

Matrix Spikes:

All percent recoveries and relative percent differences (RPDs) were within acceptance criteria with any exceptions noted below.

Additional Comments:

Analyte Comments:

QC Batch: 647110

N2: The lab does not hold NELAC/TNI accreditation for this parameter but other accreditations/certifications may apply. A complete list of accreditations/certifications is available upon request.

- Soil-1021 (Lab ID: 50300888002)
 - Ignitability, non-metallic



Project: Exide ERT Frankfort Site

Pace Project No.: 50300888

Method: SM 4500-H+B

Description: 4500H+ pH, Electrometric

Client: Advanced GeoServices Corporation

Date: November 05, 2021

General Information:

1 sample was analyzed for SM 4500-H+B by Pace Analytical Services Indianapolis. All samples were received in acceptable condition with any exceptions noted below or on the chain-of custody and/or the sample condition upon receipt form (SCUR) attached at the end of this report.

Hold Time:

The samples were analyzed within the method required hold times with any exceptions noted below.

H3: Sample was received or analysis requested beyond the recognized method holding time.

• Water-1021 (Lab ID: 50300888001)

Method Blank:

All analytes were below the report limit in the method blank, where applicable, with any exceptions noted below.

Laboratory Control Spike:

All laboratory control spike compounds were within QC limits with any exceptions noted below.

Matrix Spikes:

All percent recoveries and relative percent differences (RPDs) were within acceptance criteria with any exceptions noted below.

Duplicate Sample:

All duplicate sample results were within method acceptance criteria with any exceptions noted below.

Additional Comments:



Project: Exide ERT Frankfort Site

Pace Project No.: 50300888

Method: EPA 9045 Description: 9045 pH Soil

Client: Advanced GeoServices Corporation

Date: November 05, 2021

General Information:

1 sample was analyzed for EPA 9045 by Pace Analytical Services Indianapolis. All samples were received in acceptable condition with any exceptions noted below or on the chain-of custody and/or the sample condition upon receipt form (SCUR) attached at the end of this report.

Hold Time:

The samples were analyzed within the method required hold times with any exceptions noted below.

H3: Sample was received or analysis requested beyond the recognized method holding time.

• Soil-1021 (Lab ID: 50300888002)

Method Blank:

All analytes were below the report limit in the method blank, where applicable, with any exceptions noted below.

Laboratory Control Spike:

All laboratory control spike compounds were within QC limits with any exceptions noted below.

Matrix Spikes:

All percent recoveries and relative percent differences (RPDs) were within acceptance criteria with any exceptions noted below.

Duplicate Sample:

All duplicate sample results were within method acceptance criteria with any exceptions noted below.

Additional Comments:



Project: Exide ERT Frankfort Site

Pace Project No.: 50300888

Method: EPA 9014

Description: 733C S Reactive Cyanide

Client: Advanced GeoServices Corporation

Date: November 05, 2021

General Information:

2 samples were analyzed for EPA 9014 by Pace Analytical Services Greensburg. All samples were received in acceptable condition with any exceptions noted below or on the chain-of custody and/or the sample condition upon receipt form (SCUR) attached at the end of this report.

Hold Time:

The samples were analyzed within the method required hold times with any exceptions noted below.

Sample Preparation:

The samples were prepared in accordance with SW-846 7.3.3.2 with any exceptions noted below.

Method Blank:

All analytes were below the report limit in the method blank, where applicable, with any exceptions noted below.

Laboratory Control Spike:

All laboratory control spike compounds were within QC limits with any exceptions noted below.

Matrix Spikes:

All percent recoveries and relative percent differences (RPDs) were within acceptance criteria with any exceptions noted below.

Duplicate Sample:

All duplicate sample results were within method acceptance criteria with any exceptions noted below.

Additional Comments:



Project: Exide ERT Frankfort Site

Pace Project No.: 50300888

Method: SM 4500-S2-F-2011

Description: 734S Reactive Sulfide

Client: Advanced GeoServices Corporation

Date: November 05, 2021

General Information:

2 samples were analyzed for SM 4500-S2-F-2011 by Pace Analytical Services Greensburg. All samples were received in acceptable condition with any exceptions noted below or on the chain-of custody and/or the sample condition upon receipt form (SCUR) attached at the end of this report.

Hold Time:

The samples were analyzed within the method required hold times with any exceptions noted below.

Sample Preparation:

The samples were prepared in accordance with SW-846 7.3.4.2 with any exceptions noted below.

Method Blank:

All analytes were below the report limit in the method blank, where applicable, with any exceptions noted below.

Laboratory Control Spike:

All laboratory control spike compounds were within QC limits with any exceptions noted below.

Matrix Spikes:

All percent recoveries and relative percent differences (RPDs) were within acceptance criteria with any exceptions noted below.

Duplicate Sample:

All duplicate sample results were within method acceptance criteria with any exceptions noted below.

Additional Comments:

This data package has been reviewed for quality and completeness and is approved for release.



Project: Exide ERT Frankfort Site

Pace Project No.: 50300888

Date: 11/05/2021 04:07 PM

Sample: Water-1021	Lab ID: 50300888001 Collected: 10/22/21 08:20 Received: 10/22/21 11:40 Matrix: Water									
Parameters	Results	Units	PQL	MDL	DF	Prepared	Analyzed	CAS No.	Qua	
6010 MET ICP, TCLP	Analytical	Method: EPA 6	010 Prepa	ration Meth	od: EPA	A 3010				
	Leachate	Method/Date: E	PA 1311; 1	1/01/21 20:	50 Initi	al pH: 7.39; Final	pH: 7.39			
	Pace Anal	ytical Services	- Indianapo	lis						
Arsenic	<0.050	mg/L	0.10	0.050	1	11/03/21 13:20	11/04/21 00:05	7440-38-2		
Barium	<0.25	mg/L	5.0	0.25	1	11/03/21 13:20	11/04/21 00:05	7440-39-3		
Cadmium	< 0.025	mg/L	0.050	0.025	1	11/03/21 13:20	11/04/21 00:05	7440-43-9		
Chromium	< 0.052	mg/L	0.10	0.052	1	11/03/21 13:20	11/04/21 00:05	7440-47-3		
.ead	< 0.050	mg/L	0.10	0.050	1	11/03/21 13:20	11/04/21 00:05	7439-92-1		
Selenium	< 0.050	mg/L	0.10	0.050	1	11/03/21 13:20	11/04/21 00:05	7782-49-2		
Silver	<0.050	mg/L	0.10	0.050	1	11/03/21 13:20	11/04/21 00:05	7440-22-4		
010 MET ICP	Analytical	Method: EPA 6	010 Prepa	ration Meth	od: EPA	A 3010				
	-	ytical Services								
Arsenic	3.7J	ug/L	10.0	2.6	1	10/26/21 13:30	10/27/21 17:26	7440-38-2		
Barium	74.1	ug/L	10.0	0.79	1	10/26/21 13:30	10/27/21 17:26	7440-39-3		
Cadmium	<0.41	ug/L	2.0	0.41	1	10/26/21 13:30	10/27/21 17:26	7440-43-9		
Chromium	28.8	ug/L	10.0	1.9	1	10/26/21 13:30	10/27/21 17:26	7440-47-3		
ead	<3.5	ug/L	10.0	3.5	1	10/26/21 13:30	10/27/21 17:26	7439-92-1		
Selenium	<4.5	ug/L	10.0	4.5	1	10/26/21 13:30	10/27/21 17:26	7782-49-2		
Silver	<1.4	ug/L	10.0	1.4	1	10/26/21 13:30	10/27/21 17:26	7440-22-4		
7470 Mercury, TCLP	Analytical	Method: EPA 7	470 Prena	ration Meth	od: FP/	\ 7470				
470 Mercury, TCLI	•						nU- 7 20			
					30 IIIII	al pH: 7.39; Final	pr 1. 7.39			
	Pace Anai	ytical Services	- indianapo	IIS						
Mercury	<0.0010	mg/L	0.0020	0.0010	1	11/03/21 10:41	11/04/21 11:35	7439-97-6		
7470 Mercury	Analytical	Method: EPA 7	470 Prepa	ration Meth	od: EPA	A 7470				
•	Pace Anal	ytical Services	- Indianapo	lis						
Mercury	<0.085	ug/L	2.0	0.085	1	11/04/21 10:03	11/04/21 16:43	7439-97-6		
3270 MSSV TCLP Sep Funnel	Analytical	Method: EPA 8	270 Prepa	ration Meth	od: EPA	A 3510				
	•		•			al pH: 7.39; Final	nH: 7.39			
		ytical Services			00 11111	ai pi ii 7 .00, 1 ii ai	pr 7.00			
,4-Dichlorobenzene	<0.050	mg/L	0.10	0.050	1	11/02/21 19:55	11/04/21 00:06	106-46-7		
2.4-Dinitrotoluene	<0.050	mg/L	0.10	0.050	1		11/04/21 00:06			
lexachloro-1,3-butadiene	<0.050	mg/L	0.10	0.050	1	11/02/21 19:55	11/04/21 00:06			
lexachlorobenzene	<0.050 <0.050	mg/L	0.10	0.050	1	11/02/21 19:55	11/04/21 00:06			
lexachloroethane	<0.050 <0.050	mg/L	0.10	0.050	1	11/02/21 19:55	11/04/21 00:06			
-Methylphenol(o-Cresol)	<0.050 <0.050	mg/L	0.10	0.050	1	11/02/21 19:55	11/04/21 00:06			
8&4-Methylphenol(m&p Cresol)	<0.10	mg/L	0.10	0.030	1	11/02/21 19:55	11/04/21 00:06	JJ T U-1		
litrobenzene	<0.10 <0.050	-	0.20	0.10	1	11/02/21 19:55	11/04/21 00:06	08-05 2		
	<0.050 <0.25	mg/L mg/l	0.10	0.050			11/04/21 00:06			
Pentachlorophenol Pyridine	<0.25 <0.10	mg/L	0.50		1	11/02/21 19:55				
•		mg/L		0.10	1	11/02/21 19:55	11/04/21 00:06			
2,4,5-Trichlorophenol	<0.050	mg/L	0.50	0.050	1	11/02/21 19:55	11/04/21 00:06			
2,4,6-Trichlorophenol	<0.050	mg/L	0.10	0.050	1	11/02/21 19:55	11/04/21 00:06	08-00-2		



Project: Exide ERT Frankfort Site

Pace Project No.: 50300888

Date: 11/05/2021 04:07 PM

Sample: Water-1021	Lab ID: 50300888001 Collected: 10/22/21 08:20 Received: 10/22/21 11:40 Matrix: Water									
Parameters	Results	Units	PQL	MDL	DF	Prepared	Analyzed	CAS No.	Qua	
8270 MSSV TCLP Sep Funnel	•	Method: EPA 8	•							
			•		50 Initi	ial pH: 7.39; Final	pH: 7.39			
	Pace Ana	lytical Services	- Indianapo	olis						
Surrogates	74	0/	10 115			44/00/04 40 55	44/04/04 00 00	4405.00.0		
Nitrobenzene-d5 (S)	71	%. %.	40-115		1	11/02/21 19:55	11/04/21 00:06			
2-Fluorobiphenyl (S)	61 81	%. %.	35-102 42-156		1 1	11/02/21 19:55 11/02/21 19:55	11/04/21 00:06 11/04/21 00:06			
o-Terphenyl-d14 (S)	30	%. %.	15-48		1	11/02/21 19:55	11/04/21 00:06			
Phenol-d5 (S)	43	%. %.	21-74		1	11/02/21 19:55	11/04/21 00:06			
2-Fluorophenol (S)	43 82				1					
2,4,6-Tribromophenol (S)	02	%.	47-127		!	11/02/21 19:55	11/04/21 00:06	116-79-6		
8270 100mL Combo RV	Analytical	Method: EPA 8	270 by SIM	l Preparati	on Meth	nod: EPA 3510				
	Pace Ana	lytical Services	- Indianapo	olis						
Acenaphthene	<0.015	ug/L	1.0	0.015	1	10/28/21 10:39	11/01/21 18:58	83-32-9	L2	
Acenaphthene	<0.014	ug/L	0.95	0.014	1	11/04/21 19:31	11/05/21 14:19		H2	
Acenaphthylene	< 0.013	ug/L	1.0	0.013	1	10/28/21 10:39	11/01/21 18:58	208-96-8	L2	
Acenaphthylene	<0.012	ug/L	0.95	0.012	1	11/04/21 19:31	11/05/21 14:19	208-96-8	H2	
Anthracene	<0.012	ug/L	0.10	0.012	1	10/28/21 10:39	11/01/21 18:58	120-12-7		
Anthracene	<0.012	ug/L	0.095	0.012	1	11/04/21 19:31	11/05/21 14:19	120-12-7	H2	
Benzo(a)anthracene	<0.027	ug/L	0.10	0.027	1	10/28/21 10:39	11/01/21 18:58	56-55-3		
Benzo(a)anthracene	<0.026	ug/L	0.095	0.026	1	11/04/21 19:31	11/05/21 14:19	56-55-3	H2	
Benzo(a)pyrene	<0.026	ug/L	0.10	0.026	1	10/28/21 10:39	11/01/21 18:58	50-32-8		
Benzo(a)pyrene	<0.025	ug/L	0.095	0.025	1	11/04/21 19:31	11/05/21 14:19	50-32-8	H2	
Benzo(b)fluoranthene	<0.031	ug/L	0.10	0.031	1	10/28/21 10:39	11/01/21 18:58	205-99-2		
Benzo(b)fluoranthene	< 0.030	ug/L	0.095	0.030	1	11/04/21 19:31	11/05/21 14:19	205-99-2	H2	
Benzo(g,h,i)perylene	<0.024	ug/L	0.10	0.024	1	10/28/21 10:39	11/01/21 18:58	191-24-2		
Benzo(g,h,i)perylene	<0.022	ug/L	0.095	0.022	1	11/04/21 19:31	11/05/21 14:19	191-24-2	H2	
Benzo(k)fluoranthene	<0.020	ug/L	0.10	0.020	1	10/28/21 10:39	11/01/21 18:58	207-08-9		
Benzo(k)fluoranthene	<0.019	ug/L	0.095	0.019	1	11/04/21 19:31	11/05/21 14:19	207-08-9	H2	
Chrysene	<0.020	ug/L	0.50	0.020	1	10/28/21 10:39	11/01/21 18:58	218-01-9		
Chrysene	<0.019	ug/L	0.48	0.019	1	11/04/21 19:31	11/05/21 14:19	218-01-9	H2	
Dibenz(a,h)anthracene	<0.071	ug/L	0.10	0.071	1	10/28/21 10:39	11/01/21 18:58	53-70-3		
Dibenz(a,h)anthracene	< 0.067	ug/L	0.095	0.067	1	11/04/21 19:31	11/05/21 14:19	53-70-3	H2	
Fluoranthene	<0.015	ug/L	1.0	0.015	1	10/28/21 10:39	11/01/21 18:58	206-44-0		
Fluoranthene	<0.015	ug/L	0.95	0.015	1	11/04/21 19:31	11/05/21 14:19	206-44-0	H2	
Fluorene	<0.036	ug/L	1.0	0.036	1	10/28/21 10:39	11/01/21 18:58	86-73-7	L2	
Fluorene	<0.034	ug/L	0.95	0.034	1	11/04/21 19:31	11/05/21 14:19	86-73-7	H2	
ndeno(1,2,3-cd)pyrene	<0.073	ug/L	0.10	0.073	1	10/28/21 10:39	11/01/21 18:58	193-39-5		
ndeno(1,2,3-cd)pyrene	<0.069	ug/L	0.095	0.069	1	11/04/21 19:31	11/05/21 14:19	193-39-5	H2	
2-Methylnaphthalene	<0.015	ug/L	1.0	0.015	1	10/28/21 10:39	11/01/21 18:58	91-57-6	L2	
2-Methylnaphthalene	<0.014	ug/L	0.95	0.014	1	11/04/21 19:31	11/05/21 14:19		H2	
Naphthalene	<0.014	ug/L	1.0	0.014	1	10/28/21 10:39	11/01/21 18:58	91-20-3	H7,L2	
Naphthalene	<0.013	ug/L	0.95	0.013	1	11/04/21 19:31	11/05/21 14:19		H2,H3	
Phenanthrene	<0.021	ug/L	1.0	0.021	1	10/28/21 10:39	11/01/21 18:58	85-01-8	L2	
Phenanthrene	<0.020	ug/L	0.95	0.020	1	11/04/21 19:31	11/05/21 14:19	85-01-8	H2	
Pyrene	<0.020	ug/L	1.0	0.020	1	10/28/21 10:39	11/01/21 18:58	129-00-0		
Pyrene	<0.019	ug/L	0.95	0.019	1	11/04/21 19:31	11/05/21 14:19	129-00-0	H2	



Project: Exide ERT Frankfort Site

Pace Project No.: 50300888

Date: 11/05/2021 04:07 PM

Sample: Water-1021	Lab ID: 50300888001 Collected: 10/22/21 08:20 Received: 10/22/21 11:40 Matrix: Water									
Parameters	Results	Units	PQL	MDL	DF	Prepared	Analyzed	CAS No.	Qua	
8270 100mL Combo RV	Analytical	Method: EPA 8	270 by SIM	l Preparati	on Meth	nod: EPA 3510				
	Pace Ana	ytical Services	 Indianapo 	olis						
Surrogates										
2-Fluorobiphenyl (S)	69	%.	31-98		1	11/04/21 19:31	11/05/21 14:19	321-60-8		
2-Fluorobiphenyl (S)	37	%.	31-98		1	10/28/21 10:39	11/01/21 18:58	321-60-8		
p-Terphenyl-d14 (S)	66	%.	33-115		1	10/28/21 10:39	11/01/21 18:58	1718-51-0		
p-Terphenyl-d14 (S)	84	%.	33-115		1	11/04/21 19:31	11/05/21 14:19	1718-51-0		
8270 SVOC Combo Water	Analytical	Method: EPA 8	270 Prepa	ration Meth	nod: EP	A 3510				
	Pace Ana	ytical Services	- Indianapo	olis						
Acetophenone	<2.8	ug/L	10.0	2.8	1	10/28/21 10:39	11/01/21 20:37	98-86-2		
Atrazine	<2.8	ug/L	10.0	2.8	1	10/28/21 10:39	11/01/21 20:37	1912-24-9		
Benzaldehyde	<4.7	ug/L	50.0	4.7	1	10/28/21 10:39	11/01/21 20:37	100-52-7		
Biphenyl (Diphenyl)	<5.9	ug/L	10.0	5.9	1	10/28/21 10:39	11/01/21 20:37	92-52-4		
4-Bromophenylphenyl ether	<5.6	ug/L	10.0	5.6	1	10/28/21 10:39	11/01/21 20:37	101-55-3		
Butylbenzylphthalate	<3.5	ug/L	10.0	3.5	1	10/28/21 10:39	11/01/21 20:37	85-68-7		
Caprolactam	<4.3	ug/L	10.0	4.3	1	10/28/21 10:39	11/01/21 20:37	105-60-2		
Carbazole	<3.7	ug/L	10.0	3.7	1	10/28/21 10:39	11/01/21 20:37	86-74-8		
4-Chloro-3-methylphenol	<5.6	ug/L	10.0	5.6	1	10/28/21 10:39	11/01/21 20:37	59-50-7		
4-Chloroaniline	<3.2	ug/L	10.0	3.2	1	10/28/21 10:39	11/01/21 20:37	106-47-8		
bis(2-Chloroethoxy)methane	<2.5	ug/L	10.0	2.5	1	10/28/21 10:39	11/01/21 20:37	111-91-1		
bis(2-Chloroethyl) ether	<2.9	ug/L	10.0	2.9	1	10/28/21 10:39	11/01/21 20:37	111-44-4		
2-Chloronaphthalene	<5.8	ug/L	10.0	5.8	1	10/28/21 10:39	11/01/21 20:37	91-58-7		
2-Chlorophenol	<3.6	ug/L	10.0	3.6	1	10/28/21 10:39	11/01/21 20:37			
4-Chlorophenylphenyl ether	<5.1	ug/L	10.0	5.1	1	10/28/21 10:39	11/01/21 20:37	7005-72-3		
Dibenzofuran	<7.0	ug/L	10.0	7.0	1	10/28/21 10:39	11/01/21 20:37			
3,3'-Dichlorobenzidine	<4.0	ug/L	20.0	4.0	1	10/28/21 10:39	11/01/21 20:37			
2,4-Dichlorophenol	<4.0	ug/L	10.0	4.0	1	10/28/21 10:39	11/01/21 20:37			
Diethylphthalate	<2.7	ug/L	10.0	2.7	1		11/01/21 20:37			
2,4-Dimethylphenol	<8.1	ug/L	10.0	8.1	1	10/28/21 10:39	11/01/21 20:37			
Dimethylphthalate	<3.7	ug/L	10.0	3.7	1	10/28/21 10:39	11/01/21 20:37	131-11-3		
Di-n-butylphthalate	<3.6	ug/L	10.0	3.6	1	10/28/21 10:39	11/01/21 20:37	84-74-2		
4,6-Dinitro-2-methylphenol	<5.0	ug/L	20.0	5.0	1	10/28/21 10:39	11/01/21 20:37	534-52-1		
2,4-Dinitrophenol	<6.6	ug/L	50.0	6.6	1	10/28/21 10:39	11/01/21 20:37			
2.4-Dinitrotoluene	<6.2	ug/L	10.0	6.2	1		11/01/21 20:37			
2,6-Dinitrotoluene	<4.6	ug/L	10.0	4.6	1		11/01/21 20:37			
Di-n-octylphthalate	<4.5	ug/L	10.0	4.5	1		11/01/21 20:37			
bis(2-Ethylhexyl)phthalate	<3.1	ug/L	10.0	3.1	1		11/01/21 20:37			
Hexachloro-1,3-butadiene	<4.1	ug/L	10.0	4.1	1		11/01/21 20:37			
Hexachlorobenzene	<3.0	ug/L	10.0	3.0	1		11/01/21 20:37			
Hexachlorocyclopentadiene	<3.0	ug/L	10.0	3.0	1		11/01/21 20:37			
Hexachloroethane	<2.5	ug/L	10.0	2.5	1		11/01/21 20:37			
Isophorone	<4.2	ug/L	10.0	4.2	1		11/01/21 20:37			
2-Methylphenol(o-Cresol)	<4.3	ug/L	10.0	4.3	1		11/01/21 20:37			
3&4-Methylphenol(m&p Cresol)	<5.4	ug/L	10.0	5.4	1		11/01/21 20:37	2- ·- ·		
2-Nitroaniline	<4.2	ug/L	10.0	4.2	1	10/28/21 10:39	11/01/21 20:37	88-74-4		
3-Nitroaniline	<4.8	ug/L	10.0	4.8	1		11/01/21 20:37			



Project: Exide ERT Frankfort Site

Pace Project No.: 50300888

Date: 11/05/2021 04:07 PM

Sample: Water-1021	Lab ID: 50300888001 Collected: 10/22/21 08:20 Received: 10/22/21 11:40 Matrix: W								
Parameters	Results	Units	PQL	MDL	DF	Prepared	Analyzed	CAS No.	Qua
8270 SVOC Combo Water	Analytical	Method: EPA 8	270 Prepa	ration Metho	od: EPA	3510			
	Pace Anal	ytical Services	- Indianapo	lis					
4-Nitroaniline	<4.6	ug/L	10.0	4.6	1	10/28/21 10:39	11/01/21 20:37	100-01-6	
Nitrobenzene	<3.0	ug/L	10.0	3.0	1	10/28/21 10:39	11/01/21 20:37		
2-Nitrophenol	<3.5	ug/L	10.0	3.5	1	10/28/21 10:39	11/01/21 20:37		
4-Nitrophenol	<5.6	ug/L	50.0	5.6	1	10/28/21 10:39	11/01/21 20:37	100-02-7	
N-Nitroso-di-n-propylamine	<2.9	ug/L	50.0	2.9	1	10/28/21 10:39	11/01/21 20:37		
N-Nitrosodiphenylamine	<2.9	ug/L	10.0	2.9	1	10/28/21 10:39	11/01/21 20:37		
2,2'-Oxybis(1-chloropropane)	<4.6	ug/L	10.0	4.6	1	10/28/21 10:39	11/01/21 20:37		
Pentachlorophenol	<4.0	ug/L	50.0	4.0	1	10/28/21 10:39	11/01/21 20:37		
Phenol	5.9J	ug/L	10.0	4.1	1	10/28/21 10:39	11/01/21 20:37		
2,3,4,6-Tetrachlorophenol	<4.9	ug/L	10.0	4.9	1	10/28/21 10:39	11/01/21 20:37		
2,4,5-Trichlorophenol	<2.9	ug/L	10.0	2.9	1	10/28/21 10:39	11/01/21 20:37		
2,4,6-Trichlorophenol	<4.5	ug/L	10.0	4.5	1	10/28/21 10:39	11/01/21 20:37		
Surrogates	77.0	~9, -	10.0	7.0	•	. 3/20/21 10:00	. 1, 0 1, 2 1 20.01	20 00 2	
Nitrobenzene-d5 (S)	40	%.	39-115		1	10/28/21 10:39	11/01/21 20:37	4165-60-0	
Phenol-d5 (S)	26	%.	10-55		1	10/28/21 10:39	11/01/21 20:37		
2-Fluorophenol (S)	34	%.	10-72		1	10/28/21 10:39	11/01/21 20:37		
2,4,6-Tribromophenol (S)	65	%.	34-126		1		11/01/21 20:37		
3260 MSV TCLP	Analytical	Method: EPA 5	030/8260 I	_eachate M	ethod/D	ate: EPA 1311; 1	1/01/21 20:50		
	•	ytical Services				•			
Benzene	<0.010	mg/L	0.050	0.010	1		11/04/21 05:11	71-43-2	
2-Butanone (MEK)	<0.50	mg/L	1.0	0.50	1		11/04/21 05:11		
Carbon tetrachloride	<0.025	mg/L	0.050	0.025	1		11/04/21 05:11		
Chlorobenzene	<0.025	mg/L	0.050	0.025	1		11/04/21 05:11	108-90-7	
Chloroform	<0.025	mg/L	0.050	0.025	1		11/04/21 05:11	67-66-3	
1,2-Dichloroethane	<0.025	mg/L	0.050	0.025	1		11/04/21 05:11	107-06-2	
1,1-Dichloroethene	<0.025	mg/L	0.050	0.025	1		11/04/21 05:11		
Tetrachloroethene	<0.025	mg/L	0.050	0.025	1			127-18-4	
Trichloroethene	<0.025	mg/L	0.050	0.025	1		11/04/21 05:11		
Vinyl chloride	<0.010	mg/L	0.020	0.023	1		11/04/21 05:11		
Surrogates	30.010	g/ <u>_</u>	0.020	0.010	'		. 1/0-7/21 00.11	.001-4	
4-Bromofluorobenzene (S)	94	%.	78-117		1		11/04/21 05:11	460-00-4	
Dibromofluoromethane (S)	107	%.	78-120		1		11/04/21 05:11	1868-53-7	
Toluene-d8 (S)	101	%.	77-118		1		11/04/21 05:11	2037-26-5	
3260/5030 MSV	Analytical	Method: EPA 8	260						
	•	ytical Services		lis					
Acetone	159	ug/L	100	5.5	1		11/01/21 14:54	67-64-1	
Benzene	<0.31	ug/L	5.0	0.31	1		11/01/21 14:54		
Bromochloromethane	<0.42	ug/L	5.0	0.42	1		11/01/21 14:54		
Bromodichloromethane	<0.29	ug/L	5.0	0.29	1		11/01/21 14:54		
Bromoform	<0.42	ug/L	5.0	0.42	1		11/01/21 14:54		
	<1.6	ug/L	5.0	1.6	1		11/01/21 14:54		
Bromomethane	<1.0	uu/L	5.0	1.0	- 1		11/01/21 14.14	14-00-0	
Bromomethane 2-Butanone (MEK)	<2.1	ug/L ug/L	25.0	2.1	1		11/01/21 14:54		



Project: Exide ERT Frankfort Site

Pace Project No.: 50300888

Date: 11/05/2021 04:07 PM

Sample: Water-1021 Lab ID: 50300888001 Collected: 10/22/21 08:20 Received: 10/22/21 11:40 Matrix: Water

Parameters	Results	Units	PQL	MDL	DF	Prepared	Analyzed	CAS No.	Qua
8260/5030 MSV	Analytical	Method: EPA	x 8260						
	Pace Anal	ytical Service	es - Indianapo	lis					
Carbon tetrachloride	<0.48	ug/L	5.0	0.48	1		11/01/21 14:54	56-23-5	
Chlorobenzene	<0.33	ug/L	5.0	0.33	1		11/01/21 14:54		
Chloroethane	<1.7	ug/L	5.0	1.7	1		11/01/21 14:54		
Chloroform	<0.34	ug/L	5.0	0.34	1		11/01/21 14:54		
Chloromethane	<0.48	ug/L	5.0	0.48	1		11/01/21 14:54		
Cyclohexane	<0.36	ug/L	100	0.36	1		11/01/21 14:54		
1,2-Dibromo-3-chloropropane	<1.6	ug/L	10.0	1.6	1		11/01/21 14:54		
Dibromochloromethane	<0.34	ug/L	5.0	0.34	1		11/01/21 14:54		
1,2-Dibromoethane (EDB)	<0.42	ug/L	5.0	0.42	1		11/01/21 14:54		
1,2-Dichlorobenzene	<0.30	ug/L	5.0	0.30	1		11/01/21 14:54		
1,3-Dichlorobenzene	<0.30	ug/L	5.0	0.30	1		11/01/21 14:54		
1,4-Dichlorobenzene	<0.36	ug/L	5.0	0.36	1		11/01/21 14:54		
Dichlorodifluoromethane	<0.30 <1.7	ug/L ug/L	5.0	1.7	1		11/01/21 14:54		
1,1-Dichloroethane	<0.41	•	5.0	0.41	1		11/01/21 14:54		
•	<0.41 <0.41	ug/L	5.0	0.41	1		11/01/21 14:54		
1,2-Dichloroethane		ug/L							
I,1-Dichloroethene	<0.37	ug/L	5.0	0.37	1		11/01/21 14:54		
cis-1,2-Dichloroethene	7.8	ug/L	5.0	0.46	1		11/01/21 14:54		
rans-1,2-Dichloroethene	<0.32	ug/L	5.0	0.32	1		11/01/21 14:54		
1,2-Dichloropropane	<0.35	ug/L	5.0	0.35	1		11/01/21 14:54		
cis-1,3-Dichloropropene	<0.34	ug/L	5.0	0.34	1		11/01/21 14:54		
rans-1,3-Dichloropropene	<0.27	ug/L	5.0	0.27	1		11/01/21 14:54		
Ethylbenzene	<0.26	ug/L	5.0	0.26	1		11/01/21 14:54		
2-Hexanone	<1.4	ug/L	25.0	1.4	1		11/01/21 14:54		
sopropylbenzene (Cumene)	<0.34	ug/L	5.0	0.34	1		11/01/21 14:54		
Methyl acetate	<0.76	ug/L	50.0	0.76	1		11/01/21 14:54		
Methylcyclohexane	<0.36	ug/L	50.0	0.36	1		11/01/21 14:54		
Methylene Chloride	<0.081	ug/L	5.0	0.081	1		11/01/21 14:54		
1-Methyl-2-pentanone (MIBK)	<1.4	ug/L	25.0	1.4	1		11/01/21 14:54		
Methyl-tert-butyl ether	<0.31	ug/L	4.0	0.31	1		11/01/21 14:54		
Styrene	<0.26	ug/L	5.0	0.26	1		11/01/21 14:54		
1,1,2,2-Tetrachloroethane	<0.26	ug/L	5.0	0.26	1		11/01/21 14:54		
Tetrachloroethene	<0.44	ug/L	5.0	0.44	1		11/01/21 14:54	_	
Toluene	<0.27	ug/L	5.0	0.27	1		11/01/21 14:54		
1,2,3-Trichlorobenzene	<0.50	ug/L	5.0	0.50	1		11/01/21 14:54	87-61-6	
1,2,4-Trichlorobenzene	<0.44	ug/L	5.0	0.44	1		11/01/21 14:54		
1,1,1-Trichloroethane	<0.40	ug/L	5.0	0.40	1		11/01/21 14:54	71-55-6	
1,1,2-Trichloroethane	<0.30	ug/L	5.0	0.30	1		11/01/21 14:54		
Trichloroethene	0.61J	ug/L	5.0	0.46	1		11/01/21 14:54		
Trichlorofluoromethane	<0.24	ug/L	5.0	0.24	1		11/01/21 14:54	75-69-4	
1,1,2-Trichlorotrifluoroethane	<0.49	ug/L	5.0	0.49	1		11/01/21 14:54	76-13-1	
/inyl chloride	1.9J	ug/L	2.0	0.28	1		11/01/21 14:54	75-01-4	
Xylene (Total) Surrogates	<0.68	ug/L	10.0	0.68	1		11/01/21 14:54	1330-20-7	
Dibromofluoromethane (S)	105	%.	78-120		1		11/01/21 14:54	1868-53-7	
4-Bromofluorobenzene (S)	96	%.	78-117		1		11/01/21 14:54	460-00-4	



Project: Exide ERT Frankfort Site

Pace Project No.: 50300888

Date: 11/05/2021 04:07 PM

Sample: Water-1021	Lab ID:	50300888001	Collected	d: 10/22/2	1 08:20	Received: 10/	22/21 11:40 M	latrix: Water	
Parameters	Results	Units	PQL	MDL	DF	Prepared	Analyzed	CAS No.	Qual
8260/5030 MSV	•	Method: EPA 8							
	Pace Ana	llytical Services	- Indianapol	is					
Surrogates Toluene-d8 (S)	101	%.	77-118		1		11/01/21 14:54	2037-26-5	
1020 Flashpoint,Closed Cup	•	l Method: EPA 1 llytical Services		is					
Flashpoint	>200.0	deg F			1		10/26/21 13:17	•	N2
4500H+ pH, Electrometric	•	l Method: SM 45 llytical Services		is					
pH at 25 Degrees C	8.7	Std. Units	0.10	0.10	1		10/30/21 09:37	•	НЗ
733C S Reactive Cyanide	•	l Method: EPA 9 llytical Services	•		nod: SW-	846 7.3.3.2			
Cyanide, Reactive	<0.40	mg/kg	1.0	0.40	1	10/27/21 13:27	11/02/21 12:21		
734S Reactive Sulfide	•	l Method: SM 45 llytical Services		•	ration Me	ethod: SW-846 7.	3.4.2		
Sulfide, Reactive	<10.0	mg/kg	10.0	10.0	1	10/29/21 14:32	10/29/21 15:02	!	



Project: Exide ERT Frankfort Site

Pace Project No.: 50300888

Date: 11/05/2021 04:07 PM

Sample: Soil-1021 Lab ID: 50300888002 Collected: 10/22/21 08:50 Received: 10/22/21 11:40 Matrix: Solid

Results reported on a "dry weight" basis and are adjusted for percent moisture, sample size and any dilutions.

Parameters	Results	Units	PQL	MDL	DF	Prepared	Analyzed	CAS No.	Qual
6010 MET ICP	Analytical	Method: EPA	A 6010 Prepa	ration Meth	od: EP	A 3050			
	Pace Anal	ytical Service	es - Indianapo	olis					
Arsenic	7.6	mg/kg	1.2	0.25	1	10/27/21 07:17	10/27/21 12:52	7440-38-2	
Barium	76.5	mg/kg	1.2	0.042	1		10/27/21 12:52		
Cadmium	0.62	mg/kg	0.60	0.017	1	10/27/21 07:17	10/27/21 12:52	7440-43-9	
Chromium	14.4	mg/kg	1.2	0.089	1	10/27/21 07:17	10/27/21 12:52	7440-47-3	
Lead	47.7	mg/kg	1.2	0.12	1	10/27/21 07:17	10/27/21 12:52	7439-92-1	
Selenium	<0.32	mg/kg	1.2	0.32	1	10/27/21 07:17	10/27/21 12:52	7782-49-2	
Silver	<0.22	mg/kg	0.60	0.22	1	10/27/21 07:17	10/27/21 12:52	7440-22-4	
6010 MET ICP, TCLP	Analytical	Method: EPA	A 6010 Prepa	ration Meth	od: EP	A 3010			
	Leachate	Method/Date	: EPA 1311; 1	1/01/21 20:	50 Initi	al pH: 8.87; Final	pH: 6.4		
			es - Indianapo						
Arsenic	<0.050	mg/L	0.10	0.050	1	11/03/21 13:20	11/04/21 00:14	7440-38-2	
Barium	0.82J	mg/L	5.0	0.25	1	11/03/21 13:20	11/04/21 00:14	7440-39-3	
Cadmium	<0.025	mg/L	0.050	0.025	1	11/03/21 13:20	11/04/21 00:14	7440-43-9	
Chromium	<0.052	mg/L	0.10	0.052	1	11/03/21 13:20	11/04/21 00:14	7440-47-3	
Lead	<0.050	mg/L	0.10	0.050	1	11/03/21 13:20	11/04/21 00:14	7439-92-1	
Selenium	<0.050	mg/L	0.10	0.050	1	11/03/21 13:20	11/04/21 00:14	7782-49-2	
Silver	<0.050	mg/L	0.10	0.050	1	11/03/21 13:20	11/04/21 00:14	7440-22-4	
7470 Mercury, TCLP	Analytical	Method: EPA	A 7470 Prepa	ration Meth	od: EP	A 7470			
•	Leachate	Method/Date	: EPA 1311: 1	1/01/21 20:	50 Initi	ial pH: 8.87; Final	pH: 6.4		
			es - Indianapo			,	•		
Mercury	<0.0010	mg/L	0.0020	0.0010	1	11/03/21 10:41	11/04/21 11:37	7439-97-6	
7471 Mercury	Analytical	Method: EPA	A 7471 Prepa	ration Meth	od: EP	A 7471			
,,,	-		es - Indianapo						
Mercury	0.037J	mg/kg	0.26	0.031	1	10/31/21 13:27	11/01/21 10:00	7439-97-6	
8270 SVOC SS Soil	Analytical	Method: EPA	A 8270 Prepa	ration Meth	od: EP	A 3546			
	-		es - Indianapo						
Acenaphthene	<0.11	mg/kg	0.42	0.11	1	10/31/21 15:40	11/01/21 15:25	83-32-9	
Acenaphthylene	<0.13	mg/kg	0.42	0.13	1	10/31/21 15:40	11/01/21 15:25	208-96-8	
Acetophenone	<0.13	mg/kg	0.42	0.13	1	10/31/21 15:40	11/01/21 15:25	98-86-2	
Anthracene	<0.17	mg/kg	0.42	0.17	1	10/31/21 15:40	11/01/21 15:25	120-12-7	
Atrazine	<0.17	mg/kg	0.42	0.17	1	10/31/21 15:40	11/01/21 15:25	1912-24-9	N2
Benzaldehyde	<0.14	mg/kg	0.42	0.14	1	10/31/21 15:40	11/01/21 15:25	100-52-7	N2
Benzo(a)anthracene	<0.13	mg/kg	0.42	0.13	1	10/31/21 15:40	11/01/21 15:25		
Benzo(a)pyrene	<0.14	mg/kg	0.42	0.14	1	10/31/21 15:40	11/01/21 15:25		
Benzo(b)fluoranthene	<0.14	mg/kg	0.42	0.14	1	10/31/21 15:40			
Benzo(g,h,i)perylene	<0.15	mg/kg	0.42	0.15	1		11/01/21 15:25		
Benzo(k)fluoranthene	<0.15	mg/kg	0.42	0.15	1	10/31/21 15:40			
Biphenyl (Diphenyl)	<0.12	mg/kg	0.42	0.12	1	10/31/21 15:40			N2
4-Bromophenylphenyl ether	<0.16	mg/kg	0.42	0.16	1		11/01/21 15:25		
• • • •									



Project: Exide ERT Frankfort Site

Pace Project No.: 50300888

Date: 11/05/2021 04:07 PM

Sample: Soil-1021 Lab ID: 50300888002 Collected: 10/22/21 08:50 Received: 10/22/21 11:40 Matrix: Solid

Results reported on a "dry weight" basis and are adjusted for percent moisture, sample size and any dilutions.

Parameters	Results	Units	PQL	MDL	DF	Prepared	Analyzed	CAS No.	Qual
8270 SVOC SS Soil	-		A 8270 Prepa es - Indianapo		od: EP	A 3546			
Putulbanzulahthalata	<0.23		0.42	0.23	1	10/31/21 15:40	11/01/21 15:25	05 60 7	
Butylbenzylphthalate	<0.23 <0.21	mg/kg	0.42	0.23	1	10/31/21 15:40	11/01/21 15:25		N2
Carbazala		mg/kg			1				INZ
Carbazole	<0.17	mg/kg	0.42	0.17 0.17		10/31/21 15:40	11/01/21 15:25		
4-Chloro-3-methylphenol	<0.17	mg/kg	0.85		1	10/31/21 15:40	11/01/21 15:25		
4-Chloroaniline	<0.11	mg/kg	0.85	0.11	1	10/31/21 15:40	11/01/21 15:25		
bis(2-Chloroethoxy)methane	<0.14	mg/kg	0.42	0.14	1	10/31/21 15:40	11/01/21 15:25		
bis(2-Chloroethyl) ether	<0.16	mg/kg	0.42	0.16	1	10/31/21 15:40	11/01/21 15:25		
2-Chloronaphthalene	<0.12	mg/kg	0.42	0.12	1	10/31/21 15:40	11/01/21 15:25		
2-Chlorophenol	<0.15	mg/kg	0.42	0.15	1	10/31/21 15:40	11/01/21 15:25		
4-Chlorophenylphenyl ether	<0.13	mg/kg	0.42	0.13	1	10/31/21 15:40	11/01/21 15:25		
Chrysene	<0.14	mg/kg	0.42	0.14	1	10/31/21 15:40	11/01/21 15:25	218-01-9	
Dibenz(a,h)anthracene	<0.15	mg/kg	0.42	0.15	1	10/31/21 15:40	11/01/21 15:25	53-70-3	
Dibenzofuran	<0.13	mg/kg	0.42	0.13	1	10/31/21 15:40	11/01/21 15:25	132-64-9	
3,3'-Dichlorobenzidine	<0.14	mg/kg	0.85	0.14	1	10/31/21 15:40	11/01/21 15:25	91-94-1	
2,4-Dichlorophenol	<0.15	mg/kg	0.42	0.15	1	10/31/21 15:40	11/01/21 15:25	120-83-2	
Diethylphthalate	<0.14	mg/kg	0.42	0.14	1	10/31/21 15:40	11/01/21 15:25	84-66-2	
2,4-Dimethylphenol	<0.15	mg/kg	0.42	0.15	1	10/31/21 15:40	11/01/21 15:25	105-67-9	
Dimethylphthalate	<0.14	mg/kg	0.42	0.14	1	10/31/21 15:40	11/01/21 15:25	131-11-3	
Di-n-butylphthalate	<0.15	mg/kg	0.42	0.15	1	10/31/21 15:40	11/01/21 15:25	84-74-2	
4,6-Dinitro-2-methylphenol	<0.26	mg/kg	0.85	0.26	1	10/31/21 15:40	11/01/21 15:25	534-52-1	
2,4-Dinitrophenol	<0.23	mg/kg	2.1	0.23	1	10/31/21 15:40	11/01/21 15:25	51-28-5	
2,4-Dinitrotoluene	<0.14	mg/kg	0.42	0.14	1	10/31/21 15:40	11/01/21 15:25	121-14-2	
2,6-Dinitrotoluene	<0.12	mg/kg	0.42	0.12	1	10/31/21 15:40	11/01/21 15:25		
Di-n-octylphthalate	<0.16	mg/kg	0.42	0.16	1	10/31/21 15:40	11/01/21 15:25		
bis(2-Ethylhexyl)phthalate	<0.13	mg/kg	0.42	0.13	1	10/31/21 15:40	11/01/21 15:25		
Fluoranthene	<0.16	mg/kg	0.42	0.16	1	10/31/21 15:40	11/01/21 15:25		
Fluorene	<0.14	mg/kg	0.42	0.14	1	10/31/21 15:40	11/01/21 15:25		
Hexachloro-1,3-butadiene	<0.12	mg/kg	0.42	0.12	1	10/31/21 15:40	11/01/21 15:25		
Hexachlorobenzene	<0.11	mg/kg	0.42	0.12	1	10/31/21 15:40	11/01/21 15:25		
Hexachlorocyclopentadiene	<0.21	mg/kg	0.42	0.11	1	10/31/21 15:40	11/01/21 15:25		
Hexachloroethane	<0.13	mg/kg	0.42	0.21	1	10/31/21 15:40	11/01/21 15:25		
Indeno(1,2,3-cd)pyrene	<0.15		0.42	0.15	1	10/31/21 15:40	11/01/21 15:25		
· · · · // /		mg/kg				10/31/21 15:40			
Isophorone 2 Methylpophthologo	<0.14	mg/kg	0.42	0.14	1	10/31/21 15:40	11/01/21 15:25		
2-Methylnaphthalene	<0.13	mg/kg	0.42	0.13	1		11/01/21 15:25		
2-Methylphenol(o-Cresol)	<0.18	mg/kg	0.42	0.18	1	10/31/21 15:40	11/01/21 15:25	95-48-7	
3&4-Methylphenol(m&p Cresol)	<0.18	mg/kg	0.85	0.18	1	10/31/21 15:40	11/01/21 15:25		
Naphthalene	<0.12	mg/kg	0.42	0.12	1	10/31/21 15:40	11/01/21 15:25		
2-Nitroaniline	<0.17	mg/kg	0.42	0.17	1	10/31/21 15:40	11/01/21 15:25		
3-Nitroaniline	<0.15	mg/kg	0.42	0.15	1	10/31/21 15:40	11/01/21 15:25		
4-Nitroaniline	<0.17	mg/kg	0.42	0.17	1		11/01/21 15:25		
Nitrobenzene	<0.14	mg/kg	0.42	0.14	1	10/31/21 15:40	11/01/21 15:25		
2-Nitrophenol	<0.16	mg/kg	0.42	0.16	1		11/01/21 15:25		
4-Nitrophenol	<0.32	mg/kg	2.1	0.32	1		11/01/21 15:25		
N-Nitroso-di-n-propylamine	<0.16	mg/kg	0.42	0.16	1	10/31/21 15:40	11/01/21 15:25	621-64-7	



Project: Exide ERT Frankfort Site

Pace Project No.: 50300888

Date: 11/05/2021 04:07 PM

Sample: Soil-1021 Lab ID: 50300888002 Collected: 10/22/21 08:50 Received: 10/22/21 11:40 Matrix: Solid

Results reported on a "dry weight" basis and are adjusted for percent moisture, sample size and any dilutions.

Analytical Method: EPA 8270 Preparation Method: EPA 3546	
N-Nitrosodiphenylamine	
2,2'-Oxybis(1-chloropropane) <0.15	
Pentachlorophenol <0.33 mg/kg 2.1 0.33 1 10/31/21 15:40 11/01/21 15:25 8 Phenanthrene <0.17	86-30-6
Pentachlorophenol <0.33 mg/kg 2.1 0.33 1 10/31/21 15:40 11/01/21 15:25 8 Phenanthrene <0.17	108-60-1
Phenanthrene <0.17 mg/kg 0.42 0.17 1 10/31/21 15:40 11/01/21 15:25 8 Phenol <0.16	87-86-5
Phenol <0.16 mg/kg 0.42 0.16 1 10/31/21 15:40 11/01/21 15:25 1 Pyrene <0.13	85-01-8
Pyrene	108-95-2
1,2,4,5-Tetrachlorobenzene <0.10	129-00-0
2,3,4,6-Tetrachlorophenol <0.15 mg/kg 0.42 0.15 1 10/31/21 15:40 11/01/21 15:25 5 2,4,5-Trichlorophenol <0.15	95-94-3
2,4,5-Trichlorophenol <0.15 mg/kg 0.42 0.15 1 10/31/21 15:40 11/01/21 15:25 2 2,4,6-Trichlorophenol <0.13 mg/kg 0.42 0.13 1 10/31/21 15:40 11/01/21 15:25 8 Surrogates Nitrobenzene-d5 (S) 67 %. 32-105 1 10/31/21 15:40 11/01/21 15:25 4 Phenol-d5 (S) 70 %. 35-114 1 10/31/21 15:40 11/01/21 15:25 4 2-Fluorophenol (S) 65 %. 33-111 1 10/31/21 15:40 11/01/21 15:25 3 2,4,6-Tribromophenol (S) 40 %. 20-121 1 10/31/21 15:40 11/01/21 15:25 1	58-90-2
2,4,6-Trichlorophenol <0.13 mg/kg 0.42 0.13 1 10/31/21 15:40 11/01/21 15:25 8 Surrogates Nitrobenzene-d5 (S) 67 %. 32-105 1 10/31/21 15:40 11/01/21 15:25 4 Phenol-d5 (S) 70 %. 35-114 1 10/31/21 15:40 11/01/21 15:25 4 2-Fluorophenol (S) 65 %. 33-111 1 10/31/21 15:40 11/01/21 15:25 3 2,4,6-Tribromophenol (S) 40 %. 20-121 1 10/31/21 15:40 11/01/21 15:25 1	
Surrogates Nitrobenzene-d5 (S) 67 %. 32-105 1 10/31/21 15:40 11/01/21 15:25 4 Phenol-d5 (S) 70 %. 35-114 1 10/31/21 15:40 11/01/21 15:25 4 2-Fluorophenol (S) 65 %. 33-111 1 10/31/21 15:40 11/01/21 15:25 3 2,4,6-Tribromophenol (S) 40 %. 20-121 1 10/31/21 15:40 11/01/21 15:25 1	
Nitrobenzene-d5 (S) 67 %. 32-105 1 10/31/21 15:40 11/01/21 15:25 4 Phenol-d5 (S) 70 %. 35-114 1 10/31/21 15:40 11/01/21 15:25 4 2-Fluorophenol (S) 65 %. 33-111 1 10/31/21 15:40 11/01/21 15:25 3 2,4,6-Tribromophenol (S) 40 %. 20-121 1 10/31/21 15:40 11/01/21 15:25 1	00 00 2
Phenol-d5 (S) 70 %. 35-114 1 10/31/21 15:40 11/01/21 15:25 4 2-Fluorophenol (S) 65 %. 33-111 1 10/31/21 15:40 11/01/21 15:25 3 2,4,6-Tribromophenol (S) 40 %. 20-121 1 10/31/21 15:40 11/01/21 15:25 1	4165-60-0
2-Fluorophenol (S) 65 %. 33-111 1 10/31/21 15:40 11/01/21 15:25 3 2,4,6-Tribromophenol (S) 40 %. 20-121 1 10/31/21 15:40 11/01/21 15:25 1	
2,4,6-Tribromophenol (S) 40 %. 20-121 1 10/31/21 15:40 11/01/21 15:25 1	
z-mananancus or /r. 33-90 1 10/31/21 13:40 11/01/21 13:23 3	
p-Terphenyl-d14 (S) 67 %. 31-145 1 10/31/21 15:40 11/01/21 15:25 1	
8270 MSSV TCLP Sep Funnel Analytical Method: EPA 8270 Preparation Method: EPA 3510	
Leachate Method/Date: EPA 1311; 11/01/21 20:50 Initial pH: 8.87; Final pH: 6.4	
Pace Analytical Services - Indianapolis	
1,4-Dichlorobenzene <0.050 mg/L 0.10 0.050 1 11/02/21 19:55 11/04/21 00:24 1	106-46-7
2,4-Dinitrotoluene <0.050 mg/L 0.10 0.050 1 11/02/21 19:55 11/04/21 00:24 1	
Hexachloro-1,3-butadiene <0.050 mg/L 0.10 0.050 1 11/02/21 19:55 11/04/21 00:24 8	
Hexachlorobenzene <0.050 mg/L 0.10 0.050 1 11/02/21 19:55 11/04/21 00:24 1	
Hexachloroethane <0.050 mg/L 0.10 0.050 1 11/02/21 19:55 11/04/21 00:24 6	
2-Methylphenol(o-Cresol)	
3&4-Methylphenol(m&p Cresol)	33-40-7
	00 05 3
y	
Pentachlorophenol <0.25 mg/L 0.50 0.25 1 11/02/21 19:55 11/04/21 00:24 8	
Pyridine <0.10 mg/L 0.10 0.10 1 11/02/21 19:55 11/04/21 00:24 1	
2,4,5-Trichlorophenol <0.050 mg/L 0.50 0.050 1 11/02/21 19:55 11/04/21 00:24 9	
2,4,6-Trichlorophenol <0.050 mg/L 0.10 0.050 1 11/02/21 19:55 11/04/21 00:24 8	გგ- ებ-2
Surrogates Nitrobenzene-d5 (S) 65 %. 40-115 1 11/02/21 19:55 11/04/21 00:24 4	4165 GO O
p-Terphenyl-d14 (S) 75 %. 42-156 1 11/02/21 19:55 11/04/21 00:24 1	
Phenol-d5 (S) 29 %. 15-48 1 11/02/21 19:55 11/04/21 00:24 4	
2-Fluorophenol (S) 40 %. 21-74 1 11/02/21 19:55 11/04/21 00:24 3	
2,4,6-Tribromophenol (S) 82 %. 47-127 1 11/02/21 19:55 11/04/21 00:24 1	118-79-6
8260 MSV TCLP Analytical Method: EPA 5030/8260 Leachate Method/Date: EPA 1311; 11/01/21 20:50	
Pace Analytical Services - Indianapolis	
Benzene <0.010 mg/L 0.050 0.010 1 11/04/21 05:43 7	



Project: Exide ERT Frankfort Site

Pace Project No.: 50300888

Date: 11/05/2021 04:07 PM

Sample: Soil-1021 Lab ID: 50300888002 Collected: 10/22/21 08:50 Received: 10/22/21 11:40 Matrix: Solid

Results reported on a "dry weight" basis and are adjusted for percent moisture, sample size and any dilutions.

Parameters	Results	Units	PQL	MDL	DF	Prepared	Analyzed	CAS No.	Qual
8260 MSV TCLP	Analytical	Method: EPA	A 5030/8260	Leachate M	ethod/Da	ate: EPA 1311;	11/01/21 20:50		
	Pace Ana	ytical Service	es - Indianapo	olis					
2-Butanone (MEK)	<0.50	mg/L	1.0	0.50	1		11/04/21 05:43	78-93-3	
Carbon tetrachloride	<0.025	mg/L	0.050	0.025	1		11/04/21 05:43	56-23-5	
Chlorobenzene	< 0.025	mg/L	0.050	0.025	1		11/04/21 05:43	108-90-7	
Chloroform	< 0.025	mg/L	0.050	0.025	1		11/04/21 05:43	67-66-3	
1,2-Dichloroethane	< 0.025	mg/L	0.050	0.025	1		11/04/21 05:43	107-06-2	
1,1-Dichloroethene	< 0.025	mg/L	0.050	0.025	1		11/04/21 05:43	75-35-4	
Tetrachloroethene	< 0.025	mg/L	0.050	0.025	1		11/04/21 05:43	127-18-4	
Trichloroethene	< 0.025	mg/L	0.050	0.025	1		11/04/21 05:43	79-01-6	
Vinyl chloride	<0.010	mg/L	0.020	0.010	1		11/04/21 05:43		
Surrogates		· ·							
4-Bromofluorobenzene (S)	94	%.	78-117		1		11/04/21 05:43	460-00-4	
Dibromofluoromethane (S)	106	%.	78-120		1		11/04/21 05:43	1868-53-7	
Toluene-d8 (S)	100	%.	77-118		1		11/04/21 05:43	2037-26-5	
8260 MSV 5035A VOA	Analytical	Method: EPA	A 8260						
	Pace Ana	ytical Service	es - Indianapo	olis					
Acetone	0.010J	mg/kg	0.096	0.0020	1		11/04/21 21:48	67-64-1	
Benzene	< 0.00039	mg/kg	0.0048	0.00039	1		11/04/21 21:48	71-43-2	
Bromochloromethane	< 0.00054	mg/kg	0.0048	0.00054	1		11/04/21 21:48	74-97-5	
Bromodichloromethane	< 0.00036	mg/kg	0.0048	0.00036	1		11/04/21 21:48	75-27-4	
Bromoform	< 0.00036	mg/kg	0.0048	0.00036	1		11/04/21 21:48	75-25-2	
Bromomethane	< 0.00029	mg/kg	0.0048	0.00029	1		11/04/21 21:48	74-83-9	
2-Butanone (MEK)	< 0.0066	mg/kg	0.024	0.0066	1		11/04/21 21:48	78-93-3	
Carbon disulfide	< 0.00057	mg/kg	0.0096	0.00057	1		11/04/21 21:48	75-15-0	
Carbon tetrachloride	< 0.00034	mg/kg	0.0048	0.00034	1		11/04/21 21:48	56-23-5	
Chlorobenzene	< 0.00035	mg/kg	0.0048	0.00035	1		11/04/21 21:48	108-90-7	
Chloroethane	< 0.00021	mg/kg	0.0048	0.00021	1		11/04/21 21:48	75-00-3	
Chloroform	0.00096J	mg/kg	0.0048	0.00045	1		11/04/21 21:48	67-66-3	В
Chloromethane	< 0.00017	mg/kg	0.0048	0.00017	1		11/04/21 21:48	74-87-3	
Cyclohexane	< 0.00042	mg/kg	0.096	0.00042	1		11/04/21 21:48	110-82-7	N2
1,2-Dibromo-3-chloropropane	< 0.00065	mg/kg	0.0096	0.00065	1		11/04/21 21:48	96-12-8	
Dibromochloromethane	< 0.00035	mg/kg	0.0048	0.00035	1		11/04/21 21:48	124-48-1	
1,2-Dibromoethane (EDB)	< 0.00047	mg/kg	0.0048	0.00047	1		11/04/21 21:48	106-93-4	
1,2-Dichlorobenzene	< 0.00037	mg/kg	0.0048	0.00037	1		11/04/21 21:48		
1.3-Dichlorobenzene	<0.00028	mg/kg	0.0048	0.00028	1		11/04/21 21:48		
1,4-Dichlorobenzene	< 0.00035	mg/kg	0.0048	0.00035	1		11/04/21 21:48		
Dichlorodifluoromethane	<0.00015	mg/kg	0.0048	0.00015	1		11/04/21 21:48		
1,1-Dichloroethane	< 0.00045	mg/kg	0.0048	0.00045	1		11/04/21 21:48		
1,2-Dichloroethane	< 0.00049	mg/kg	0.0048	0.00049	1		11/04/21 21:48		
1,1-Dichloroethene	<0.00054	mg/kg	0.0048	0.00054	1		11/04/21 21:48		
cis-1,2-Dichloroethene	0.0037J	mg/kg	0.0048	0.00046	1		11/04/21 21:48		
trans-1,2-Dichloroethene	<0.00045	mg/kg	0.0048	0.00045	1		11/04/21 21:48		
1,2-Dichloropropane	< 0.00041	mg/kg	0.0048	0.00041	1		11/04/21 21:48		
cis-1,3-Dichloropropene	< 0.00037	mg/kg	0.0048	0.00037	1		11/04/21 21:48		
,			2.00.0		-				



Project: Exide ERT Frankfort Site

Pace Project No.: 50300888

Date: 11/05/2021 04:07 PM

Sample: Soil-1021 Lab ID: 50300888002 Collected: 10/22/21 08:50 Received: 10/22/21 11:40 Matrix: Solid

Results reported on a "dry weight" basis and are adjusted for percent moisture, sample size and any dilutions.

Parameters	Results	Units	PQL	MDL	DF	Prepared	Analyzed	CAS No.	Qual
- raidilleleis	- Nesults	Office				- Frepareu	- Allalyzeu		- Quai
8260 MSV 5035A VOA		Method: EPA							
	Pace Ana	lytical Service	s - Indianap	olis					
trans-1,3-Dichloropropene	< 0.00032	mg/kg	0.0048	0.00032	1		11/04/21 21:48	10061-02-6	
1,4-Dioxane (p-Dioxane)	<0.051	mg/kg	0.48	0.051	1		11/04/21 21:48	123-91-1	
Ethylbenzene	<0.00027	mg/kg	0.0048	0.00027	1		11/04/21 21:48	100-41-4	
2-Hexanone	<0.0011	mg/kg	0.096	0.0011	1		11/04/21 21:48	591-78-6	
Isopropylbenzene (Cumene)	<0.00036	mg/kg	0.0048	0.00036	1		11/04/21 21:48	98-82-8	
Methyl acetate	<0.00080	mg/kg	0.0048	0.00080	1		11/04/21 21:48	79-20-9	N2
Methylcyclohexane	< 0.00034	mg/kg	0.0048	0.00034	1		11/04/21 21:48	108-87-2	N2
Methylene Chloride	< 0.0047	mg/kg	0.019	0.0047	1		11/04/21 21:48	75-09-2	
4-Methyl-2-pentanone (MIBK)	< 0.0016	mg/kg	0.024	0.0016	1		11/04/21 21:48	108-10-1	
Methyl-tert-butyl ether	<0.00027	mg/kg	0.0048	0.00027	1		11/04/21 21:48	1634-04-4	
Styrene	< 0.00034	mg/kg	0.0048	0.00034	1		11/04/21 21:48	100-42-5	
1,1,2,2-Tetrachloroethane	<0.00040	mg/kg	0.0048	0.00040	1		11/04/21 21:48	79-34-5	
Tetrachloroethene	< 0.00036	mg/kg	0.0048	0.00036	1		11/04/21 21:48		
Toluene	< 0.00051	mg/kg	0.0048	0.00051	1		11/04/21 21:48	108-88-3	
1,2,3-Trichlorobenzene	<0.00040	mg/kg	0.0048	0.00040	1		11/04/21 21:48		
1,2,4-Trichlorobenzene	< 0.00039	mg/kg	0.0048	0.00039	1		11/04/21 21:48		
1,1,1-Trichloroethane	< 0.00040	mg/kg	0.0048	0.00040	1		11/04/21 21:48		
1,1,2-Trichloroethane	< 0.00041	mg/kg	0.0048	0.00041	1		11/04/21 21:48		
Trichloroethene	0.00092J	mg/kg	0.0048	0.00043	1		11/04/21 21:48		
Trichlorofluoromethane	< 0.00014	mg/kg	0.0048	0.00014	1		11/04/21 21:48		
1,1,2-Trichlorotrifluoroethane	< 0.00053	mg/kg	0.0048	0.00053	1		11/04/21 21:48		
Vinyl chloride	0.00064J	mg/kg	0.0048	0.000095	1		11/04/21 21:48		
Xylene (Total)	<0.00078	mg/kg	0.0096	0.00078	1		11/04/21 21:48		
Surrogates	10.0001.0	9,9	0.0000	0.000.0	•		,,	.000 _0 .	
Dibromofluoromethane (S)	99	%.	73-132		1		11/04/21 21:48	1868-53-7	
Toluene-d8 (S)	98	%.	66-148		1		11/04/21 21:48	2037-26-5	
4-Bromofluorobenzene (S)	94	%.	40-149		1		11/04/21 21:48	460-00-4	
Percent Moisture	Analytical	Method: SM 2	2540G						
	Pace Ana	lytical Service	s - Indianap	olis					
Percent Moisture	22.2	%	0.10	0.10	1		10/25/21 11:52		N2
1030 Ignitability of Solids	•	Method: 1030 lytical Service		olis					
Ignitability, non-metallic	<2.2 mm/sec	mm/sec	2.2	2.2	1		10/27/21 10:05		N2
9045 pH Soil	•	Method: EPA lytical Service		olis					
pH at 25 Degrees C	6.8	Std. Units	0.10	0.10	1		10/29/21 13:07		НЗ
733C S Reactive Cyanide		Method: EPA lytical Service			od: SW	-846 7.3.3.2			
Cyanide, Reactive	<0.51	mg/kg	1.3	0.51	1	10/27/21 13:27	11/02/21 12:21		



Project: Exide ERT Frankfort Site

Pace Project No.: 50300888

Date: 11/05/2021 04:07 PM

Sample: Soil-1021 Lab ID: 50300888002 Collected: 10/22/21 08:50 Received: 10/22/21 11:40 Matrix: Solid

Results reported on a "dry weight" basis and are adjusted for percent moisture, sample size and any dilutions.

Parameters Results Units **PQL** MDL DF Prepared Analyzed CAS No. Qual 734S Reactive Sulfide Analytical Method: SM 4500-S2-F-2011 Preparation Method: SW-846 7.3.4.2 Pace Analytical Services - Greensburg Sulfide, Reactive <12.9 mg/kg 12.9 12.9 10/29/21 14:32 10/29/21 15:02



Project: Exide ERT Frankfort Site

Pace Project No.: 50300888

QC Batch: 648297 Analysis Method: EPA 7470

QC Batch Method: EPA 7470 Analysis Description: 7470 Mercury TCLP

Laboratory: Pace Analytical Services - Indianapolis

Associated Lab Samples: 50300888001, 50300888002

METHOD BLANK: Matrix: Water

Associated Lab Samples: 50300888001, 50300888002

> Blank Reporting Parameter Units Result Limit MDL Analyzed Qualifiers

< 0.00033 0.00067 0.00033 11/04/21 11:15 Mercury mg/L

LABORATORY CONTROL SAMPLE: 2986871

Spike LCS LCS % Rec % Rec Limits Qualifiers Parameter Units Conc. Result Mercury 0.005 0.0050 101 80-120 mg/L

MATRIX SPIKE SAMPLE: 2986872

MS 50298356057 Spike MS % Rec Parameter Units Result Conc. Result % Rec Limits Qualifiers <0.0010 0.015 Mercury mg/L 0.016 108 75-125

MATRIX SPIKE SAMPLE: 2986873

Date: 11/05/2021 04:07 PM

50300491001 MS MS % Rec Spike % Rec Parameter Units Result Conc. Result Limits Qualifiers

ND Mercury mg/L 0.015 0.016 107 75-125

MATRIX SPIKE & MATRIX SPIKE DUPLICATE: 2986874 2986875

MS MSD MSD 50300888002 Spike Spike MS MS MSD % Rec Max Parameter Units Result Conc. Conc. Result Result % Rec % Rec Limits **RPD** RPD Qual

Mercury <0.0010 0.015 0.015 0.016 0.016 105 106 75-125 0 20 mg/L

MATRIX SPIKE SAMPLE: 2986876

50300927001 MS MS % Rec Spike Parameter Units Result Conc. Result % Rec Limits Qualifiers < 0.040 105 Mercury mg/L 0.3 0.32 75-125

Results presented on this page are in the units indicated by the "Units" column except where an alternate unit is presented to the right of the result.



Project: Exide ERT Frankfort Site

Pace Project No.: 50300888

Date: 11/05/2021 04:07 PM

MATRIX SPIKE SAMPLE:	2986877						
		50301317001	Spike	MS	MS	% Rec	
Parameter	Units	Result	Conc.	Result	% Rec	Limits	Qualifiers
Mercury	mg/L	<0.0020	0.015	0.013	87	75-125	
MATRIX SPIKE SAMPLE:	2986879						
Parameter	Units	50301380001 Result	Spike Conc.	MS Result	MS % Rec	% Rec Limits	Qualifiers
Mercury	mg/L	ND	0.015	0.016	103	75-125	
MATRIX SPIKE SAMPLE:	2986880						
Parameter	Units	50301209001 Result	Spike Conc.	MS Result	MS % Rec	% Rec Limits	Qualifiers
Mercury	mg/L	ND	0.015	0.016	105	75-125	
MATRIX SPIKE SAMPLE:	2986881						
Parameter	Units	50301220001 Result	Spike Conc.	MS Result	MS % Rec	% Rec Limits	Qualifiers
Mercury	mg/L	ND	0.015	0.013	89	75-125	
MATRIX SPIKE SAMPLE:	2986882						
Parameter	Units	50301536002 Result	Spike Conc.	MS Result	MS % Rec	% Rec Limits	Qualifiers
Mercury	mg/L	0.011	0.015	0.027	105	75-125	
MATRIX SPIKE SAMPLE:	2986883						
Parameter	Units	50301362001 Result	Spike Conc.	MS Result	MS % Rec	% Rec Limits	Qualifiers
Mercury	mg/L	ND ND	0.015	0.0079	52	75-125	ИΟ
MATRIX SPIKE SAMPLE:	2986884						
Parameter	Units	50300517005 Result	Spike Conc.	MS Result	MS % Rec	% Rec Limits	Qualifiers
Mercury	mg/L	ND .	0.015	0.016	107	75-125	

Results presented on this page are in the units indicated by the "Units" column except where an alternate unit is presented to the right of the result.



Project: Exide ERT Frankfort Site

Pace Project No.: 50300888

Date: 11/05/2021 04:07 PM

QC Batch: 648095 Analysis Method: EPA 7470

QC Batch Method: EPA 7470 Analysis Description: 7470 Mercury

Laboratory: Pace Analytical Services - Indianapolis

Associated Lab Samples: 50300888001

METHOD BLANK: 2985983 Matrix: Water

Associated Lab Samples: 50300888001

Blank Reporting
Parameter Units Result Limit MDL Analyzed Qualifiers

Mercury ug/L <0.085 2.0 0.085 11/04/21 15:51

LABORATORY CONTROL SAMPLE: 2985984

Spike LCS LCS % Rec Conc. Result % Rec Limits Qualifiers Parameter Units Mercury ug/L 5.2 103 80-120

MATRIX SPIKE & MATRIX SPIKE DUPLICATE: 2985985 2985986

MS MSD

50300876002 Spike Spike MS MSD MS MSD % Rec Max Parameter Units Conc. Conc. Result Result % Rec % Rec Limits **RPD** RPD Qual Result ND 5 100 20 Mercury ug/L 5 5.0 4.9 99 75-125

Results presented on this page are in the units indicated by the "Units" column except where an alternate unit is presented to the right of the result.



Project: Exide ERT Frankfort Site

Pace Project No.: 50300888

Date: 11/05/2021 04:07 PM

QC Batch: 647537 QC Batch Method: EPA 7471 Analysis Method: EPA 7471

Analysis Description: 7471 Mercury

Laboratory:

Pace Analytical Services - Indianapolis

Associated Lab Samples: 50300888002

METHOD BLANK: 2983633 Matrix: Solid

Associated Lab Samples: 50300888002

Blank Reporting

Parameter Units Result Limit MDL Analyzed Qualifiers

Mercury mg/kg <0.025 0.20 0.025 11/01/21 09:38

LABORATORY CONTROL SAMPLE: 2983634

Spike LCS LCS % Rec Conc. Result % Rec Limits Qualifiers Parameter Units Mercury 0.49 0.52 107 80-120 mg/kg

MATRIX SPIKE & MATRIX SPIKE DUPLICATE: 2983635 2983636

MS MSD

50300743001 Spike Spike MS MSD MS MSD % Rec Max Parameter Units Conc. Conc. Result Result % Rec % Rec Limits **RPD** RPD Qual Result ND 0.54 0.61 106 20 Mercury mg/kg 0.59 0.66 105 75-125

Results presented on this page are in the units indicated by the "Units" column except where an alternate unit is presented to the right of the result.



Project: Exide ERT Frankfort Site

Pace Project No.: 50300888

Date: 11/05/2021 04:07 PM

QC Batch: 646594 Analysis Method: EPA 6010
QC Batch Method: EPA 3050 Analysis Description: 6010 MET

Laboratory: Pace Analytical Services - Indianapolis

Associated Lab Samples: 50300888002

METHOD BLANK: 2979584 Matrix: Solid

Associated Lab Samples: 50300888002

Parameter	Units	Blank Result	Reporting Limit	MDL	Analyzed	Qualifiers
Arsenic	 mg/kg	<0.19	0.91	0.19	10/27/21 11:44	
Barium	mg/kg	0.041J	0.91	0.032	10/27/21 11:44	
Cadmium	mg/kg	< 0.013	0.45	0.013	10/27/21 11:44	
Chromium	mg/kg	< 0.067	0.91	0.067	10/27/21 11:44	
Lead	mg/kg	< 0.094	0.91	0.094	10/27/21 11:44	
Selenium	mg/kg	<0.24	0.91	0.24	10/27/21 11:44	
Silver	mg/kg	<0.16	0.45	0.16	10/27/21 11:44	

LABORATORY CONTROL SAMPLE:	2979585					
		Spike	LCS	LCS	% Rec	
Parameter	Units	Conc.	Result	% Rec	Limits	Qualifiers
Arsenic	mg/kg	47.8	44.4	93	80-120	
Barium	mg/kg	47.8	46.2	97	80-120	
Cadmium	mg/kg	47.8	43.2	90	80-120	
hromium	mg/kg	47.8	44.3	93	80-120	
ead	mg/kg	47.8	42.6	89	80-120	
Selenium	mg/kg	47.8	43.6	91	80-120	
lilver	mg/kg	23.9	21.1	88	80-120	

MATRIX SPIKE & MATRIX	SPIKE DUPL	ICATE: 2979		MOD	2979587							
		50300963003	MS Spike	MSD Spike	MS	MSD	MS	MSD	% Rec		Max	
Parameter	Units	Result	Conc.	Conc.	Result	Result	% Rec	% Rec	Limits	RPD	RPD	Qual
Arsenic	mg/kg	5.3	48.7	46.3	53.6	53.8	99	105	75-125	0	20	
Barium	mg/kg	35.0	48.7	46.3	83.2	87.8	99	114	75-125	5	20	
Cadmium	mg/kg	ND	48.7	46.3	46.2	46.6	95	100	75-125	1	20	
Chromium	mg/kg	7.3	48.7	46.3	49.6	52.3	87	97	75-125	5	20	
Lead	mg/kg	17.9	48.7	46.3	58.6	61.7	84	95	75-125	5	20	
Selenium	mg/kg	ND	48.7	46.3	45.0	46.0	92	99	75-125	2	20	
Silver	mg/kg	ND	24.3	23.2	22.8	23.0	94	99	75-125	1	20	

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Project: Exide ERT Frankfort Site

Pace Project No.: 50300888

Date: 11/05/2021 04:07 PM

QC Batch: 648249 Analysis Method: EPA 6010
QC Batch Method: EPA 3010 Analysis Description: 6010 MET TCLP

Laboratory: Pace Analytical Services - Indianapolis

Associated Lab Samples: 50300888001, 50300888002

METHOD BLANK: 2986644 Matrix: Water

Associated Lab Samples: 50300888001, 50300888002

Parameter	Units	Blank Result	Reporting Limit	MDL	Analyzed	Qualifiers
Arsenic	mg/L	<0.0050	0.010	0.0050	11/03/21 23:47	
Barium	mg/L	< 0.025	0.50	0.025	11/03/21 23:47	
Cadmium	mg/L	< 0.0025	0.0050	0.0025	11/03/21 23:47	
Chromium	mg/L	< 0.0052	0.010	0.0052	11/03/21 23:47	
Lead	mg/L	< 0.0050	0.010	0.0050	11/03/21 23:47	
Selenium	mg/L	< 0.0050	0.010	0.0050	11/03/21 23:47	
Silver	mg/L	< 0.0050	0.010	0.0050	11/03/21 23:47	

LABORATORY CONTROL SAMPLE:	2986645					
		Spike	LCS	LCS	% Rec	
Parameter	Units	Conc.	Result	% Rec	Limits	Qualifiers
Arsenic	mg/L		1.0	100	80-120	
Barium	mg/L	1	0.99	99	80-120	
Cadmium	mg/L	1	0.99	99	80-120	
Chromium	mg/L	1	1.0	100	80-120	
Lead	mg/L	1	0.98	98	80-120	
Selenium	mg/L	1	0.99	99	80-120	
Silver	mg/L	0.5	0.47	94	80-120	

MATRIX SPIKE & MATRIX	SPIKE DUPLI	CATE: 2986			2986647							
Parameter	Units	50300491001 Result	MS Spike Conc.	MSD Spike Conc.	MS Result	MSD Result	MS % Rec	MSD % Rec	% Rec Limits	RPD	Max RPD	Qual
Arsenic	mg/L	ND	10	10	10.2	10.0	102	100	50-150	2	20	
Barium	mg/L	ND	10	10	11.5	11.2	98	95	50-150	2	20	
Cadmium	mg/L	0.10	10	10	10	9.8	99	97	50-150	2	20	
Chromium	mg/L	ND	10	10	10	9.7	99	97	50-150	2	20	
Lead	mg/L	5.1	10	10	14.5	14.2	94	91	50-150	2	20	
Selenium	mg/L	ND	10	10	10.1	9.9	101	99	50-150	2	20	
Silver	mg/L	ND	5	5	4.6	4.5	91	90	50-150	2	20	

MATRIX SPIKE SAMPLE:	2986648						
		50300844003	Spike	MS	MS	% Rec	
Parameter	Units	Result	Conc.	Result	% Rec	Limits	Qualifiers
Arsenic	 mg/L	ND ND	10	10.2	102	50-150	
Barium	mg/L	ND	10	9.9	98	50-150	

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Project: Exide ERT Frankfort Site

Pace Project No.: 50300888

Date: 11/05/2021 04:07 PM

MATRIX SPIKE SAMPLE:	2986648						
	11.7	50300844003	Spike	MS	MS	% Rec	0 ""
Parameter	Units	Result	Conc.	Result	% Rec	Limits	Qualifiers
Cadmium	mg/L	ND	10	10	100	50-150	
Chromium	mg/L	0.97	10	10.8	99	50-150	
Lead	mg/L	ND	10	9.4	94	50-150	
Selenium	mg/L	ND	10	10.0	100	50-150	
Silver	mg/L	ND	5	4.7	94	50-150	
MATRIX SPIKE SAMPLE:	2986649						
		50300888001	Spike	MS	MS	% Rec	
Parameter	Units	Result	Conc.	Result	% Rec	Limits	Qualifiers
Arsenic	 mg/L	<0.050	10	9.5	95	50-150	
Barium	mg/L	<0.25	10	9.6	96	50-150	
Cadmium	mg/L	<0.025	10	9.4	94	50-150	
Chromium	mg/L	< 0.052	10	9.6	95	50-150	
_ead	mg/L	< 0.050	10	9.2	92	50-150	
Selenium	mg/L	< 0.050	10	9.5	95	50-150	
Silver	mg/L	<0.050	5	4.5	89	50-150	
MATRIX SPIKE SAMPLE:	2986650						
		50300836001	Spike	MS	MS	% Rec	
Parameter	Units	Result	Conc.	Result	% Rec	Limits	Qualifiers
Arsenic	 mg/L	ND ND	10	10.1	101	50-150	
3arium	mg/L	ND	10	10.9	96	50-150	
Cadmium	mg/L	ND	10	9.7	97	50-150	
Chromium	mg/L	ND	10	9.6	95	50-150	
_ead	mg/L	ND	10	9.0	90	50-150	
Selenium	mg/L	ND	10	10	99	50-150	
Silver	mg/L	ND	5	4.5	90	50-150	
MATRIX SPIKE SAMPLE:	2986651						
- -		50300963001	Spike	MS	MS	% Rec	
Parameter	Units	Result	Conc.	Result	% Rec	Limits	Qualifiers
Arsenic	mg/L	ND	10	10.1	101	50-150	
Barium	mg/L	ND	10	10.9	96	50-150	
Cadmium	mg/L	ND	10	9.7	97	50-150	
Chromium	mg/L	ND	10	9.8	98	50-150	
_ead	mg/L	ND	10	9.2	91	50-150	
Selenium	mg/L	ND	10	10	100	50-150	
Silver	mg/L	ND	5	4.6	91	50-150	

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Project: Exide ERT Frankfort Site

Pace Project No.: 50300888

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Parameter Arsenic Barium Cadmium Chromium Lead Selenium Silver MATRIX SPIKE SAMPLE: Parameter Arsenic Barium Cadmium Chromium	Units mg/L mg/L mg/L mg/L mg/L mg/L mg/L mg/	50301209001 Result	Spike Conc. 10 10 10 10 10 5 Spike	MS Result 9.9 9.5 9.5 9.6 9.0 9.8 4.5	MS % Rec 98 95 95 96 90 98 90	% Rec Limits 50-150 50-150 50-150 50-150 50-150 50-150	Qualifiers
Arsenic Barium Cadmium Chromium Lead Selenium Silver MATRIX SPIKE SAMPLE: Parameter Arsenic Barium Cadmium	mg/L mg/L mg/L mg/L mg/L mg/L mg/L mg/L	ND ND ND ND ND ND ND	10 10 10 10 10 10 5 Spike	9.9 9.5 9.5 9.6 9.0 9.8 4.5	98 95 95 96 90 98 90	50-150 50-150 50-150 50-150 50-150 50-150	Qualifiers
Barium Cadmium Chromium Lead Selenium Silver MATRIX SPIKE SAMPLE: Parameter Arsenic Barium Cadmium	mg/L mg/L mg/L mg/L mg/L mg/L mg/L mg/L	ND ND ND ND ND ND	10 10 10 10 10 5 Spike	9.5 9.5 9.6 9.0 9.8 4.5	95 95 96 90 98 90	50-150 50-150 50-150 50-150 50-150 50-150	
Cadmium Chromium Lead Gelenium Silver MATRIX SPIKE SAMPLE: Parameter Arsenic Barium Cadmium	mg/L mg/L mg/L mg/L mg/L mg/L mg/L mg/L	ND ND ND ND ND	10 10 10 10 10 5 Spike	9.5 9.6 9.0 9.8 4.5	95 96 90 98 90	50-150 50-150 50-150 50-150 50-150	
Cadmium Chromium Lead Selenium Silver MATRIX SPIKE SAMPLE: Parameter Arsenic Sarium Cadmium	mg/L mg/L mg/L mg/L mg/L mg/L mg/L	ND ND ND ND	10 10 10 10 5 Spike	9.5 9.6 9.0 9.8 4.5	95 96 90 98 90	50-150 50-150 50-150 50-150	
Chromium Lead Selenium Silver MATRIX SPIKE SAMPLE: Parameter Arsenic Barium Cadmium	mg/L mg/L mg/L mg/L 2986653 Units mg/L	ND ND ND ND	10 10 10 5 5	9.6 9.0 9.8 4.5	96 90 98 90	50-150 50-150 50-150 50-150	
Lead Selenium Silver MATRIX SPIKE SAMPLE: Parameter Arsenic Barium Cadmium	mg/L mg/L mg/L 2986653 Units mg/L	ND ND 50301209001	10 10 5 Spike	9.0 9.8 4.5	90 98 90	50-150 50-150 50-150	
Selenium Silver MATRIX SPIKE SAMPLE: Parameter Arsenic Barium Cadmium	mg/L mg/L 2986653 Units mg/L	ND ND 50301209001	10 5 Spike	9.8 4.5	98 90	50-150 50-150	
MATRIX SPIKE SAMPLE: Parameter Arsenic Barium Cadmium	2986653 Units mg/L	ND 50301209001	5 Spike	4.5	90	50-150	
Parameter Arsenic Barium Cadmium	Units mg/L			MS	MS		
Parameter Arsenic Barium Cadmium	Units mg/L			MS	MC		
Arsenic Barium Cadmium	mg/L				IVIO	% Rec	
Arsenic Barium Cadmium	mg/L		Conc.	Result	% Rec	Limits	Qualifiers
Barium Cadmium	_	ND	10	9.7	97	50-150	
Cadmium	man	ND ND	10	10.1	97 95	50-150	
	mg/L mg/L	ND ND	10	9.4	95 94	50-150 50-150	
inomium		ND ND					
	mg/L	ND ND	10	9.4	94	50-150	
_ead	mg/L		10	8.9	89	50-150	
Selenium	mg/L	ND	10	9.6	96	50-150	
Silver	mg/L	ND	5	4.5	89	50-150	
MATRIX SPIKE SAMPLE:	2986654						
		50301220001	Spike	MS	MS	% Rec	
Parameter	Units	Result	Conc.	Result	% Rec	Limits	Qualifiers
Arsenic	mg/L	ND	10	10.0	100	50-150	
Barium	mg/L	ND	10	9.6	95	50-150	
Cadmium	mg/L	ND	10	9.6	96	50-150	
Chromium	mg/L	ND	10	9.7	97	50-150	
Lead	mg/L	ND	10	9.1	91	50-150	
Selenium	mg/L	ND	10	10.1	101	50-150	
Silver	mg/L	ND	5	4.5	90	50-150	
MATRIX SPIKE SAMPLE:	2986655						
		50301536001	Spike	MS	MS	% Rec	
Parameter	Units	Result	Conc.	Result	% Rec	Limits	Qualifiers
Arsenic	mg/L		10	9.9	98	50-150	
Barium	mg/L	ND	10	9.6	96	50-150	
Cadmium	mg/L	ND	10	9.5	95	50-150	
Chromium	mg/L	ND	10	9.6	96	50-150	
-ead	mg/L	ND	10	9.1	91	50-150	
Selenium	mg/L	ND	10	9.1	98	50-150	
Silver	mg/L	ND ND	5	9.6 4.5	96 91	50-150 50-150	

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REPORT OF LABORATORY ANALYSIS

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Project: Exide ERT Frankfort Site

Pace Project No.: 50300888

Date: 11/05/2021 04:07 PM

MATRIX SPIKE SAMPLE:	2986656						
		50301362001	Spike	MS	MS	% Rec	
Parameter	Units	Result	Conc.	Result	% Rec	Limits	Qualifiers
Arsenic	mg/L	ND	10	10.0	100	50-150	
Barium	mg/L	ND	10	11.0	94	50-150	
Cadmium	mg/L	ND	10	9.5	95	50-150	
Chromium	mg/L	ND	10	9.6	95	50-150	
Lead	mg/L	47.0	10	54.8	78	50-150	
Selenium	mg/L	ND	10	9.8	98	50-150	
Silver	mg/L	ND	5	4.4	88	50-150	

MATRIX SPIKE SAMPLE:	2986657						
Parameter	Units	50300517005 Result	Spike Conc.	MS Result	MS % Rec	% Rec Limits	Qualifiers
							Qualificity
Arsenic	mg/L	ND	10	9.9	99	50-150	
Barium	mg/L	ND	10	10.4	95	50-150	
Cadmium	mg/L	ND	10	9.5	95	50-150	
Chromium	mg/L	ND	10	9.5	95	50-150	
Lead	mg/L	ND	10	8.9	89	50-150	
Selenium	mg/L	ND	10	9.8	98	50-150	
Silver	mg/L	ND	5	4.5	89	50-150	

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Project: Exide ERT Frankfort Site

Pace Project No.: 50300888

QC Batch: 646674
QC Batch Method: EPA 3010

Analysis Method: EPA 6010

Analysis Description: Laboratory: 6010 MET
Pace Analytical Services - Indianapolis

Associated Lab Samples: 50300888001

METHOD BLANK: 2979830

Matrix: Water

Associated Lab Samples: 50300888001

LABORATORY CONTROL SAMPLE:

Date: 11/05/2021 04:07 PM

Parameter	Units	Blank Result	Reporting Limit	MDL	Analyzed	Qualifiers
Arsenic	ug/L	<2.6	10.0	2.6	10/27/21 16:28	
Barium	ug/L	< 0.79	10.0	0.79	10/27/21 16:28	
Cadmium	ug/L	< 0.41	2.0	0.41	10/27/21 16:28	
Chromium	ug/L	<1.9	10.0	1.9	10/27/21 16:28	
Lead	ug/L	<3.5	10.0	3.5	10/27/21 16:28	
Selenium	ug/L	<4.5	10.0	4.5	10/27/21 16:28	
Silver	ua/L	<1.4	10.0	1.4	10/27/21 16:28	

Parameter	Units	Spike Conc.	LCS Result	LCS % Rec	% Rec Limits	Qualifiers
Arsenic	ug/L	1000	936	94	80-120	
Barium	ug/L	1000	953	95	80-120	
Cadmium	ug/L	1000	906	91	80-120	
Chromium	ua/L	1000	940	94	80-120	

80-120 Lead ug/L 1000 891 89 Selenium ug/L 1000 912 91 80-120 Silver ug/L 500 464 93 80-120

2979831

MATRIX SPIKE & MATRIX	SPIKE DUPL	ICATE: 2979	832 MS	MSD	2979833							
		50300796003	Spike	Spike	MS	MSD	MS	MSD	% Rec		Max	
Parameter	Units	Result	Conc.	Conc.	Result	Result	% Rec	% Rec	Limits	RPD	RPD	Qual
Arsenic	ug/L	ND	1000	1000	1000	984	100	98	75-125	2	20	
Barium	ug/L	12.0	1000	1000	1010	988	99	98	75-125	2	20	
Cadmium	ug/L	0.54J	1000	1000	944	932	94	93	75-125	1	20	
Chromium	ug/L	ND	1000	1000	948	936	95	94	75-125	1	20	
Lead	ug/L	ND	1000	1000	874	862	87	86	75-125	1	20	
Selenium	ug/L	ND	1000	1000	958	943	96	94	75-125	2	20	
Silver	ug/L	ND	500	500	485	473	97	95	75-125	3	20	

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Project: Exide ERT Frankfort Site

Pace Project No.: 50300888

Date: 11/05/2021 04:07 PM

QC Batch: 648494 Analysis Method: EPA 5030/8260
QC Batch Method: EPA 5030/8260 Analysis Description: 8260 MSV TCLP

Laboratory: Pace Analytical Services - Indianapolis

Associated Lab Samples: 50300888001, 50300888002

METHOD BLANK: 2987966 Matrix: Water

Associated Lab Samples: 50300888001, 50300888002

_		Blank	Reporting			
Parameter	Units	Result	Limit	MDL	Analyzed	Qualifiers
1,1-Dichloroethene	mg/L	< 0.025	0.050	0.025	11/04/21 03:03	
1,2-Dichloroethane	mg/L	< 0.025	0.050	0.025	11/04/21 03:03	
2-Butanone (MEK)	mg/L	< 0.50	1.0	0.50	11/04/21 03:03	
Benzene	mg/L	< 0.010	0.050	0.010	11/04/21 03:03	
Carbon tetrachloride	mg/L	< 0.025	0.050	0.025	11/04/21 03:03	
Chlorobenzene	mg/L	< 0.025	0.050	0.025	11/04/21 03:03	
Chloroform	mg/L	< 0.025	0.050	0.025	11/04/21 03:03	
Tetrachloroethene	mg/L	< 0.025	0.050	0.025	11/04/21 03:03	
Trichloroethene	mg/L	< 0.025	0.050	0.025	11/04/21 03:03	
Vinyl chloride	mg/L	< 0.010	0.020	0.010	11/04/21 03:03	
4-Bromofluorobenzene (S)	%.	91	78-117		11/04/21 03:03	
Dibromofluoromethane (S)	%.	106	78-120		11/04/21 03:03	
Toluene-d8 (S)	%.	100	77-118		11/04/21 03:03	

LABORATORY CONTROL SAMPLE:	2987967					
		Spike	LCS	LCS	% Rec	
Parameter	Units	Conc.	Result	% Rec	Limits	Qualifiers
1,1-Dichloroethene	mg/L	0.5	0.48	97	67-136	
1,2-Dichloroethane	mg/L	0.5	0.49	97	69-135	
2-Butanone (MEK)	mg/L	2.5	2.1	85	56-164	
Benzene	mg/L	0.5	0.43	85	77-128	
Carbon tetrachloride	mg/L	0.5	0.51	102	61-139	
Chlorobenzene	mg/L	0.5	0.44	89	76-124	
Chloroform	mg/L	0.5	0.47	94	77-120	
Tetrachloroethene	mg/L	0.5	0.47	94	70-124	
Trichloroethene	mg/L	0.5	0.44	89	75-130	
Vinyl chloride	mg/L	0.5	0.51	103	51-140	
4-Bromofluorobenzene (S)	%.			93	78-117	
Dibromofluoromethane (S)	%.			108	78-120	
Toluene-d8 (S)	%.			98	77-118	

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Project: Exide ERT Frankfort Site

Pace Project No.: 50300888

Date: 11/05/2021 04:07 PM

QC Batch: 647925 Analysis Method: EPA 8260
QC Batch Method: EPA 8260 Analysis Description: 8260 MSV

Laboratory: Pace Analytical Services - Indianapolis

Associated Lab Samples: 50300888001

METHOD BLANK: 2985354 Matrix: Water

Associated Lab Samples: 50300888001

		Blank	Reporting			
Parameter	Units	Result	Limit	MDL	Analyzed	Qualifiers
1,1,1-Trichloroethane	ug/L	<0.40	5.0	0.40	11/01/21 11:41	
1,1,2,2-Tetrachloroethane	ug/L	<0.26	5.0	0.26	11/01/21 11:41	
1,1,2-Trichloroethane	ug/L	< 0.30	5.0	0.30	11/01/21 11:41	
1,1,2-Trichlorotrifluoroethane	ug/L	< 0.49	5.0	0.49	11/01/21 11:41	
1,1-Dichloroethane	ug/L	<0.41	5.0	0.41	11/01/21 11:41	
1,1-Dichloroethene	ug/L	< 0.37	5.0	0.37	11/01/21 11:41	
1,2,3-Trichlorobenzene	ug/L	< 0.50	5.0	0.50	11/01/21 11:41	
1,2,4-Trichlorobenzene	ug/L	< 0.44	5.0	0.44	11/01/21 11:41	
1,2-Dibromo-3-chloropropane	ug/L	<1.6	10.0	1.6	11/01/21 11:41	
1,2-Dibromoethane (EDB)	ug/L	< 0.42	5.0	0.42	11/01/21 11:41	
1,2-Dichlorobenzene	ug/L	< 0.30	5.0	0.30	11/01/21 11:41	
1,2-Dichloroethane	ug/L	<0.41	5.0	0.41	11/01/21 11:41	
1,2-Dichloropropane	ug/L	< 0.35	5.0	0.35	11/01/21 11:41	
1,3-Dichlorobenzene	ug/L	< 0.30	5.0	0.30	11/01/21 11:41	
1,4-Dichlorobenzene	ug/L	< 0.36	5.0	0.36	11/01/21 11:41	
2-Butanone (MEK)	ug/L	<2.1	25.0	2.1	11/01/21 11:41	
2-Hexanone	ug/L	<1.4	25.0	1.4	11/01/21 11:41	
4-Methyl-2-pentanone (MIBK)	ug/L	<1.4	25.0	1.4	11/01/21 11:41	
Acetone	ug/L	<5.5	100	5.5	11/01/21 11:41	
Benzene	ug/L	<0.31	5.0	0.31	11/01/21 11:41	
Bromochloromethane	ug/L	< 0.42	5.0	0.42	11/01/21 11:41	
Bromodichloromethane	ug/L	<0.29	5.0	0.29	11/01/21 11:41	
Bromoform	ug/L	< 0.42	5.0	0.42	11/01/21 11:41	
Bromomethane	ug/L	<1.6	5.0	1.6	11/01/21 11:41	
Carbon disulfide	ug/L	< 0.32	10.0	0.32	11/01/21 11:41	
Carbon tetrachloride	ug/L	<0.48	5.0	0.48	11/01/21 11:41	
Chlorobenzene	ug/L	< 0.33	5.0	0.33	11/01/21 11:41	
Chloroethane	ug/L	<1.7	5.0	1.7	11/01/21 11:41	
Chloroform	ug/L	< 0.34	5.0	0.34	11/01/21 11:41	
Chloromethane	ug/L	<0.48	5.0	0.48	11/01/21 11:41	
cis-1,2-Dichloroethene	ug/L	<0.46	5.0	0.46	11/01/21 11:41	
cis-1,3-Dichloropropene	ug/L	< 0.34	5.0	0.34	11/01/21 11:41	
Cyclohexane	ug/L	< 0.36	100	0.36	11/01/21 11:41	
Dibromochloromethane	ug/L	< 0.34	5.0	0.34	11/01/21 11:41	
Dichlorodifluoromethane	ug/L	<1.7	5.0	1.7	11/01/21 11:41	
Ethylbenzene	ug/L	<0.26	5.0	0.26	11/01/21 11:41	
Isopropylbenzene (Cumene)	ug/L	< 0.34	5.0	0.34	11/01/21 11:41	
Methyl acetate	ug/L	<0.76	50.0	0.76	11/01/21 11:41	
Methyl-tert-butyl ether	ug/L	<0.31	4.0	0.31	11/01/21 11:41	
Methylcyclohexane	ug/L	< 0.36	50.0	0.36	11/01/21 11:41	

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Project: Exide ERT Frankfort Site

Pace Project No.: 50300888

Date: 11/05/2021 04:07 PM

METHOD BLANK: 2985354 Matrix: Water

Associated Lab Samples: 50300888001

		Blank	Reporting			
Parameter	Units	Result	Limit	MDL	Analyzed	Qualifiers
Methylene Chloride	ug/L	<0.081	5.0	0.081	11/01/21 11:41	
Styrene	ug/L	<0.26	5.0	0.26	11/01/21 11:41	
Tetrachloroethene	ug/L	< 0.44	5.0	0.44	11/01/21 11:41	
Toluene	ug/L	<0.27	5.0	0.27	11/01/21 11:41	
trans-1,2-Dichloroethene	ug/L	< 0.32	5.0	0.32	11/01/21 11:41	
trans-1,3-Dichloropropene	ug/L	<0.27	5.0	0.27	11/01/21 11:41	
Trichloroethene	ug/L	< 0.46	5.0	0.46	11/01/21 11:41	
Trichlorofluoromethane	ug/L	< 0.24	5.0	0.24	11/01/21 11:41	
Vinyl chloride	ug/L	<0.28	2.0	0.28	11/01/21 11:41	
Xylene (Total)	ug/L	<0.68	10.0	0.68	11/01/21 11:41	
4-Bromofluorobenzene (S)	%.	96	78-117		11/01/21 11:41	
Dibromofluoromethane (S)	%.	105	78-120		11/01/21 11:41	
Toluene-d8 (S)	%.	99	77-118		11/01/21 11:41	

LABORATORY CONTROL SAMPLE:	2985355					
		Spike	LCS	LCS	% Rec	
Parameter	Units	Conc.	Result	% Rec	Limits	Qualifiers
1,1,1-Trichloroethane	ug/L	50	53.9	108	73-132	
1,1,2,2-Tetrachloroethane	ug/L	50	52.9	106	65-131	
1,1,2-Trichloroethane	ug/L	50	52.5	105	74-127	
1,1,2-Trichlorotrifluoroethane	ug/L	50	60.8	122	70-133	
1,1-Dichloroethane	ug/L	50	48.7	97	73-133	
1,1-Dichloroethene	ug/L	50	54.7	109	67-136	
1,2,3-Trichlorobenzene	ug/L	50	58.6	117	58-136	
1,2,4-Trichlorobenzene	ug/L	50	59.6	119	48-149	
1,2-Dibromo-3-chloropropane	ug/L	50	56.0	112	71-133	
1,2-Dibromoethane (EDB)	ug/L	50	57.0	114	76-126	
1,2-Dichlorobenzene	ug/L	50	53.7	107	75-114	
1,2-Dichloroethane	ug/L	50	57.9	116	69-135	
1,2-Dichloropropane	ug/L	50	49.8	100	78-134	
1,3-Dichlorobenzene	ug/L	50	53.9	108	70-119	
1,4-Dichlorobenzene	ug/L	50	53.0	106	69-117	
2-Butanone (MEK)	ug/L	250	278	111	56-164	
2-Hexanone	ug/L	250	270	108	63-137	
4-Methyl-2-pentanone (MIBK)	ug/L	250	280	112	64-134	
Acetone	ug/L	250	290	116	46-140	
Benzene	ug/L	50	47.3	95	77-128	
Bromochloromethane	ug/L	50	51.3	103	71-124	
Bromodichloromethane	ug/L	50	54.1	108	70-124	
Bromoform	ug/L	50	55.0	110	65-116	
Bromomethane	ug/L	50	60.1	120	10-200	
Carbon disulfide	ug/L	50	46.1	92	70-131	
Carbon tetrachloride	ug/L	50	58.3	117	61-139	
Chlorobenzene	ug/L	50	53.4	107	76-124	

Results presented on this page are in the units indicated by the "Units" column except where an alternate unit is presented to the right of the result.



Project: Exide ERT Frankfort Site

Pace Project No.: 50300888

Date: 11/05/2021 04:07 PM

Parameter Units Spike Conc. LCS Result LCS % Rec Limits Qualifier oethane of form ug/L 50 54.6 109 56-142 omethane ug/L 50 51.6 103 77-120 omethane ug/L 50 43.2 86 29-141
oethane ug/L 50 54.6 109 56-142 oform ug/L 50 51.6 103 77-120
oform ug/L 50 51.6 103 77-120
amethons 100/1 50 43.0 96 20.444
omethane ug/L 50 43.2 86 29-141
2-Dichloroethene ug/L 50 53.7 107 72-127
3-Dichloropropene ug/L 50 54.8 110 71-131
hexane ug/L 50 49.6J 99 58-141
mochloromethane ug/L 50 57.8 116 69-132
prodifluoromethane ug/L 50 43.3 87 23-139
penzene ug/L 50 52.9 106 76-119
ppylbenzene (Cumene) ug/L 50 54.5 109 77-128
rl acetate ug/L 250 250 100 33-200
/l-tert-butyl ether ug/L 50 54.3 109 75-129
rlcyclohexane ug/L 50 40.4J 81 71-136
rlene Chloride ug/L 50 47.9 96 72-129
ne ug/L 50 54.0 108 66-123
chloroethene ug/L 50 56.9 114 70-124
ne ug/L 50 52.7 105 72-117
1,2-Dichloroethene ug/L 50 53.0 106 75-133
1,3-Dichloropropene ug/L 50 54.4 109 75-111
oroethene ug/L 50 52.0 104 75-130
orofluoromethane ug/L 50 70.1 140 63-162
chloride ug/L 50 56.8 114 51-140
e (Total) ug/L 150 161 107 73-117
mofluorobenzene (S) %. 99 78-117
mofluoromethane (S) %. 104 78-120
ne-d8 (S) %. 102 77-118

Results presented on this page are in the units indicated by the "Units" column except where an alternate unit is presented to the right of the result.



Project: Exide ERT Frankfort Site

Pace Project No.: 50300888

Date: 11/05/2021 04:07 PM

QC Batch: 648672 Analysis Method: EPA 8260

QC Batch Method: EPA 8260 Analysis Description: 8260 MSV 5035A Volatile Organics

Laboratory: Pace Analytical Services - Indianapolis

Associated Lab Samples: 50300888002

METHOD BLANK: 2988853 Matrix: Solid

Associated Lab Samples: 50300888002

		Blank	Reporting			
Parameter	Units	Result	Limit	MDL	Analyzed	Qualifiers
1,1,1-Trichloroethane	 mg/kg	<0.00042	0.0050	0.00042	11/04/21 13:58	
1,1,2,2-Tetrachloroethane	mg/kg	< 0.00042	0.0050	0.00042	11/04/21 13:58	
1,1,2-Trichloroethane	mg/kg	< 0.00043	0.0050	0.00043	11/04/21 13:58	
1,1,2-Trichlorotrifluoroethane	mg/kg	< 0.00055	0.0050	0.00055	11/04/21 13:58	
1,1-Dichloroethane	mg/kg	< 0.00046	0.0050	0.00046	11/04/21 13:58	
1,1-Dichloroethene	mg/kg	< 0.00056	0.0050	0.00056	11/04/21 13:58	
1,2,3-Trichlorobenzene	mg/kg	0.00097J	0.0050	0.00041	11/04/21 13:58	
1,2,4-Trichlorobenzene	mg/kg	0.00079J	0.0050	0.00041	11/04/21 13:58	
1,2-Dibromo-3-chloropropane	mg/kg	<0.00068	0.010	0.00068	11/04/21 13:58	
1,2-Dibromoethane (EDB)	mg/kg	< 0.00049	0.0050	0.00049	11/04/21 13:58	
1,2-Dichlorobenzene	mg/kg	< 0.00039	0.0050	0.00039	11/04/21 13:58	
1,2-Dichloroethane	mg/kg	< 0.00051	0.0050	0.00051	11/04/21 13:58	
1,2-Dichloropropane	mg/kg	< 0.00043	0.0050	0.00043	11/04/21 13:58	
1,3-Dichlorobenzene	mg/kg	< 0.00029	0.0050	0.00029	11/04/21 13:58	
1,4-Dichlorobenzene	mg/kg	< 0.00036	0.0050	0.00036	11/04/21 13:58	
1,4-Dioxane (p-Dioxane)	mg/kg	< 0.053	0.50	0.053	11/04/21 13:58	
2-Butanone (MEK)	mg/kg	<0.0068	0.025	0.0068	11/04/21 13:58	
2-Hexanone	mg/kg	< 0.0012	0.10	0.0012	11/04/21 13:58	
4-Methyl-2-pentanone (MIBK)	mg/kg	< 0.0016	0.025	0.0016	11/04/21 13:58	
Acetone	mg/kg	< 0.0021	0.10	0.0021	11/04/21 13:58	
Benzene	mg/kg	< 0.00040	0.0050	0.00040	11/04/21 13:58	
Bromochloromethane	mg/kg	< 0.00056	0.0050	0.00056	11/04/21 13:58	
Bromodichloromethane	mg/kg	<0.00038	0.0050	0.00038	11/04/21 13:58	
Bromoform	mg/kg	<0.00038	0.0050	0.00038	11/04/21 13:58	
Bromomethane	mg/kg	< 0.00030	0.0050	0.00030	11/04/21 13:58	
Carbon disulfide	mg/kg	< 0.00059	0.010	0.00059	11/04/21 13:58	
Carbon tetrachloride	mg/kg	< 0.00035	0.0050	0.00035	11/04/21 13:58	
Chlorobenzene	mg/kg	< 0.00037	0.0050	0.00037	11/04/21 13:58	
Chloroethane	mg/kg	< 0.00022	0.0050	0.00022	11/04/21 13:58	
Chloroform	mg/kg	0.0015J	0.0050	0.00047	11/04/21 13:58	
Chloromethane	mg/kg	<0.00018	0.0050	0.00018	11/04/21 13:58	
cis-1,2-Dichloroethene	mg/kg	<0.00048	0.0050	0.00048	11/04/21 13:58	
cis-1,3-Dichloropropene	mg/kg	<0.00038	0.0050	0.00038	11/04/21 13:58	
Cyclohexane	mg/kg	< 0.00044	0.10	0.00044	11/04/21 13:58	N2
Dibromochloromethane	mg/kg	< 0.00037	0.0050	0.00037	11/04/21 13:58	
Dichlorodifluoromethane	mg/kg	< 0.00016	0.0050	0.00016	11/04/21 13:58	
Ethylbenzene	mg/kg	<0.00028	0.0050	0.00028	11/04/21 13:58	
Isopropylbenzene (Cumene)	mg/kg	< 0.00037	0.0050	0.00037	11/04/21 13:58	
Methyl acetate	mg/kg	< 0.00084	0.0050	0.00084	11/04/21 13:58	N2
Methyl-tert-butyl ether	mg/kg	<0.00028	0.0050	0.00028	11/04/21 13:58	

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Project: Exide ERT Frankfort Site

Pace Project No.: 50300888

Date: 11/05/2021 04:07 PM

METHOD BLANK: 2988853 Matrix: Solid

Associated Lab Samples: 50300888002

		Blank	Reporting			
Parameter	Units	Result	Limit	MDL	Analyzed	Qualifiers
Methylcyclohexane	mg/kg	<0.00036	0.0050	0.00036	11/04/21 13:58	N2
Methylene Chloride	mg/kg	< 0.0049	0.020	0.0049	11/04/21 13:58	
Styrene	mg/kg	< 0.00035	0.0050	0.00035	11/04/21 13:58	
Tetrachloroethene	mg/kg	< 0.00037	0.0050	0.00037	11/04/21 13:58	
Toluene	mg/kg	< 0.00053	0.0050	0.00053	11/04/21 13:58	
trans-1,2-Dichloroethene	mg/kg	< 0.00047	0.0050	0.00047	11/04/21 13:58	
trans-1,3-Dichloropropene	mg/kg	< 0.00033	0.0050	0.00033	11/04/21 13:58	
Trichloroethene	mg/kg	< 0.00045	0.0050	0.00045	11/04/21 13:58	
Trichlorofluoromethane	mg/kg	< 0.00014	0.0050	0.00014	11/04/21 13:58	
Vinyl chloride	mg/kg	< 0.000099	0.0050	0.000099	11/04/21 13:58	
Xylene (Total)	mg/kg	<0.00081	0.010	0.00081	11/04/21 13:58	
4-Bromofluorobenzene (S)	%.	97	40-149		11/04/21 13:58	
Dibromofluoromethane (S)	%.	100	73-132		11/04/21 13:58	
Toluene-d8 (S)	%.	97	66-148		11/04/21 13:58	

LABORATORY CONTROL SAMPLE:	2988854					
		Spike	LCS	LCS	% Rec	
Parameter	Units	Conc.	Result	% Rec	Limits	Qualifiers
1,1,1-Trichloroethane	mg/kg	0.05	0.041	83	68-129	
1,1,2,2-Tetrachloroethane	mg/kg	0.05	0.044	88	67-137	
1,1,2-Trichloroethane	mg/kg	0.05	0.044	87	68-137	
1,1,2-Trichlorotrifluoroethane	mg/kg	0.05	0.041	83	76-135	
1,1-Dichloroethane	mg/kg	0.05	0.042	84	69-126	
1,1-Dichloroethene	mg/kg	0.05	0.043	86	53-135	
1,2,3-Trichlorobenzene	mg/kg	0.05	0.043	86	57-117	
1,2,4-Trichlorobenzene	mg/kg	0.05	0.044	88	46-134	
1,2-Dibromo-3-chloropropane	mg/kg	0.05	0.045	89	65-132	
1,2-Dibromoethane (EDB)	mg/kg	0.05	0.042	85	68-125	
1,2-Dichlorobenzene	mg/kg	0.05	0.042	85	63-122	
1,2-Dichloroethane	mg/kg	0.05	0.043	85	69-128	
1,2-Dichloropropane	mg/kg	0.05	0.041	82	70-130	
1,3-Dichlorobenzene	mg/kg	0.05	0.042	85	61-121	
1,4-Dichlorobenzene	mg/kg	0.05	0.042	85	59-117	
1,4-Dioxane (p-Dioxane)	mg/kg	0.25	0.24J	95	53-151	
2-Butanone (MEK)	mg/kg	0.25	0.21	83	57-149	
2-Hexanone	mg/kg	0.25	0.21	83	54-140	
4-Methyl-2-pentanone (MIBK)	mg/kg	0.25	0.22	86	65-150	
Acetone	mg/kg	0.25	0.20	82	48-151	
Benzene	mg/kg	0.05	0.043	87	69-125	
Bromochloromethane	mg/kg	0.05	0.043	86	64-136	
Bromodichloromethane	mg/kg	0.05	0.044	87	70-124	
Bromoform	mg/kg	0.05	0.044	87	61-119	
Bromomethane	mg/kg	0.05	0.040	80	15-185	
Carbon disulfide	mg/kg	0.05	0.040	81	52-125	

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Project: Exide ERT Frankfort Site

Pace Project No.: 50300888

Date: 11/05/2021 04:07 PM

LABORATORY CONTROL SAMPLE:	2988854					
		Spike	LCS	LCS	% Rec	
Parameter	Units	Conc.	Result	% Rec	Limits	Qualifiers
Carbon tetrachloride	mg/kg	0.05	0.043	87	65-129	
Chlorobenzene	mg/kg	0.05	0.042	83	66-121	
Chloroethane	mg/kg	0.05	0.032	63	50-146	
Chloroform	mg/kg	0.05	0.039	77	66-123	
Chloromethane	mg/kg	0.05	0.029	58	22-144	
cis-1,2-Dichloroethene	mg/kg	0.05	0.042	83	67-122	
cis-1,3-Dichloropropene	mg/kg	0.05	0.046	92	68-136	
Cyclohexane	mg/kg	0.05	0.042J	84	48-136	N2
Dibromochloromethane	mg/kg	0.05	0.045	89	69-129	
Dichlorodifluoromethane	mg/kg	0.05	0.022	43	10-161	
Ethylbenzene	mg/kg	0.05	0.042	84	57-126	
Isopropylbenzene (Cumene)	mg/kg	0.05	0.042	83	62-132	
Methyl acetate	mg/kg	0.25	0.20	80	49-200	N2
Methyl-tert-butyl ether	mg/kg	0.05	0.041	82	66-136	
Methylcyclohexane	mg/kg	0.05	0.041	81	52-121	N2
Methylene Chloride	mg/kg	0.05	0.044	87	59-148	
Styrene	mg/kg	0.05	0.042	84	67-125	
Tetrachloroethene	mg/kg	0.05	0.043	86	61-123	
Toluene	mg/kg	0.05	0.042	83	67-128	
trans-1,2-Dichloroethene	mg/kg	0.05	0.042	84	61-127	
trans-1,3-Dichloropropene	mg/kg	0.05	0.045	91	69-131	
Trichloroethene	mg/kg	0.05	0.042	84	64-122	
Trichlorofluoromethane	mg/kg	0.05	0.033	66	59-129	
Vinyl chloride	mg/kg	0.05	0.034	67	42-148	
Xylene (Total)	mg/kg	0.15	0.12	81	62-126	
4-Bromofluorobenzene (S)	%.			99	40-149	
Dibromofluoromethane (S)	%.			101	73-132	
Toluene-d8 (S)	%.			103	66-148	

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Project: Exide ERT Frankfort Site

Pace Project No.: 50300888

Date: 11/05/2021 04:07 PM

QC Batch: 647839 Analysis Method: EPA 8270

QC Batch Method: EPA 3546 Analysis Description: 8270 Solid MSSV Microwave Short Spike

Laboratory: Pace Analytical Services - Indianapolis

Associated Lab Samples: 50300888002

METHOD BLANK: 2985029 Matrix: Solid

Associated Lab Samples: 50300888002

Parameter	Units	Blank Result	Reporting	MDL	Analyzed	Qualifiers
			Limit			
1,2,4,5-Tetrachlorobenzene	mg/kg	<0.079	0.33	0.079	11/01/21 13:59	
2,2'-Oxybis(1-chloropropane)	mg/kg	<0.11	0.33	0.11	11/01/21 13:59	
2,3,4,6-Tetrachlorophenol	mg/kg	<0.12	0.33	0.12	11/01/21 13:59	
2,4,5-Trichlorophenol	mg/kg	<0.12	0.33	0.12	11/01/21 13:59	
2,4,6-Trichlorophenol	mg/kg	<0.10	0.33	0.10	11/01/21 13:59	
2,4-Dichlorophenol	mg/kg	<0.11	0.33	0.11	11/01/21 13:59	
2,4-Dimethylphenol	mg/kg	<0.11	0.33	0.11	11/01/21 13:59	
2,4-Dinitrophenol	mg/kg	<0.18	1.6	0.18	11/01/21 13:59	
2,4-Dinitrotoluene	mg/kg	<0.11	0.33	0.11	11/01/21 13:59	
2,6-Dinitrotoluene	mg/kg	< 0.094	0.33	0.094	11/01/21 13:59	
2-Chloronaphthalene	mg/kg	< 0.093	0.33	0.093	11/01/21 13:59	
2-Chlorophenol	mg/kg	<0.12	0.33	0.12	11/01/21 13:59	
2-Methylnaphthalene	mg/kg	< 0.098	0.33	0.098	11/01/21 13:59	
2-Methylphenol(o-Cresol)	mg/kg	<0.14	0.33	0.14	11/01/21 13:59	
2-Nitroaniline	mg/kg	<0.14	0.33	0.14	11/01/21 13:59	
2-Nitrophenol	mg/kg	<0.13	0.33	0.13	11/01/21 13:59	
3&4-Methylphenol(m&p Cresol)	mg/kg	<0.14	0.66	0.14	11/01/21 13:59	
3,3'-Dichlorobenzidine	mg/kg	<0.11	0.66	0.11	11/01/21 13:59	
3-Nitroaniline	mg/kg	<0.12	0.33	0.12	11/01/21 13:59	
4,6-Dinitro-2-methylphenol	mg/kg	<0.20	0.66	0.20	11/01/21 13:59	
4-Bromophenylphenyl ether	mg/kg	<0.12	0.33	0.12	11/01/21 13:59	
4-Chloro-3-methylphenol	mg/kg	<0.13	0.66	0.13	11/01/21 13:59	
4-Chloroaniline	mg/kg	< 0.087	0.66	0.087	11/01/21 13:59	
4-Chlorophenylphenyl ether	mg/kg	<0.10	0.33	0.10	11/01/21 13:59	
4-Nitroaniline	mg/kg	<0.13	0.33	0.13	11/01/21 13:59	
4-Nitrophenol	mg/kg	<0.25	1.6	0.25	11/01/21 13:59	
Acenaphthene	mg/kg	<0.088	0.33	0.088	11/01/21 13:59	
Acenaphthylene	mg/kg	< 0.099	0.33	0.099	11/01/21 13:59	
Acetophenone	mg/kg	< 0.099	0.33	0.099	11/01/21 13:59	
Anthracene	mg/kg	<0.14	0.33	0.14	11/01/21 13:59	
Atrazine	mg/kg	<0.14	0.33	0.14	11/01/21 13:59	N2
Benzaldehyde	mg/kg	<0.11	0.33	0.11	11/01/21 13:59	N2
Benzo(a)anthracene	mg/kg	<0.098	0.33	0.098	11/01/21 13:59	
Benzo(a)pyrene	mg/kg	<0.11	0.33	0.11	11/01/21 13:59	
Benzo(b)fluoranthene	mg/kg	<0.11	0.33	0.11	11/01/21 13:59	
Benzo(g,h,i)perylene	mg/kg	<0.12	0.33	0.12	11/01/21 13:59	
Benzo(k)fluoranthene	mg/kg	<0.12	0.33	0.12	11/01/21 13:59	
Biphenyl (Diphenyl)	mg/kg	<0.091	0.33	0.091	11/01/21 13:59	N2
bis(2-Chloroethoxy)methane	mg/kg	<0.11	0.33	0.11	11/01/21 13:59	
bis(2-Chloroethyl) ether	mg/kg	<0.13	0.33	0.13	11/01/21 13:59	
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Results presented on this page are in the units indicated by the "Units" column except where an alternate unit is presented to the right of the result.



Project: Exide ERT Frankfort Site

Pace Project No.: 50300888

Date: 11/05/2021 04:07 PM

METHOD BLANK: 2985029 Matrix: Solid

Associated Lab Samples: 50300888002

	Blank	Reporting			
Units	Result	Limit	MDL	Analyzed	Qualifiers
mg/kg	<0.10	0.33	0.10	11/01/21 13:59	
mg/kg	<0.18	0.33	0.18	11/01/21 13:59	
mg/kg	<0.16	0.33	0.16	11/01/21 13:59	N2
mg/kg	<0.13	0.33	0.13	11/01/21 13:59	
mg/kg	<0.11	0.33	0.11	11/01/21 13:59	
mg/kg	<0.12	0.33	0.12	11/01/21 13:59	
mg/kg	<0.12	0.33	0.12	11/01/21 13:59	
mg/kg	<0.12	0.33	0.12	11/01/21 13:59	
mg/kg	<0.10	0.33	0.10	11/01/21 13:59	
mg/kg	<0.11	0.33	0.11	11/01/21 13:59	
mg/kg	<0.11	0.33	0.11	11/01/21 13:59	
mg/kg	<0.13	0.33	0.13	11/01/21 13:59	
mg/kg	<0.11	0.33	0.11	11/01/21 13:59	
mg/kg	< 0.090	0.33	0.090	11/01/21 13:59	
mg/kg	< 0.084	0.33	0.084	11/01/21 13:59	
mg/kg	<0.16	0.33	0.16	11/01/21 13:59	
mg/kg	<0.10	0.33	0.10	11/01/21 13:59	
mg/kg	<0.12	0.33	0.12	11/01/21 13:59	
mg/kg	<0.11	0.33	0.11	11/01/21 13:59	
mg/kg	<0.13	0.33	0.13	11/01/21 13:59	
mg/kg	<0.11	0.33	0.11	11/01/21 13:59	
mg/kg	< 0.095	0.33	0.095	11/01/21 13:59	
mg/kg	<0.11	0.33	0.11	11/01/21 13:59	
mg/kg	< 0.25	1.6	0.25	11/01/21 13:59	
mg/kg	<0.13	0.33	0.13	11/01/21 13:59	
mg/kg	<0.12	0.33	0.12	11/01/21 13:59	
mg/kg	<0.10	0.33	0.10	11/01/21 13:59	
%.	87	20-121		11/01/21 13:59	
%.	77	35-96		11/01/21 13:59	
%.	76	33-111		11/01/21 13:59	
%.	65	32-105		11/01/21 13:59	
%.	86	31-145		11/01/21 13:59	
%.	80	35-114		11/01/21 13:59	
	mg/kg	Units Result mg/kg <0.10	Units Result Limit mg/kg <0.10	Units Result Limit MDL mg/kg <0.10	Units Result Limit MDL Analyzed mg/kg <0.10

LABORATORY CONTROL SAMPLE:	2985030					
		Spike	LCS	LCS	% Rec	
Parameter	Units	Conc.	Result	% Rec	Limits	Qualifiers
2,4-Dinitrotoluene	mg/kg	1.7	1.4	82	48-116	
2-Chlorophenol	mg/kg	1.7	1.3	77	58-100	
2-Methylnaphthalene	mg/kg	1.7	1.3	77	53-99	
4-Chloro-3-methylphenol	mg/kg	1.7	1.4	85	57-112	
4-Nitrophenol	mg/kg	1.7	1.2J	74	36-133	
Acenaphthene	mg/kg	1.7	1.4	82	61-98	
Acenaphthylene	mg/kg	1.7	1.3	79	61-98	

Results presented on this page are in the units indicated by the "Units" column except where an alternate unit is presented to the right of the result.



Project: Exide ERT Frankfort Site

Pace Project No.: 50300888

Date: 11/05/2021 04:07 PM

ABORATORY CONTROL SAMPLE:	2985030	0. "			0/ D	
Parameter	Units	Spike Conc.	LCS Result	LCS % Rec	% Rec Limits	Qualifiers
	-					Quamore
nthracene	mg/kg	1.7	1.4	86	62-100	
enzo(a)anthracene	mg/kg	1.7	1.4	86	64-101	
enzo(a)pyrene	mg/kg	1.7	1.4	82	60-104	
enzo(b)fluoranthene	mg/kg	1.7	1.6	93	65-107	
enzo(g,h,i)perylene	mg/kg	1.7	1.5	90	61-108	
enzo(k)fluoranthene	mg/kg	1.7	1.5	90	61-109	
hrysene	mg/kg	1.7	1.5	88	64-101	
ibenz(a,h)anthracene	mg/kg	1.7	1.5	90	65-106	
uoranthene	mg/kg	1.7	1.5	88	63-111	
uorene	mg/kg	1.7	1.4	84	64-100	
deno(1,2,3-cd)pyrene	mg/kg	1.7	1.5	89	64-106	
Nitroso-di-n-propylamine	mg/kg	1.7	1.3	76	48-101	
aphthalene	mg/kg	1.7	1.3	75	58-93	
entachlorophenol	mg/kg	1.7	1.3J	76	33-117	
nenanthrene	mg/kg	1.7	1.5	88	62-102	
nenol	mg/kg	1.7	1.3	79	56-101	
yrene	mg/kg	1.7	1.3	80	60-105	
4,6-Tribromophenol (S)	%.			87	20-121	
Fluorobiphenyl (S)	%.			79	35-96	
Fluorophenol (S)	%.			70	33-111	
robenzene-d5 (S)	%.			70	32-105	
Terphenyl-d14 (S)	%.			84	31-145	
nenol-d5 (S)	%.			82	35-114	

MATRIX SPIKE & MATRIX S	PIKE DUPLIC	ATE: 2985	031		2985032							
			MS	MSD								
	50	0300743001	Spike	Spike	MS	MSD	MS	MSD	% Rec		Max	
Parameter	Units	Result	Conc.	Conc.	Result	Result	% Rec	% Rec	Limits	RPD	RPD	Qua
2,4-Dinitrotoluene	mg/kg	ND	1.8	1.9	1.5	1.5	81	80	13-119	0	20	
2-Chlorophenol	mg/kg	ND	1.8	1.9	1.5	1.5	82	77	16-116	4	20	
2-Methylnaphthalene	mg/kg	ND	1.8	1.9	1.5	1.5	78	81	19-120	5	20	
4-Chloro-3-methylphenol	mg/kg	ND	1.8	1.9	1.5	1.5	83	81	22-124	1	20	
4-Nitrophenol	mg/kg	ND	1.8	1.9	1.5J	1.4J	80	76	10-139		20	
Acenaphthene	mg/kg	ND	1.8	1.9	1.5	1.5	83	81	25-114	1	20	
Acenaphthylene	mg/kg	ND	1.8	1.9	1.5	1.5	78	77	21-116	0	20	
Anthracene	mg/kg	ND	1.8	1.9	1.6	1.6	84	84	23-116	2	20	
Benzo(a)anthracene	mg/kg	ND	1.8	1.9	1.6	1.6	84	85	14-128	2	20	
Benzo(a)pyrene	mg/kg	ND	1.8	1.9	1.5	1.5	79	79	12-127	1	20	
Benzo(b)fluoranthene	mg/kg	ND	1.8	1.9	1.7	1.7	89	93	10-142	4	20	
Benzo(g,h,i)perylene	mg/kg	ND	1.8	1.9	1.6	1.6	84	85	16-120	2	20	
Benzo(k)fluoranthene	mg/kg	ND	1.8	1.9	1.6	1.6	86	85	15-131	0	20	
Chrysene	mg/kg	ND	1.8	1.9	1.5	1.6	82	84	11-132	3	20	
Dibenz(a,h)anthracene	mg/kg	ND	1.8	1.9	1.6	1.6	84	85	21-117	2	20	
Fluoranthene	mg/kg	ND	1.8	1.9	1.6	1.6	87	87	10-143	1	20	
Fluorene	mg/kg	ND	1.8	1.9	1.6	1.6	83	83	18-122	0	20	

Results presented on this page are in the units indicated by the "Units" column except where an alternate unit is presented to the right of the result.



Project: Exide ERT Frankfort Site

Pace Project No.: 50300888

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MATRIX SPIKE & MATRIX SF	PIKE DUPLIC	ATE: 2985	031 MS	MSD	2985032							
	50	0300743001	Spike	Spike	MS	MSD	MS	MSD	% Rec		Max	
Parameter	Units	Result	Conc.	Conc.	Result	Result	% Rec	% Rec	Limits	RPD	RPD	Qual
Indeno(1,2,3-cd)pyrene	mg/kg	ND	1.8	1.9	1.6	1.6	84	85	19-120	2	20	
N-Nitroso-di-n-propylamine	mg/kg	ND	1.8	1.9	1.7	1.4	91	75	24-109	18	20	
Naphthalene	mg/kg	ND	1.8	1.9	1.5	1.4	78	77	22-112	1	20	
Pentachlorophenol	mg/kg	ND	1.8	1.9	1.3J	1.4J	71	76	10-123		20	
Phenanthrene	mg/kg	ND	1.8	1.9	1.6	1.7	85	88	10-136	4	20	
Phenol	mg/kg	ND	1.8	1.9	1.5	1.5	80	78	14-122	2	20	
Pyrene	mg/kg	ND	1.8	1.9	1.6	1.7	88	89	10-144	2	20	
2,4,6-Tribromophenol (S)	%.						84	90	20-121			
2-Fluorobiphenyl (S)	%.						76	79	35-96			
2-Fluorophenol (S)	%.						80	76	33-111			
Nitrobenzene-d5 (S)	%.						74	70	32-105			
p-Terphenyl-d14 (S)	%.						84	84	31-145			
Phenol-d5 (S)	%.						83	80	35-114			



Project: Exide ERT Frankfort Site

Pace Project No.: 50300888

Date: 11/05/2021 04:07 PM

QC Batch: 648252 Analysis Method: EPA 8270

QC Batch Method: EPA 3510 Analysis Description: 8270 TCLP MSSV

Laboratory: Pace Analytical Services - Indianapolis

Associated Lab Samples: 50300888001, 50300888002

METHOD BLANK: 2986665 Matrix: Water

Associated Lab Samples: 50300888001, 50300888002

Associated Lab Campies. 30300000	1001, 30300888002					
		Blank	Reporting			
Parameter	Units	Result	Limit	MDL	Analyzed	Qualifiers
1,4-Dichlorobenzene	mg/L	<0.0050	0.010	0.0050	11/03/21 18:37	
2,4,5-Trichlorophenol	mg/L	< 0.0050	0.050	0.0050	11/03/21 18:37	
2,4,6-Trichlorophenol	mg/L	< 0.0050	0.010	0.0050	11/03/21 18:37	
2,4-Dinitrotoluene	mg/L	< 0.0050	0.010	0.0050	11/03/21 18:37	
2-Methylphenol(o-Cresol)	mg/L	< 0.0050	0.010	0.0050	11/03/21 18:37	
3&4-Methylphenol(m&p Cresol)	mg/L	< 0.010	0.020	0.010	11/03/21 18:37	
Hexachloro-1,3-butadiene	mg/L	< 0.0050	0.010	0.0050	11/03/21 18:37	
Hexachlorobenzene	mg/L	< 0.0050	0.010	0.0050	11/03/21 18:37	
Hexachloroethane	mg/L	< 0.0050	0.010	0.0050	11/03/21 18:37	
Nitrobenzene	mg/L	< 0.0050	0.010	0.0050	11/03/21 18:37	
Pentachlorophenol	mg/L	< 0.025	0.050	0.025	11/03/21 18:37	
Pyridine	mg/L	< 0.010	0.010	0.010	11/03/21 18:37	
2,4,6-Tribromophenol (S)	%.	79	47-127		11/03/21 18:37	
2-Fluorobiphenyl (S)	%.	55	35-102		11/03/21 18:37	
2-Fluorophenol (S)	%.	37	21-74		11/03/21 18:37	
Nitrobenzene-d5 (S)	%.	62	40-115		11/03/21 18:37	
p-Terphenyl-d14 (S)	%.	86	42-156		11/03/21 18:37	
Phenol-d5 (S)	%.	25	15-48		11/03/21 18:37	
Thomas (o)	70.	25	13-40		11/00/21 10.07	

		Spike	LCS	LCS	% Rec	
Parameter	Units	Conc.	Result	% Rec	Limits	Qualifiers
Dichlorobenzene	mg/L	0.05	0.028	57	30-85	
5-Trichlorophenol	mg/L	0.05	0.041J	82	52-117	
6-Trichlorophenol	mg/L	0.05	0.039	78	52-114	
Dinitrotoluene	mg/L	0.05	0.040	80	58-107	
lethylphenol(o-Cresol)	mg/L	0.05	0.032	65	40-95	
I-Methylphenol(m&p Cresol)	mg/L	0.1	0.061	61	37-89	
achloro-1,3-butadiene	mg/L	0.05	0.027	54	22-78	
achlorobenzene	mg/L	0.05	0.032	64	46-79	
achloroethane	mg/L	0.05	0.027	54	17-85	
benzene	mg/L	0.05	0.038	77	50-110	
tachlorophenol	mg/L	0.05	0.041J	83	32-126	
dine	mg/L	0.05	0.017	33	18-69	
6-Tribromophenol (S)	%.			80	47-127	
luorobiphenyl (S)	%.			66	35-102	
luorophenol (S)	%.			43	21-74	
obenzene-d5 (S)	%.			72	40-115	
erphenyl-d14 (S)	%.			87	42-156	

Results presented on this page are in the units indicated by the "Units" column except where an alternate unit is presented to the right of the result.



Project: Exide ERT Frankfort Site

Pace Project No.: 50300888

Date: 11/05/2021 04:07 PM

LABORATORY CONTROL SAMPLE: 2986666

Spike LCS LCS % Rec

Parameter Units Conc. Result % Rec Limits Qualifiers

Phenol-d5 (S) %. 30 15-48

MATRIX SPIKE SAMPLE: 2986667 50300799004 MS MS % Rec Spike Parameter Units Result Conc. Result % Rec Limits Qualifiers ND 1,4-Dichlorobenzene mg/L 0.5 0.31 63 24-81 ND 0.5 0.47J 34-129 2,4,5-Trichlorophenol mg/L 94 ND 2,4,6-Trichlorophenol mg/L 0.5 0.43 85 33-123 ND 2,4-Dinitrotoluene mg/L 0.5 0.43 87 35-116 ND 2-Methylphenol(o-Cresol) mg/L 0.5 0.32 64 24-102 ND 3&4-Methylphenol(m&p Cresol) mg/L 1 0.63 63 18-99 Hexachloro-1,3-butadiene mg/L ND 0.5 0.31 61 15-79 Hexachlorobenzene mg/L ND 0.5 0.34 68 29-86 Hexachloroethane ND 0.5 0.31 62 14-79 mg/L ND 0.5 Nitrobenzene mg/L 0.40 79 27-117 mg/L ND 0.40J Pentachlorophenol 0.5 81 15-151 ND Pyridine 0.5 0.26 52 12-75 mg/L 2,4,6-Tribromophenol (S) 87 47-127 %. 2-Fluorobiphenyl (S) %. 64 35-102 2-Fluorophenol (S) %. 46 21-74 Nitrobenzene-d5 (S) %. 75 40-115 p-Terphenyl-d14 (S) %. 85 42-156 Phenol-d5 (S) %. 33 15-48

Results presented on this page are in the units indicated by the "Units" column except where an alternate unit is presented to the right of the result.



Project: Exide ERT Frankfort Site

Pace Project No.: 50300888

Date: 11/05/2021 04:07 PM

QC Batch: 647400 Analysis Method: EPA 8270 by SIM

QC Batch Method: EPA 3510 Analysis Description: 8270 Water PAH Low Volume

Laboratory: Pace Analytical Services - Indianapolis

Associated Lab Samples: 50300888001

METHOD BLANK: 2982637 Matrix: Water

Associated Lab Samples: 50300888001

Parameter	Units	Blank Result	Reporting Limit	MDL	Analyzed	Qualifiers
2-Methylnaphthalene	ug/L	<0.015	1.0	0.015	11/01/21 17:15	
Acenaphthene	ug/L	<0.015	1.0	0.015	11/01/21 17:15	
Acenaphthylene	ug/L	< 0.013	1.0	0.013	11/01/21 17:15	
Anthracene	ug/L	< 0.012	0.10	0.012	11/01/21 17:15	
Benzo(a)anthracene	ug/L	< 0.027	0.10	0.027	11/01/21 17:15	
Benzo(a)pyrene	ug/L	< 0.026	0.10	0.026	11/01/21 17:15	
Benzo(b)fluoranthene	ug/L	< 0.031	0.10	0.031	11/01/21 17:15	
Benzo(g,h,i)perylene	ug/L	< 0.024	0.10	0.024	11/01/21 17:15	
Benzo(k)fluoranthene	ug/L	< 0.020	0.10	0.020	11/01/21 17:15	
Chrysene	ug/L	< 0.020	0.50	0.020	11/01/21 17:15	
Dibenz(a,h)anthracene	ug/L	0.090J	0.10	0.071	11/01/21 17:15	
Fluoranthene	ug/L	< 0.015	1.0	0.015	11/01/21 17:15	
Fluorene	ug/L	< 0.036	1.0	0.036	11/01/21 17:15	
Indeno(1,2,3-cd)pyrene	ug/L	< 0.073	0.10	0.073	11/01/21 17:15	
Naphthalene	ug/L	0.18J	1.0	0.014	11/01/21 17:15	
Phenanthrene	ug/L	< 0.021	1.0	0.021	11/01/21 17:15	
Pyrene	ug/L	< 0.020	1.0	0.020	11/01/21 17:15	
2-Fluorobiphenyl (S)	%.	56	31-98		11/01/21 17:15	
p-Terphenyl-d14 (S)	%.	92	33-115		11/01/21 17:15	

LABORATORY CONTROL SAMPLE:	2982638					
		Spike	LCS	LCS	% Rec	
Parameter	Units	Conc.	Result	% Rec	Limits	Qualifiers
2-Methylnaphthalene	ug/L	10	3.3	33	46-95	L2
Acenaphthene	ug/L	10	3.7	37	49-103	L2
cenaphthylene	ug/L	10	4.6	46	53-102	L2
Inthracene	ug/L	10	5.1	51	47-104	
enzo(a)anthracene	ug/L	10	6.6	66	44-107	
enzo(a)pyrene	ug/L	10	6.6	66	33-101	
enzo(b)fluoranthene	ug/L	10	6.3	63	34-105	
enzo(g,h,i)perylene	ug/L	10	5.2	52	21-95	
enzo(k)fluoranthene	ug/L	10	5.1	51	29-113	
hrysene	ug/L	10	5.3	53	48-96	
ibenz(a,h)anthracene	ug/L	10	5.3	53	21-102	
uoranthene	ug/L	10	5.6	56	50-116	
uorene	ug/L	10	4.6	46	51-103	L2
deno(1,2,3-cd)pyrene	ug/L	10	5.3	53	22-102	
aphthalene	ug/L	10	3.4	34	44-97	L2
henanthrene	ug/L	10	4.9	49	53-101	L2

Results presented on this page are in the units indicated by the "Units" column except where an alternate unit is presented to the right of the result.



Project: Exide ERT Frankfort Site

Pace Project No.: 50300888

Date: 11/05/2021 04:07 PM

LABORATORY CONTROL SAMPLE:	2982638					
		Spike	LCS	LCS	% Rec	
Parameter	Units	Conc.	Result	% Rec	Limits	Qualifiers
Pyrene	ug/L	10	6.0	60	58-106	
2-Fluorobiphenyl (S)	%.			37	31-98	
p-Terphenyl-d14 (S)	%.			64	33-115	



Project: Exide ERT Frankfort Site

Pace Project No.: 50300888

Date: 11/05/2021 04:07 PM

QC Batch: 648745 Analysis Method: EPA 8270 by SIM

QC Batch Method: EPA 3510 Analysis Description: 8270 Water PAH Low Volume

Laboratory: Pace Analytical Services - Indianapolis

Associated Lab Samples: 50300888001

METHOD BLANK: 2989319 Matrix: Water

Associated Lab Samples: 50300888001

Parameter	Units	Blank Result	Reporting Limit	MDL	Analyzed	Qualifiers
2-Methylnaphthalene	ug/L	<0.015	1.0	0.015	11/05/21 13:56	
Acenaphthene	ug/L	<0.015	1.0	0.015	11/05/21 13:56	
Acenaphthylene	ug/L	< 0.013	1.0	0.013	11/05/21 13:56	
Anthracene	ug/L	< 0.012	0.10	0.012	11/05/21 13:56	
Benzo(a)anthracene	ug/L	< 0.027	0.10	0.027	11/05/21 13:56	
Benzo(a)pyrene	ug/L	< 0.026	0.10	0.026	11/05/21 13:56	
Benzo(b)fluoranthene	ug/L	< 0.031	0.10	0.031	11/05/21 13:56	
Benzo(g,h,i)perylene	ug/L	< 0.024	0.10	0.024	11/05/21 13:56	
Benzo(k)fluoranthene	ug/L	< 0.020	0.10	0.020	11/05/21 13:56	
Chrysene	ug/L	< 0.020	0.50	0.020	11/05/21 13:56	
Dibenz(a,h)anthracene	ug/L	< 0.071	0.10	0.071	11/05/21 13:56	
Fluoranthene	ug/L	< 0.015	1.0	0.015	11/05/21 13:56	
Fluorene	ug/L	< 0.036	1.0	0.036	11/05/21 13:56	
Indeno(1,2,3-cd)pyrene	ug/L	< 0.073	0.10	0.073	11/05/21 13:56	
Naphthalene	ug/L	< 0.014	1.0	0.014	11/05/21 13:56	
Phenanthrene	ug/L	< 0.021	1.0	0.021	11/05/21 13:56	
Pyrene	ug/L	< 0.020	1.0	0.020	11/05/21 13:56	
2-Fluorobiphenyl (S)	%.	64	31-98		11/05/21 13:56	
p-Terphenyl-d14 (S)	%.	89	33-115		11/05/21 13:56	

LABORATORY CONTROL SAMPLE:	2989320					
		Spike	LCS	LCS	% Rec	
Parameter	Units	Conc.	Result	% Rec	Limits	Qualifiers
2-Methylnaphthalene	ug/L	10	6.7	67	46-95	
Acenaphthene	ug/L	10	7.0	70	49-103	
Acenaphthylene	ug/L	10	8.3	83	53-102	
Anthracene	ug/L	10	8.2	82	47-104	
Benzo(a)anthracene	ug/L	10	9.7	97	44-107	
Benzo(a)pyrene	ug/L	10	8.8	88	33-101	
Benzo(b)fluoranthene	ug/L	10	6.9	69	34-105	
Benzo(g,h,i)perylene	ug/L	10	6.2	62	21-95	
Benzo(k)fluoranthene	ug/L	10	8.0	80	29-113	
Chrysene	ug/L	10	7.7	77	48-96	
Dibenz(a,h)anthracene	ug/L	10	6.1	61	21-102	
Fluoranthene	ug/L	10	8.5	85	50-116	
Fluorene	ug/L	10	8.2	82	51-103	
Indeno(1,2,3-cd)pyrene	ug/L	10	6.2	62	22-102	
Naphthalene	ug/L	10	6.9	69	44-97	
Phenanthrene	ug/L	10	8.1	81	53-101	

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Project: Exide ERT Frankfort Site

Pace Project No.: 50300888

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LABORATORY CONTROL SAMPLE:	2989320					
		Spike	LCS	LCS	% Rec	
Parameter	Units	Conc.	Result	% Rec	Limits	Qualifiers
Pyrene	ug/L	10	9.0	90	58-106	
2-Fluorobiphenyl (S)	%.			70	31-98	
p-Terphenyl-d14 (S)	%.			81	33-115	



Project: Exide ERT Frankfort Site

Pace Project No.: 50300888

Date: 11/05/2021 04:07 PM

QC Batch: 647394 Analysis Method: EPA 8270

QC Batch Method: EPA 3510 Analysis Description: 8270 Water Scan LV

Laboratory: Pace Analytical Services - Indianapolis

Associated Lab Samples: 50300888001

METHOD BLANK: 2982615 Matrix: Water

Associated Lab Samples: 50300888001

_		Blank	Reporting			
Parameter	Units	Result	Limit	MDL	Analyzed	Qualifiers
2,2'-Oxybis(1-chloropropane)	ug/L	<4.6	10.0	4.6	11/01/21 18:11	
2,3,4,6-Tetrachlorophenol	ug/L	<4.9	10.0	4.9	11/01/21 18:11	
2,4,5-Trichlorophenol	ug/L	<2.9	10.0	2.9	11/01/21 18:11	
2,4,6-Trichlorophenol	ug/L	<4.5	10.0	4.5	11/01/21 18:11	
2,4-Dichlorophenol	ug/L	<4.0	10.0	4.0	11/01/21 18:11	
2,4-Dimethylphenol	ug/L	<8.1	10.0	8.1	11/01/21 18:11	
2,4-Dinitrophenol	ug/L	<6.6	50.0	6.6	11/01/21 18:11	
2,4-Dinitrotoluene	ug/L	<6.2	10.0	6.2	11/01/21 18:11	
2,6-Dinitrotoluene	ug/L	<4.6	10.0	4.6	11/01/21 18:11	
2-Chloronaphthalene	ug/L	<5.8	10.0	5.8	11/01/21 18:11	
2-Chlorophenol	ug/L	<3.6	10.0	3.6	11/01/21 18:11	
2-Methylphenol(o-Cresol)	ug/L	<4.3	10.0	4.3	11/01/21 18:11	
2-Nitroaniline	ug/L	<4.2	10.0	4.2	11/01/21 18:11	
2-Nitrophenol	ug/L	<3.5	10.0	3.5	11/01/21 18:11	
3&4-Methylphenol(m&p Cresol)	ug/L	< 5.4	10.0	5.4	11/01/21 18:11	
3,3'-Dichlorobenzidine	ug/L	<4.0	20.0	4.0	11/01/21 18:11	
3-Nitroaniline	ug/L	<4.8	10.0	4.8	11/01/21 18:11	
4,6-Dinitro-2-methylphenol	ug/L	< 5.0	20.0	5.0	11/01/21 18:11	
4-Bromophenylphenyl ether	ug/L	< 5.6	10.0	5.6	11/01/21 18:11	
4-Chloro-3-methylphenol	ug/L	< 5.6	10.0	5.6	11/01/21 18:11	
4-Chloroaniline	ug/L	<3.2	10.0	3.2	11/01/21 18:11	
4-Chlorophenylphenyl ether	ug/L	<5.1	10.0	5.1	11/01/21 18:11	
4-Nitroaniline	ug/L	<4.6	10.0	4.6	11/01/21 18:11	
4-Nitrophenol	ug/L	< 5.6	50.0	5.6	11/01/21 18:11	
Acetophenone	ug/L	<2.8	10.0	2.8	11/01/21 18:11	
Atrazine	ug/L	<2.8	10.0	2.8	11/01/21 18:11	
Benzaldehyde	ug/L	<4.7	50.0	4.7	11/01/21 18:11	
Biphenyl (Diphenyl)	ug/L	< 5.9	10.0	5.9	11/01/21 18:11	
bis(2-Chloroethoxy)methane	ug/L	<2.5	10.0	2.5	11/01/21 18:11	
bis(2-Chloroethyl) ether	ug/L	<2.9	10.0	2.9	11/01/21 18:11	
bis(2-Ethylhexyl)phthalate	ug/L	<3.1	10.0	3.1	11/01/21 18:11	
Butylbenzylphthalate	ug/L	<3.5	10.0	3.5	11/01/21 18:11	
Caprolactam	ug/L	<4.3	10.0	4.3	11/01/21 18:11	
Carbazole	ug/L	<3.7	10.0	3.7	11/01/21 18:11	
Di-n-butylphthalate	ug/L	<3.6	10.0	3.6	11/01/21 18:11	
Di-n-octylphthalate	ug/L	<4.5	10.0	4.5	11/01/21 18:11	
Dibenzofuran	ug/L	<7.0	10.0	7.0	11/01/21 18:11	
Diethylphthalate	ug/L	<2.7	10.0	2.7	11/01/21 18:11	
Dimethylphthalate	ug/L	<3.7	10.0	3.7	11/01/21 18:11	
Hexachloro-1,3-butadiene	ug/L	<4.1	10.0	4.1	11/01/21 18:11	

Results presented on this page are in the units indicated by the "Units" column except where an alternate unit is presented to the right of the result.



Project: Exide ERT Frankfort Site

Pace Project No.: 50300888

Date: 11/05/2021 04:07 PM

METHOD BLANK: 2982615 Matrix: Water

Associated Lab Samples: 50300888001

		Blank	Reporting			
Parameter	Units	Result	Limit	MDL	Analyzed	Qualifiers
Hexachlorobenzene	ug/L	<3.0	10.0	3.0	11/01/21 18:11	
Hexachlorocyclopentadiene	ug/L	<3.0	10.0	3.0	11/01/21 18:11	
Hexachloroethane	ug/L	<2.5	10.0	2.5	11/01/21 18:11	
Isophorone	ug/L	<4.2	10.0	4.2	11/01/21 18:11	
N-Nitroso-di-n-propylamine	ug/L	<2.9	50.0	2.9	11/01/21 18:11	
N-Nitrosodiphenylamine	ug/L	<2.9	10.0	2.9	11/01/21 18:11	
Nitrobenzene	ug/L	<3.0	10.0	3.0	11/01/21 18:11	
Pentachlorophenol	ug/L	<4.0	50.0	4.0	11/01/21 18:11	
Phenol	ug/L	<4.1	10.0	4.1	11/01/21 18:11	
2,4,6-Tribromophenol (S)	%.	112	34-126		11/01/21 18:11	
2-Fluorophenol (S)	%.	60	10-72		11/01/21 18:11	
Nitrobenzene-d5 (S)	%.	89	39-115		11/01/21 18:11	
Phenol-d5 (S)	%.	46	10-55		11/01/21 18:11	

LABORATORY CONTROL SAMPLE	E: 2982616					
		Spike	LCS	LCS	% Rec	
Parameter	Units	Conc.	Result	% Rec	Limits	Qualifiers
2,4-Dimethylphenol	ug/L	100	54.9	55	26-142	
2,4-Dinitrotoluene	ug/L	100	63.0	63	55-141	
2-Chlorophenol	ug/L	100	47.4	47	26-110	
4-Chloro-3-methylphenol	ug/L	100	63.4	63	34-140	
4-Nitrophenol	ug/L	100	55.6	56	10-94	
bis(2-Ethylhexyl)phthalate	ug/L	100	75.5	76	48-160	
Dibenzofuran	ug/L	100	51.0	51	38-125	
N-Nitroso-di-n-propylamine	ug/L	100	50.6	51	48-126	
Pentachlorophenol	ug/L	100	69.9	70	43-144	
Phenol	ug/L	100	41.1	41	10-73	
2,4,6-Tribromophenol (S)	%.			68	34-126	
2-Fluorophenol (S)	%.			48	10-72	
Nitrobenzene-d5 (S)	%.			54	39-115	
Phenol-d5 (S)	%.			42	10-55	

Results presented on this page are in the units indicated by the "Units" column except where an alternate unit is presented to the right of the result.



Project: Exide ERT Frankfort Site

Pace Project No.: 50300888

QC Batch: 646639 Analysis Method: SM 2540G

QC Batch Method: SM 2540G Analysis Description: Dry Weight/Percent Moisture

Laboratory: Pace Analytical Services - Indianapolis

Associated Lab Samples: 50300888002

SAMPLE DUPLICATE: 2979704

 Parameter
 Units
 50300947003 Result
 Dup Result
 Max RPD
 RPD
 Qualifiers

 Percent Moisture
 %
 54.6
 54.9
 1
 5 N2

SAMPLE DUPLICATE: 2979705

Date: 11/05/2021 04:07 PM

		50300913001	Dup		Max	
Parameter	Units	Result	Result	RPD	RPD	Qualifiers
Percent Moisture	%		11.9	2		5 N2



Project: Exide ERT Frankfort Site

Pace Project No.: 50300888

QC Batch: 647806 Analysis Method: SM 4500-H+B
QC Batch Method: SM 4500-H+B Analysis Description: 4500H+B pH

Laboratory: Pace Analytical Services - Indianapolis

Associated Lab Samples: 50300888001

SAMPLE DUPLICATE: 2984774

50301006004 Dup Max Units RPD RPD Qualifiers Parameter Result Result 7.2 7.2 2 H3 pH at 25 Degrees C Std. Units 0

SAMPLE DUPLICATE: 2984775

Date: 11/05/2021 04:07 PM

50301172001 Dup Max Parameter Units Result Result **RPD RPD** Qualifiers 8.4 pH at 25 Degrees C 0 2 H3 Std. Units 8.5

Results presented on this page are in the units indicated by the "Units" column except where an alternate unit is presented to the right of the result.



Project: Exide ERT Frankfort Site

Pace Project No.: 50300888

QC Batch: 647576 Analysis Method: EPA 9045
QC Batch Method: EPA 9045 Analysis Description: 9045 pH

Laboratory: Pace Analytical Services - Indianapolis

Associated Lab Samples: 50300888002

SAMPLE DUPLICATE: 2983806

50301213001 Dup Max Units RPD RPD Qualifiers Parameter Result Result 10.2 2 H3,PO pH at 25 Degrees C Std. Units 10.2 0

SAMPLE DUPLICATE: 2983807

Date: 11/05/2021 04:07 PM

 Parameter
 Units
 50301215001 Result
 Dup Result
 Max RPD
 Max RPD
 Qualifiers

 pH at 25 Degrees C
 Std. Units
 6.9
 6.8
 1
 2 H3



Project: Exide ERT Frankfort Site

Pace Project No.: 50300888

QC Batch: 469753 Analysis Method: EPA 9014

QC Batch Method: SW-846 7.3.3.2 Analysis Description: 733C Reactive Cyanide

Laboratory: Pace Analytical Services - Greensburg

Associated Lab Samples: 50300888001, 50300888002

METHOD BLANK: 2268143 Matrix: Solid

Associated Lab Samples: 50300888001, 50300888002

Blank Reporting
Parameter Units Result Limit MDL Analyzed Qualifiers

Cyanide, Reactive mg/kg <0.40 1.0 0.40 11/02/21 12:13

LABORATORY CONTROL SAMPLE: 2268144

Spike LCS LCS % Rec Parameter Conc. Result % Rec Limits Qualifiers Units Cyanide, Reactive mg/kg < 0.40 4 0-8

SAMPLE DUPLICATE: 2268145

Date: 11/05/2021 04:07 PM

Parameter Units Result Result RPD APD Qualifiers

Cyanide, Reactive mg/kg ND <0.70 20

Qualifiers



QUALITY CONTROL DATA

Project: Exide ERT Frankfort Site

Pace Project No.: 50300888

QC Batch: 470138
QC Batch Method: SW-846 7.3.4.2

Analysis Method: SM 4500-S2-F-2011
Analysis Description: 734S Reactive Sulfide

Laboratory:

Pace Analytical Services - Greensburg

Analyzed

Associated Lab Samples: 50300888001, 50300888002

METHOD BLANK: 2269458 Matrix: Solid

Associated Lab Samples: 50300888001, 50300888002

Blank Reporting
Parameter Units Result Limit MDL

Sulfide, Reactive mg/kg <10.0 10.0 10.0 10/29/21 15:02

LABORATORY CONTROL SAMPLE: 2269459

Spike LCS LCS % Rec Parameter Conc. Result % Rec Limits Qualifiers Units Sulfide, Reactive mg/kg 200 <10.0 4 0-52

SAMPLE DUPLICATE: 2269460

Date: 11/05/2021 04:07 PM

Parameter Units Esult Dup Max Result RPD Qualifiers

Sulfide, Reactive mg/kg <10.0 <10.0 20



QUALIFIERS

Project: Exide ERT Frankfort Site

Pace Project No.: 50300888

DEFINITIONS

DF - Dilution Factor, if reported, represents the factor applied to the reported data due to dilution of the sample aliquot.

ND - Not Detected at or above adjusted reporting limit.

TNTC - Too Numerous To Count

J - Estimated concentration above the adjusted method detection limit and below the adjusted reporting limit.

MDL - Adjusted Method Detection Limit.

PQL - Practical Quantitation Limit.

RL - Reporting Limit - The lowest concentration value that meets project requirements for quantitative data with known precision and bias for a specific analyte in a specific matrix.

S - Surrogate

1,2-Diphenylhydrazine decomposes to and cannot be separated from Azobenzene using Method 8270. The result for each analyte is a combined concentration.

Consistent with EPA guidelines, unrounded data are displayed and have been used to calculate % recovery and RPD values.

LCS(D) - Laboratory Control Sample (Duplicate)

MS(D) - Matrix Spike (Duplicate)

DUP - Sample Duplicate

RPD - Relative Percent Difference

NC - Not Calculable.

SG - Silica Gel - Clean-Up

U - Indicates the compound was analyzed for, but not detected.

N-Nitrosodiphenylamine decomposes and cannot be separated from Diphenylamine using Method 8270. The result reported for each analyte is a combined concentration.

Reported results are not rounded until the final step prior to reporting. Therefore, calculated parameters that are typically reported as "Total" may vary slightly from the sum of the reported component parameters.

Pace Analytical is TNI accredited. Contact your Pace PM for the current list of accredited analytes.

TNI - The NELAC Institute.

ANALYTE QUALIFIERS

Date: 11/05/2021 04:07 PM

В	Analyte was detected in the associated method blank.
H2	Extraction or preparation conducted outside EPA method holding time.
H3	Sample was received or analysis requested beyond the recognized method holding time.
H7	Re-extraction or re-analysis could not be performed within method holding time.
L2	Analyte recovery in the laboratory control sample (LCS) was below QC limits. Results for this analyte in associated samples may be biased low.
M0	Matrix spike recovery and/or matrix spike duplicate recovery was outside laboratory control limits.
N2	The lab does not hold NELAC/TNI accreditation for this parameter but other accreditations/certifications may apply. A complete list of accreditations/certifications is available upon request.
PO	The reported result is outside the range of the pH buffer solutions used to check the calibration of the pH meter.



QUALITY CONTROL DATA CROSS REFERENCE TABLE

Project: Exide ERT Frankfort Site

Pace Project No.: 50300888

Date: 11/05/2021 04:07 PM

Lab ID	Sample ID	QC Batch Method	QC Batch	Analytical Method	Analytical Batch
50300888002	Soil-1021	EPA 3050	646594	EPA 6010	647161
50300888001 50300888002	Water-1021 Soil-1021	EPA 3010 EPA 3010	648249 648249	EPA 6010 EPA 6010	648527 648527
50300888001	Water-1021	EPA 3010	646674	EPA 6010	647125
50300888001 50300888002	Water-1021 Soil-1021	EPA 7470 EPA 7470	648297 648297	EPA 7470 EPA 7470	648541 648541
50300888001	Water-1021	EPA 7470	648095	EPA 7470	648661
50300888002	Soil-1021	EPA 7471	647537	EPA 7471	647854
50300888002	Soil-1021	EPA 3546	647839	EPA 8270	647946
50300888001 50300888002	Water-1021 Soil-1021	EPA 3510 EPA 3510	648252 648252	EPA 8270 EPA 8270	648457 648457
50300888001	Water-1021	EPA 3510	647400	EPA 8270 by SIM	648006
50300888001	Water-1021	EPA 3510	648745	EPA 8270 by SIM	648909
50300888001	Water-1021	EPA 3510	647394	EPA 8270	648005
50300888001 50300888002	Water-1021 Soil-1021	EPA 5030/8260 EPA 5030/8260	648494 648494		
50300888001	Water-1021	EPA 8260	647925		
50300888002	Soil-1021	EPA 8260	648672		
50300888002	Soil-1021	SM 2540G	646639		
50300888001	Water-1021	EPA 1020B	646840		
50300888002	Soil-1021	1030	647110		
50300888001	Water-1021	SM 4500-H+B	647806		
0300888002	Soil-1021	EPA 9045	647576		
50300888001 50300888002	Water-1021 Soil-1021	SW-846 7.3.3.2 SW-846 7.3.3.2	469753 469753	EPA 9014 EPA 9014	470778 470778
50300888001 50300888002	Water-1021 Soil-1021	SW-846 7.3.4.2 SW-846 7.3.4.2	470138 470138	SM 4500-S2-F-2011 SM 4500-S2-F-2011	470445 470445

1055 Andrew Drive, Suite A West Chester, PA 19380-4293 tel 610.840.9100 fax 610.840.9199

ADVANCED GEOSERVICES CORP. **CHAIN OF CUSTODY**



Project Name: EkideTr.	st-Fr	nkfor	+	MI C	INTER S	1,144	Proj	ect N	lo.:_	20	26-	41	23	_	_ Sh	nipmo	ent N	No.:	1		
AGC Contact Person: Any	Graham	of a You	ala din sh	5=0(1)	Nav		Ship	men	t Tra	cking	g No	.: L	ub	De 1	ive	ry	OF A	ed Name	joy9		
Laboratory Name/Location: 1	Pace-I	n digo	polis	, most	DE PRO			A		2	to the	_	ANAI								
Sampler's Name(s) (Print):	on's Pet	kr/B	vnie	Bee	51e	_	/	/ /	/	RURA	Such	Jon	your o		//	//	/	/ / AM	9 /		
Sanata IDII aasiia	Dete	Times	Sample Type	Sample	Field Filtered	Total Number	18	1/3	3/3	Tapulal	3/3	+ 1/4	1100					PRESERVATIVE			
Sample ID/Location	Date	Times	C or G	Matrix	Y or N	of Container	2	5/01	12	4	Ha	12	1					PRE		Remarks	
Water-1021	(0/24)21	0820	C	GV	N		D	X	X	X	×	X	332	14112	102	MIE II	(in a r	134	(105)	001	
Soil-1021	10/22/21	0850	C	an	N		X	X	X	X	X	X	1 2-3	. 31591	1.110-6	10.50	1190	1,7,8	10/19	005	
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the service of the se	ed AAC or s	gestack f	apark a	DOLA 19	(100)				125,000	- na	200	DISE S	1671 34	1 100	d Salta	0.8-20	FE 100	75mm7	745		
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Relinquished by:		Date/Time:			-		Received	by:								Г	Date/Time	e:			П
Relinquished by:		Date/Time:	×				Received	by:			Albino Oul):	of V		100	8 (6) 1 17810	g d c	Date/Time	e told ske	0.07		

Preservative: 1-ice, 2-H₂SO₄, 3-HCl, 4-HNO₃, 5-NaOH, 6-ZnOAC, 7-Act hand F-DI water Sample Matrix: SW - Surface Water, GW - Groundwater, Sed - Sediment, S - Soil, Sld - Sludge, A - Air

F-IN-Q-290-rev.21, 02Feb2021

Pace Analytical*

SAMPLE CONDITION UPON RECEIPT FORM

1. Courier: ☐ FED EX ☐ UPS ☐ CLIENT ☐ PAGE		J010 L	5. Packing Material	☐ Bubble Wrap	Bubbl	e Bags	
2. Custody Seal on Cooler/Box Present: Yes	₽ No			□None	☐ Other		
(If yes)Seals Intact:	if no seals	were pres	ent)				
3. Thermometer: 123456 ABC (DEF			6. Ice Type: 🗗 🛚	et 🗆 Blue 🗆 None	9		
4. Cooler Temperature: 3.0/3.0°C Temp should be above freezing to 6°C (Initial/Corrected)	_		7. If temp. is over 6°C	or under 0°C, was the PN	1 notified?	∵ ☐ Yes	□ No
All	discrepand	cies will be	written out in the comments section below	v.			
	Yes	No			Yes	No	N/A
USDA Regulated Soils? (HI, ID, NY, WA, OR,CA, NM, TX, OK, AR, LA, TN, AL, MS, NC, SC, GA, FL, or Puerto Rico)		-	All containers needing acid/base pres. Have <u>CHECKED</u> ?: exceptions: VOA, coliform, LL container with a septum cap or preserved with	Hg, O&G, and any			
Short Hold Time Analysis (48 hours or less)? Analysis: DITC		_	Circle: HN03 (<2) H2SO4 (<2) NaOH (>10) NaO Any non-conformance to pH recommendations v	H/ZnAc (>9) vill be noted on the container			
Time 5035A TC placed in Freezer or Short Holds To Lab	Time:		Residual Chlorine Check (SVOC 625 Pest/P	CB 608)	Present	Absent	N/A
Rush TAT Requested (4 days or less):		-	Residual Chlorine Check (Total/Amenable/F	ree Cyanide)			_
Custody Signatures Present?	/		Headspace Wisconsin Sulfide?				_
Containers Intact?:	_		Headspace in VOA Vials (>6mm): See Containter Count form for details		Present	Absent	No VOA Vials Ser
Sample Label (IDs/Dates/Times) Match COC?: Except TCs, which only require sample ID	/		Trip Blank Present?			-	
Extra labels on Terracore Vials? (soils only)			Trip Blank Custody Seals?:			_	
COMMENTS:							

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000	DACE	-6
COC	PAGE	of

		SBS MeOH (only) BK	-																											
		Kit	-																							.2				ontainers mance **
COC Line Item	WGFU	R	Hegel Gegh	VOA VIAL HS (>6mm)	VG9U	DG90	VG9T	AGOU	AG1H	AG10	AG2U	AG3S	AG3SF	AG3C	BP1U	BP1N	BP2U	вьзп	BP3N	ВРЗЕ	BP3S	врзв	BP3Z	ССЗН	Syringe Kit	WSK	×	HNO3/	NaOH/	
1			3					2		2								2	1								ws	~		
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12																					-				-					

Container Codes

	Gla	SS			
DG9H	40mL HCl amber voa vial	BG1T	1L Na Thiosulfate clear glass	BP1B	1L NaOH plastic
DG9P	40mL TSP amber vial	BG1U	1L unpreserved glass	BP1N	1L HNO3 plastic
DG9S	40mL H2SO4 amber vial	BG3H	250mL HCI Clear Glass	BP1S	1L H2SO4 plastic
DG9T	40mL Na Thio amber vial	BG3U	250mL Unpres Clear Glass	BP1U	1L unpreserved plastic
DG9U	40mL unpreserved amber vial	AG0U	100mL unpres amber glass	BP1Z	1L NaOH, Zn, Ac
VG9H	40mL HCl clear vial	AG1H	1L HCl amber glass	BP2N	500mL HNO3 plastic
VG9T	40mL Na Thio. clear vial	AG1S	1L H2SO4 amber glass	BP2C	500mL NaOH plastic
VG9U	40mL unpreserved clear vial	AG1T	1L Na Thiosulfate amber glass	BP2S	500mL H2SO4 plastic
I	40mL w/hexane wipe vial	AG1U	1liter unpres amber glass	BP2U	500mL unpreserved plastic
WGKU	8oz unpreserved clear jar	AG2N	500mL HNO3 amber glass	BP2Z	500mL NaOH, Zn Ac
WGFU	4oz clear soil jar	AG2S	500mL H2SO4 amber glass	врзв	250mL NaOH plastic
JGFU	4oz unpreserved amber wide	AG2U	500mL unpres amber glass	BP3N	250mL HNO3 plastic
CG3H	250mL clear glass HCl	AG3S	250mL H2SO4 amber glass	BP3F	250mL HNO3 plastic-field filtered
BG1H	1L HCl clear glass	AG3SF	250mL H2SO4 amb glass -field filtered	BP3U	250mL unpreserved plastic
BG1S	1L H2SO4 clear glass	AG3U	250mL unpres amber glass	BP3S	250mL H2SO4 plastic
GN	General	AG3C	250mL NaOH amber glass	BP3Z	250mL NaOH, ZnAc plastic

	Plastic / Misc.						
7	BP4U	125mL unpreserved plastic	1				
	BP4N	125mL HNO3 plastic	1				
1	BP4S	125mL H2SO4 plastic	1				

Syringe Kit LL Cr+6 sampling kit

AF	Air Filter
С	Air Cassettes
R	Terracore kit
SP5T	120mL Coliform Na Thiosulfate
U	Summa Can
ZPLC	Ziploc Bag

WT		Water	
SL		Solid	
	OL	Non-aqueous liquid	Oil
WP		Wipe	P



RE-APPROVAL NOTICE

Customer Account: 601139 December 17, 2021

Vanessa Bravo VISION ENVIRONMENTAL, LLC 57 4TH STREET, UNIT B SOMERVILLE, NJ 08876

Thank you for selecting US Ecology ("USE") as your environmental management partner. In the event that a waste stream has not changed, the generator may use this form to re-approve the waste profile.

Generator Name: EXIDE ENVIRONMENTAL RESPONSE TRUST EPA ID No.: IND001647460

Waste Common Name: Drill Cuttings & PPE

Waste Code(s):

Approval No.: K196084DET Expiration Date: 11/13/2020

USE Facility Name & ID Number: EQ Detroit, Inc. (MID980991566)

This Re-approval Notice acknowledges the acceptability of waste material(s) into the noted USE facility(s) and ensures each facility has the appropriate permit(s) issued by federal and state regulatory agencies to properly transport, treat, and/or dispose of the waste material(s). Upon signature and submittal of this form, the waste stream will be reviewed by USE. The expiration will be extended for one year, unless you are contacted otherwise.

I certify that all information (including attachments) is complete and factual and is an accurate representation of the known and suspected hazards, pertaining to the waste described herein. I authorize USE to add supplemental information to the waste approval file, provided I am contacted and give verbal permission. I authorize USE to obtain a sample from any waste shipment for purposes of verification and confirmation. I agree that, if USE approves the waste described herein, all such wastes that are transported, delivered, or tendered to USE by Generator or on Generator's behalf shall be subject to, and Generator shall be bound by, the Standard Terms and Conditions associated with the original Waste Profile Form. (The Standard Terms and Conditions are incorporated into the Waste Profile Form as Page 4.)

Generator Signature:	Printed Name:
Company Name:	Date:

Please return this form via fax (800) 592-5329 or email customer.service@usecology.com. Questions? Please call (800) 592-5489.

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RE-APPROVAL NOTICE

Customer Account: 601139 December 17, 2021

Vanessa Bravo VISION ENVIRONMENTAL, LLC 57 4TH STREET, UNIT B SOMERVILLE, NJ 08876

Thank you for selecting US Ecology ("USE") as your environmental management partner. In the event that a waste stream has not changed, the generator may use this form to re-approve the waste profile.

Generator Name: EXIDE ENVIRONMENTAL RESPONSE TRUST EPA ID No.: IND001647460

Waste Common Name: Non Haz Soil

Waste Code(s):

Approval No.: L199075DET Expiration Date: 11/13/2020

USE Facility Name & ID Number: EQ Detroit, Inc. (MID980991566)

This Re-approval Notice acknowledges the acceptability of waste material(s) into the noted USE facility(s) and ensures each facility has the appropriate permit(s) issued by federal and state regulatory agencies to properly transport, treat, and/or dispose of the waste material(s). Upon signature and submittal of this form, the waste stream will be reviewed by USE. The expiration will be extended for one year, unless you are contacted otherwise.

I certify that all information (including attachments) is complete and factual and is an accurate representation of the known and suspected hazards, pertaining to the waste described herein. I authorize USE to add supplemental information to the waste approval file, provided I am contacted and give verbal permission. I authorize USE to obtain a sample from any waste shipment for purposes of verification and confirmation. I agree that, if USE approves the waste described herein, all such wastes that are transported, delivered, or tendered to USE by Generator or on Generator's behalf shall be subject to, and Generator shall be bound by, the Standard Terms and Conditions associated with the original Waste Profile Form. (The Standard Terms and Conditions are incorporated into the Waste Profile Form as Page 4.)

Generator Signature:	Printed Name:
Company Name:	Date:

Please return this form via fax (800) 592-5329 or email customer.service@usecology.com. Questions? Please call (800) 592-5489.

Rev. 04/19 Page 1 of 1 -1184431 - 1



RE-APPROVAL NOTICE

Customer Account: 601139 December 17, 2021

Vanessa Bravo VISION ENVIRONMENTAL, LLC 57 4TH STREET, UNIT B SOMERVILLE, NJ 08876

Thank you for selecting US Ecology ("USE") as your environmental management partner. In the event that a waste stream has not changed, the generator may use this form to re-approve the waste profile.

Generator Name: EXIDE ENVIRONMENTAL RESPONSE TRUST EPA ID No.: IND001647460

Waste Common Name: Drilling Water

Waste Code(s): 029L

Approval No.: K196082DET Expiration Date: 11/13/2020

USE Facility Name & ID Number: EQ Detroit, Inc. (MID980991566)

This Re-approval Notice acknowledges the acceptability of waste material(s) into the noted USE facility(s) and ensures each facility has the appropriate permit(s) issued by federal and state regulatory agencies to properly transport, treat, and/or dispose of the waste material(s). Upon signature and submittal of this form, the waste stream will be reviewed by USE. The expiration will be extended for one year, unless you are contacted otherwise.

I certify that all information (including attachments) is complete and factual and is an accurate representation of the known and suspected hazards, pertaining to the waste described herein. I authorize USE to add supplemental information to the waste approval file, provided I am contacted and give verbal permission. I authorize USE to obtain a sample from any waste shipment for purposes of verification and confirmation. I agree that, if USE approves the waste described herein, all such wastes that are transported, delivered, or tendered to USE by Generator or on Generator's behalf shall be subject to, and Generator shall be bound by, the Standard Terms and Conditions associated with the original Waste Profile Form. (The Standard Terms and Conditions are incorporated into the Waste Profile Form as Page 4.)

Generator Signature:	PrintedName:
Company Name:	Date:

Please return this form via fax (800) 592-5329 or email customer.service@usecology.com. Questions? Please call (800) 592-5489.

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APPENDIX F

Bio-Trap Protocols and Results



10515 Research Drive Knoxville, TN 37932 Phone: (865) 573-8188 Fax: (865) 573-8133





Client: Gregory Smoot Phone: 610-745-4624

Advanced GeoServices Corp

1055 Andrew Drive

Suite A

West Chester, PA 19380

Client Project #: 2020-4123-02 Client Project Name: Exide Trust - Frankfort

Fax:

Purchase Order #: 2020-4123-02

Test results provided for: CENSUS

Charles Slater

Reviewed By:

NOTICE: This report is intended only for the addressee shown above and may contain confidential or privileged information. If the recipient of this material is not the intended recipient or if you have received this in error, please notify Microbial Insights, Inc. immediately. The data and other information in this report represent only the sample(s) analyzed and are rendered upon condition that it is not to be reproduced without approval from Microbial Insights, Inc. Thank you for your cooperation.

Results relate only to the items tested and the sample(s) as received by the laboratory.

MICROBIAL INSIGHTS, INC.

10515 Research Dr., Knoxville, TN 37932

Tel. (865) 573-8188 Fax. (865) 573-8133

Client: **Advanced GeoServices Corp** Project: Exide Trust - Frankfort

041TC MI Project Number: 03/10/2022 Date Received:

Sample Information

Client Sample ID:		MW-1 MNA	MW-1 BioStim	MW-1 BioAug	MW-4 MNA	MW-4 BioStim	
			SRS	SRS SDC-9		SRS	
Sample Date:		03/09/2022	03/09/2022	03/09/2022	03/09/2022	03/09/2022	
Units:		cells/bead	cells/bead	cells/bead	cells/bead	cells/bead	
Analyst/Reviewer:		BB/CS	BB/CS	BB/CS	BB/CS	BB/CS	
hlorinating Bacteria							
Dehalococcoides	DHC	2.03E+04	8.45E+03	7.18E+06	9.12E+03	2.66E+04	
Dehalococcoides tceA Reductase	DHC TCE	2.03E+04 1.59E+03	8.45E+03 5.95E+02	7.18E+06 6.80E+05	9.12E+03 4.68E+02	2.66E+04 1.77E+03	

Legend:

NA = Not Analyzed NS = Not Sampled J = Estimated gene copies below PQL but above LQL I = Inhibited

CENSUS

< = Result not detected

MICROBIAL INSIGHTS, INC.

10515 Research Dr., Knoxville, TN 37932

Tel. (865) 573-8188 Fax. (865) 573-8133

Client: **Advanced GeoServices Corp** Project: Exide Trust - Frankfort

MI Project Number: Date Received:

041TC 03/10/2022 **CENSUS**

Sample Information

Client Sample ID: MW-4 BioAug MW-9 MNA MW-9 BioStim MW-9 BioAug SRS SDC-9 SRS SDC-9 SRS 03/09/2022 03/09/2022 03/09/2022 Sample Date: 03/09/2022 cells/bead cells/bead cells/bead cells/bead Units: BB/CS BB/CS BB/CS BB/CS Analyst/Reviewer:

Dechlorinating Bacteria

Dehalococcoides DHC 8.08E+03 4.28E+03 1.27E+07 3.00E+06 tceA Reductase TCE 1.28E+06 4.16E+02 3.66E+02 3.45E+05 **BAV1 Vinyl Chloride Reductase** BVC <2.50E+01 6.98E+01 <2.50E+01 7.80E+00 (J) Vinyl Chloride Reductase VCR 8.72E+05 4.89E+02 3.89E+02 2.42E+05

Legend:

J = Estimated gene copies below PQL but above LQL NA = Not Analyzed NS = Not Sampled I = Inhibited

< = Result not detected

Quality Assurance/Quality Control Data

Samples Received 3/10/2022

Component	Date Prepared	Date Analyzed	Arrival Temperature	Positive Control	Extraction Blank	Negative Control	
BVC	03/10/2022	03/16/2022	0 °C	105%	non-detect	non-detect	
TCE	03/10/2022	03/16/2022	0 °C	101%	non-detect	non-detect	
DHC	03/10/2022	03/16/2022	0 °C	100%	non-detect	non-detect	
VCR	03/10/2022	03/16/2022	0 °C	106%	non-detect	non-detect	

REPORT TO: Reports will be provid below will require prid	ded to the contact(s) listed below. Parties other than the contact(s) listed or approval.	INVOICE TO: For Invoices paid by a corresponding reference	third party it is imperative that contact information & se No. be provided.	microbial insights
Name:	greg Smoot	Name:		
Company:	Montrose Favikonmenter	Company:		10515 Research Drive.
Address:	1055 Andrew BR.	Address:		Knoxville TN 37932
	West chester PA 19380			phone (865) 573-8188 fax: (865) 573-8133
email:	9P5mooto mintrose - envicom	email:		email: info@microbe.com
Phone:	(6/0) 840-9142	Phone:	()	www.microbe.com
Fax:	(610) 840 -9199	Fax:	()	
Project Manager: Project Name:	SAN Abinsky Exide Toust - FRANK FORT	Purchase Order No. Subcontract No.	2020 -4123-02	Please Check One: ☐ More samples to follow ☐ No Additional Samples
Project No.:	2020 -4123 - 02			1
Daniel Times	M Standard (C Comprehensive (159/ gurehama)	□ Weterical /	200/	Saturday Delivery
Report Type:	Standard (Comprehensive (15% surcharge)	Li Historicai (3	30% surcharge)	Please see sampling protocol for instructions
Please contact us pri	ior to submitting samples regarding questions about the analyses you are requesting	at (865) 573-8188 (8:00 a	m to 4:00 pm M-F). After these hours please call (865) 300-8053	

	Samp	ole Information				MICRO		COC		GEO
MI ID Laboratory Use Only)	Monitoring Well	Unit Type	. Data Deployed	Date Retrieved	CENSUS DHC CENSUS BVC. TCE. VCR		COC Dissolved Gases (MFE)	Dissolved Gases (WEE)	Volatile Fatty Acids (VFAs) Anions	
041TC 1	MW-1	MNA (200	12/0/2	3/9/22	x x		X X	x	XX	
2	mw-1	BioStim SRS 1145	12/10/21	3/9/22	x x		X X	x	X X	
3	mlw-1	BioAug SRS SDC-9 1130	12/10/21	3/9/22	хх		X X	x	хх	
	MW-4 MW-4 MW-4	MNA 1300 BioStim SRS 1245 BioAug SRS SDC-9 1230	1 1/2	3/9/22 3/9/22	x x x x		x x x x x x x	X I	X X X X X X X X X X X X X X X X X X X X X X X X X X X X X X X X X X X X X X X X X X X X X X X	
7	MW-9 MW-9	MNA 1400 BioStim SRS 1345	12/10/21	3/9/22 3/9/22	x x		X X		X X X X	
9		BioAug SRS SDC-9 1330		3/9/2	хх		X X		x x	
elinquished by:							19	2 Hessale	3/10/22	

In order for analysis to be completed correctly, it is vital that chain of custody is filled out correctly & that all relative information is provided. Failure to provide sufficient and/or correct information regarding reporting, invoicing & analyses requested information may result in delays for which MI will not be liable. *additional cost and sample preservation are associated with RNA samples.



10515 Research Drive Knoxville, TN 37932 Phone: 865.573.8188 Fax: 865.573.8133 Web: www.microbe.com

SITE LOGIC Report

Bio-Trap In Situ Microcosm Study

Contact: Gregory Smoot Phone: 610-745-4624

Address: Advanced GeoServices Corp

1055 Andrew Drive Email: gpsmoot@montrose-env.com

Suite A

West Chester, PA 19380

MI Identifier: 041TC Report Date: March 25, 2022

Project: Exide Trust – Frankfort; 2020-4123-02

Comments:

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Executive Summary

A Bio-Trap® *In Situ* Microcosm (ISM) study was performed in monitoring wells MW-1, MW-4, and MW-9 to investigate whether the addition of an exogenous amendment and/or bioaugmentation would enhance the biodegradation of chlorinated ethenes. The ISM assemblies deployed in all three wells consisted of three Bio-Trap units each: (i) a control monitored natural attenuation (MNA) unit with no exogenous amendment, (ii) a BioStim unit amended with SRS as the electron donor, and (iii) a BioAug unit amended with SRS as the electron donor and the exogenous SDC-9 dechlorinating bacterial culture. Following the deployment period, the Bio-Trap units were recovered for CENSUS® analysis and quantification of contaminant concentrations, dissolved gases, volatile fatty acids (VFAs), and anions. A summary of the data is provided in Tables 1 - 3. Key observations from the results obtained for each *in situ* microcosm are described below.

MW-1 MNA, BioStim and BioAug Units

- The concentrations of *Dehalococcoides* (DHC) in the MNA and BioAug SRS SDC-9 units were measured at concentrations of 10⁴ cells/bead and 10⁶ cells/bead, respectively, which met the 10⁴ cells/mL density threshold proposed by Lu et al. as a screening criterion for generally useful rates of biological reductive dechlorination¹. However, the DHC concentration in the BioStim SRS unit (10³ cells/bead) was below the 10⁴ cells/mL density threshold proposed by Lu et al. (2006). DHC is capable of mediating the complete reductive dechlorination of tetrachloroethene (PCE) and trichloroethene (TCE) to ethene under anaerobic conditions.
- In addition, the TCE reductase gene and vinyl chloride reductase gene VCR were detected at higher concentrations in BioAug unit (10⁵ cells/bead, each) compared to the MNA unit (10³ cells/bead, each). However, in the BioStim unit, the TCE reductase gene and vinyl chloride reductase gene VCR were an order of magnitude lower (10² cells/bead, each) compared to the MNA unit. The vinyl chloride reductase gene BVC was below the detection limit under all conditions. Collectively, the microbial and functional gene data suggest that the potential for the complete reductive dechlorination of PCE and TCE to ethene at well MW-1 is moderate under MNA conditions, low in the BioStim unit, and high under the BioAug condition assessed.
- CENSUS® analysis indicated that the genetic potential for the anaerobic biodegradation of chlorinated ethenes was
 highest in the BioAug unit with SRS amendment with SDC-9 culture compared to the MNA and SRS BioStim units
 deployed at this well location.
- Contaminant analysis indicated that *cis*-1,2-DCE was the primary chlorinated ethene present in all units deployed in MW-1 and was detected at concentrations of 24.3 µg/L, 316 µg/L, and 278 µg/L, in MNA, BioStim and BioAug units, respectively. Vinyl chloride was the second highest contaminant detected in the BioStim (27.0 µg/L) and BioAug (28.6 µg/L) units, whereas in the MNA unit, it was TCE (23.5 µg/L). The elevated concentration of chlorinated compounds in the BioStim and BioAug units relative to the MNA unit may be due to vertical heterogeneity of contaminant distribution in the subsurface. The detection of vinyl chloride and ethene daughter products in all ISM units suggested that complete reductive dechlorination occurred during the deployment period.

¹ Lu, X., Wilson, J. T. & Kampbell, D. H. Relationship between *Dehalococcoides* DNA in ground water and rates of reductive dechlorination at field scale. Water Research **40**, 3131–3140 (2006).



- Dissolved methane ranged from 9 μ g/L in the MNA unit to 30 μ g/L in the BioAug unit, and the ethene concentration was less than 3 μ g/L in all units.
- Sulfate was detected at concentrations of 559 mg/L, 401 mg/L, 271 mg/L, in the MNA, BioStim and BioAug units, respectively. The presence of alternative electron acceptors such as sulfate suggests that dechlorinating bacteria may be competing with other hydrogen-consuming microorganisms (*e.g.*, sulfate-reducing bacteria) for shared electron donors.
- Acetic acid was noticeably higher in the BioStim (51 mg/L) and BioAug (110 mg/L) units, compared to the MNA unit (0.2 mg/L, below the practical quantitation limit). Lactic acid was also detected in the MNA (2 mg/L), BioStim (2.1 mg/L), and BioAug (0.72 mg/L) units. Concentrations of other volatile fatty acids were below 1 mg/L. These results suggest that microorganisms were actively fermenting the electron donor components of the SRS amendments.

MW-4 MNA, BioStim and BioAug Units

- DHC concentrations were noticeably higher in the BioStim and BioAug Units deployed in well MW-4 (detected at concentrations of 10⁴ cells/bead, and 10⁷ cells/bead respectively) compared to the MNA unit (10³ cells/bead). The higher DHC concentrations indicate an enhancement of the genetic potential for complete reductive dechlorination under SRS biostimulation and SDC-9 amendments.
- TCE and VCR reductase gene concentrations were detected on the order of 10² cells/bead in the MNA unit and 10³ cells/bead in the BioStim unit, whereas TCE and VCR reductase gene concentrations were measured at concentrations of 10⁶ cells/bead and 10⁵ cells/bead, respectively, in the BioAug unit. The BVC gene was detected at a similar concentration of 10¹ cells/bead in all units. The results indicate that the both SRS amendment and SDC-9 bioaugmentation stimulated the overall growth of DHC and functional genes.
- The primary chlorinated contaminant in all Bio-Trap ISM units was *cis*-1,2-DCE, followed by TCE. The concentrations of *cis*-1,2-DCE were measured at 216000 µg/L, 389000 µg/L and 145000 µg/L in the MNA, BioStim and BioAug units, respectively. Ethene was detected in all units at high concentrations ranging from 2600 µg/L to 3000 µg/L. These results suggest that complete reductive dechlorination to ethene occurred in all three Bio-Trap ISM units during the deployment period.
- Methane concentrations were 3100 $\mu g/L$, 2900 $\mu g/L$ and 2400 $\mu g/L$ in MNA, BioStim and BioAug units, respectively. In addition, sulfate was detected in all units at concentrations ranging from 16.5 mg/L to 32.1 mg/L. The geochemical data suggest that site-specific well conditions may have been strongly reducing in all ISM units deployed in MW-4.
- Only acetic acid was detected in the BioAug unit (14 mg/L), whereas the concentrations of the other volatile fatty
 acids, including lactic acids, pyruvic acid, propionic acid, and butyric acid were either below the detection limit or
 below the practical quantitation limit for all units deployed at the MW-4 well location.

MW-9 MNA, BioStim and BioAug Units

• In the MNA and BioStim Units deployed in well MW-9, DHC was detected at a similar concentration of 10³ cells/bead, whereas the DHC concentration was three orders of magnitude higher in the BioAug unit, indicating an enhancement of the genetic potential for complete reductive dechlorination under bioaugmentation with SDC-9 and SRS. TCE and VCR reductase genes were also higher in the BioAug unit (10⁵ cells/bead each) compared to



the MNA and BioStim units (10² cells/bead each), while BVC was either below the detection limit or below the practical quantitation limit in all units. These results indicate an increase in the genetic potential for the complete anaerobic reductive dechlorination of PCE and TCE under bioaugmentation with SDC-9 and SRS amendment at this well location.

- The primary chlorinated contaminant in all three Bio-Trap ISM units was cis-1,2-DCE, followed by vinyl chloride. cis-1,2-DCE measured at concentrations of 678 μ g/L, 590 μ g/L and 452 μ g/L in the MNA, BioStim and BioAug units, respectively. Ethene was detected at concentrations of 14 μ g/L, 12 μ g/L and 10 μ g/L in MNA, BioStim and BioAug units, suggesting that some complete reductive dechlorination had occurred in all three units. Ethane was also detected in all unit, at concentrations ranging from 74 μ g/L to 82 μ g/L.
- High methane concentrations were detected in the MNA ($1200 \,\mu g/L$), BioStim ($1400 \,\mu g/L$) and BioAug ($1800 \,\mu g/L$) units. Sulfate was only detected in the BioAug unit at a low concentration of 0.4 mg/L. The geochemical data suggest that site-specific well conditions may be reducing in all ISM units at this location.
- The VFA analysis indicated that acetic acid and propionic acid were detected in the MNA (51 mg/L, and 46 mg/L, respectively), BioStim (57 mg/L, and 48 mg/L, respectively) and BioAug (15 mg/L, and 18 mg/L, respectively) units. Concentrations of other volatile fatty acids were below 2 mg/L.



Overview of Approach

Site managers have frequently turned to laboratory microcosms or small pilot studies to evaluate bioremediation. However, duplication of *in situ* conditions in the laboratory is difficult and the results often do not correlate to the field. Pilot studies are performed in the field but are often prohibitively expensive as an investigative tool. Bio-Trap studies serve as cost-effective, *in situ* microcosms providing microbial, chemical, and geochemical evidence to evaluate biodegradation as a treatment mechanism and to screen remedial alternatives.

Typically each Bio-Trap Unit will contain samplers to evaluate the following:

Geochemical Fingerprint (GEO)

•40 mL VOA vial with a nylon screened cap designed for assessment of a variety of geochemical parameters including anions and metabolic acids.

Contaminant of Concern (COC)

 Passive diffusion bag designed for analysis of a variety of COCs including chlorinated solvents and petroleum hydrocarbons.

Microbial Populations (MICRO)

•PVC cassette containing Bio-Sep® beads, which provide a large surface area for microbial attachment and were designed for analysis by a variety of molecular biological tools (MBTs).

How does it work?

The MICRO sampler (microbial populations) contains Bio-Sep® beads, an engineered composite of Nomex® and powdered activated carbon which provides an incredibly large surface area (~600 m²/g) that is readily colonized by subsurface microorganisms. In addition to a matrix for microbial growth, the Bio-Sep® beads can be "baited" with amendments including electron donors (e.g. hydrogen releasing compounds) to investigate biostimulation approaches to enhance biodegradation. The Bio-Trap units also contain a COC (contaminant of concern) sampler to measure contaminant concentrations, daughter product formation, and dissolved gases and a GEO (geochemical fingerprint) sampler for quantification of geochemical parameters (nitrate, iron, sulfate, etc.), chloride production, and metabolic acids (pyruvic, lactic, acetic, propionic, etc.).

Bio-Trap® *In Situ* Microcosm studies at chlorinated solvent sites typically include three types of Bio-Trap Units deployed within a monitoring well. Each Bio-Trap Unit corresponds to one of the three most common remedial options: monitored natural attenuation (MNA), Biostimulation (BioStim), and Bioaugmentation (BioAug). All three Bio-Trap Units contain COC and GEO samplers for chemical and geochemical analyses. The key difference between the Bio-Trap Units is in the MICRO sampler.



Types of Bio-Trap Units typically deployed and MICRO sampler configurations:

Control

(MNA)

 \bullet Bio-Sep® beads contain no additional amendment and represent current aquifer conditions.

Biostimulation

(BioStim)

• An amendment supplier is used to release the desired specified electron donor (sodium lactate, molasses, EVO, etc.) or electron acceptor (oxygen release compound, sulfate, etc.).

Bioaugmentation

(BioAug)

•Bio-Sep® beads are pre-inoculated with a bioaugmentation culture, such as *Dehalococcoides*. These units can also be baited with an additional amendment.

MNA Unit: The purpose of the Control Bio-Trap Unit is to quantify contaminant degrading bacteria and daughter product formation under monitored natural attenuation (MNA) conditions and to serve as a baseline for comparison to BioStim and/or BioAug Units.

Following in-well deployment, DNA or phospholipid fatty acids (PLFA) can be extracted from the Bio-Sep beads for further analysis. For example, DNA extracted from the Bio-Sep beads can be used in CENSUS analysis of *Dehalococcoides* (DHC) and vinyl chloride reductase (*bvcA* and *vcrA*) genes to evaluate the potential for complete reductive dechlorination of PCE to ethene under MNA conditions. The VOC and anion samplers can be used to determine concentrations of contaminants, daughter products, dissolved gases, terminal electron acceptors, and chloride.

BioStim Unit: The Biostimulation Bio-Trap Unit is designed to test the hypothesis that electron donor addition will stimulate growth of dechlorinating bacteria and enhance biodegradation. As with the MNA Unit, the BioStim Unit contains COC and GEO samplers for chemical analyses. The BioStim Unit contains an amendment supplier to release the desired amendment over the incubation time.

BioAug Unit: The Bioaugmentation Bio-Trap Unit is designed to evaluate bioaugmentation as a treatment technology. The MICRO sampler contains Bio-Sep® beads pre-inoculated with the desired commercial culture. An amendment supplier may also be used to deliver an amendment. As with the MNA and BioStim Units, the BioAug Unit also contains a COC and GEO samplers for chemical analyses.

www.microbe.com



Results

Table 1. Summary of the results obtained for *In Situ* Microcosm Units.

Sample Information	MW-1	MW-1	MW-1
Treatment	MNA	BioStim SRS	BioAug SRS SDC-9
Sample Date	03/09/2022	03/09/2022	03/09/2022
MI ID	041TC-1	041TC-2	041TC-3
Microbial Populations (cells/bd)			
Dehalococcoides (DHC)	2.03E+04	8.45E+03	7.18E+06
tceA Reductase (TCE)	1.59E+03	5.95E+02	6.80E+05
bvcA Reductase (BVC)	< 2.50E+01	< 2.50E+01	< 2.50E+01
vcrA Reductase (VCR)	1.62E+03	7.11E+02	3.23E+05
Contaminant of Concern (µg/L)			
Tetrachloroethene	<5.00	<25.0	<25.0
Trichloroethene	23.5	8.8 J	7.0 J
cis-1,2-Dichloroethene	24.3	316	278
trans-1,2-Dichloroethene	0.5 J	1.2 J	<25.0
Vinyl chloride	2.8	27.0	28.6
Dissolved Gases (µg/L)			
Methane	9	16	30
Ethane	<1.0	<1.0	0.2 J
Ethene	2.7	1.3	1.1
VFAs (mg/L)			
Lactic Acid	2	2.1	0.72
Pyruvic Acid	< 0.50	< 0.50	< 0.50
Acetic Acid	0.2 J	51	110
Propionic Acid	< 0.50	< 0.50	< 0.50
Butyric Acid	< 0.50	0.5	0.5
Anions (mg/L)			
Chloride	2.5	5.3 J	6.1 J
Nitrate	< 0.4	< 0.4	< 0.4
Nitrite	< 0.4	< 0.4	< 0.4
Sulfate	559	401	271

Legend: NA = Not analyzed NS = Not sampled J = Estimated result below PQL but above LQL I = Inhibited <= Result not detected.



Table 2. Summary of the results obtained for *In Situ* Microcosm Units.

Treatment MNA BioStim SRS BioAug SRS SDC-9 Sample Date MI ID 03/09/2022 04/10 <th>Sample Information</th> <th>MW-4</th> <th>MW-4</th> <th>MW-4</th>	Sample Information	MW-4	MW-4	MW-4
MI ID 041TC-4 041TC-5 041TC-6	Treatment	MNA	BioStim SRS	BioAug SRS SDC-9
MI ID 041TC-4 041TC-5 041TC-6	Sample Date	03/09/2022	03/09/2022	03/09/2022
Dehalococcoides (DHC) 9.12E+03 2.66E+04 1.27E+07 tceA Reductase (TCE) 4.68E+02 1.77E+03 1.28E+06 bvcA Reductase (BVC) 5.75E+01 3.83E+01 6.98E+01 vcrA Reductase (VCR) 5.14E+02 2.17E+03 8.72E+05 Contaminant of Concern (μg/L) Tetrachloroethene <25000 <50000 <12500 Trichloroethene 69900 98700 11200 J Cis-1,2-Dichloroethene 216000 389000 145000 Trichloroethene <25000 <50000 <12500 Vinyl choride 11100 19200 J 4390 J Dissolved Gases (μg/L) Methane 3100 2900 2400 Ethane 1900 1500 1100 Ethane 1900 1500 100 VFAs (mg/L) Lactic Acid <10 <10 <10 Pyruvic Acid <10 <10 <10 <th>-</th> <th>041TC-4</th> <th>041TC-5</th> <th>041TC-6</th>	-	041TC-4	041TC-5	041TC-6
tecA Reductase (TCE)	Microbial Populations (cells/bd)			
bvcA Reductase (BVC) vcrA Reductase (VCR) 5.75E+01 3.83E+01 6.98E+01 vcrA Reductase (VCR) 5.14E+02 2.17E+03 8.72E+05	Dehalococcoides (DHC)	9.12E+03	2.66E+04	1.27E+07
VerA Reductase (VCR) 5.14E+02 2.17E+03 8.72E+05 Contaminant of Concern (μg/L) Contaminant of Concern (μg/L) 5.0000 <12500	tceA Reductase (TCE)	4.68E+02	1.77E+03	1.28E+06
Contaminant of Concern (μg/L) Tetrachloroethene <25000	bvcA Reductase (BVC)	5.75E+01	3.83E+01	6.98E+01
Tetrachloroethene <25000	vcrA Reductase (VCR)	5.14E+02	2.17E+03	8.72E+05
Trichloroethene 69000 98700 11200 J cis-1,2-Dichloroethene 216000 389000 145000 trans-1,2-Dichloroethene <25000	Contaminant of Concern (µg/L)			
cis-1,2-Dichloroethene 216000 389000 145000 trans-1,2-Dichloroethene <25000	Tetrachloroethene	<25000	<50000	<12500
trans-1,2-Dichloroethene <25000 <50000 <12500 Vinyl chloride 11100 19200 J 4390 J Dissolved Gases (μg/L) Secondary (μg/L) Secondary (μg/L) Secondary (μg/L) Secondary (μg/L) 2900 2400 2400 2400 2400 2400 2600 2600 2600 2700 2600 2600 2600 2700 2600 </td <td>Trichloroethene</td> <td>69000</td> <td>98700</td> <td>11200 J</td>	Trichloroethene	69000	98700	11200 J
Vinyl chloride 11100 19200 J 4390 J Dissolved Gases (μg/L) Stanta 2900 2400 Ethane 1900 1500 1100 Ethene 3000 2700 2600 VFAs (mg/L) Lactic Acid <10	cis-1,2-Dichloroethene	216000	389000	145000
Dissolved Gases (μg/L) Methane 3100 2900 2400 Ethane 1900 1500 1100 Ethene 3000 2700 2600 VFAs (mg/L) Lactic Acid <10	trans-1,2-Dichloroethene	<25000	< 50000	<12500
Methane 3100 2900 2400 Ethane 1900 1500 1100 Ethene 3000 2700 2600 VFAs (mg/L) Lactic Acid <10	Vinyl chloride	11100	19200 J	4390 J
Ethane 1900 1500 1100 Ethene 3000 2700 2600 VFAs (mg/L) Lactic Acid <10 <10 <10 <10 <10 <10 <10 <10 Anous (mg/L) <10 <10 <10 <10 <10 <10 <10 <10 <10 <10 <10 <10 <10 <10 <10 <10 <10 <10 <10 <10 <10 <10 <10 <10 <10 <10 <10 <10 <10 <10 <10 <10 <10 <10 <10 <10 <10 <10 <10 <10 <10 <10 <10 <10 <10 <10 <10 <10 <10 <10 <10 <10 <10 <10 <10 <10 <10 <10 <10 <10 <10 <10 <10 <10 <10 <10 <10 <10 <10 <10 <10 <10 <10 <10 <10 <10 <10 <10 <10 <10 <10<	Dissolved Gases (μg/L)			
Ethene 3000 2700 2600 VFAs (mg/L) Lactic Acid <10	Methane	3100	2900	2400
VFAs (mg/L) Lactic Acid <10	Ethane	1900	1500	1100
Lactic Acid <10	Ethene	3000	2700	2600
Pyruvic Acid <10	VFAs (mg/L)			
Acetic Acid 8.8 J 10.0 J 14 Propionic Acid <10	Lactic Acid	<10	<10	<10
Propionic Acid <10 7.4 J 6.6 J Butyric Acid <10	Pyruvic Acid	<10	<10	<10
Butyric Acid <10 <10 <10 Anions (mg/L) 378 440 549 Nitrate <20.0	Acetic Acid	8.8 J	10.0 J	14
Anions (mg/L) Chloride 378 440 549 Nitrate <20.0	Propionic Acid	<10	7.4 J	6.6 J
Chloride 378 440 549 Nitrate <20.0	Butyric Acid	<10	<10	<10
Nitrate <20.0 <0.4 <40.0 Nitrite <20.0	Anions (mg/L)			
Nitrite <20.0 <20.0 <1.0	Chloride	378	440	549
	Nitrate	<20.0	< 0.4	<40.0
Sulfate 32.1 30.9 16.5	Nitrite	<20.0	<20.0	<1.0
	Sulfate	32.1	30.9	16.5

Legend: NA = Not analyzed NS = Not sampled J = Estimated result below PQL but above LQL I = Inhibited <= Result not detected.



Table 3. Summary of the results obtained for *In Situ* Microcosm Units.

Sample Information	MW-9	MW-9	MW-9
Treatment	MNA	BioStim SRS	BioAug SRS SDC-9
Sample Date MI ID	03/09/2022 041TC-7	03/09/2022 041TC-8	03/09/2022 041TC-9
Microbial Populations (cells/bd)			
Dehalococcoides (DHC)	8.08E+03	4.28E+03	3.00E+06
tceA Reductase (TCE)	4.16E+02	3.66E+02	3.45E+05
bvcA Reductase (BVC)	< 2.50E+01	7.80E+00 J	< 2.50E+01
vcrA Reductase (VCR)	4.89E+02	3.89E+02	2.42E+05
Contaminant of Concern (µg/L)			
Tetrachloroethene	< 500	<250	<125
Trichloroethene	< 500	<250	<125
cis-1,2-Dichloroethene	678	590	452
trans-1,2-Dichloroethene	< 500	<250	<125
Vinyl chloride	146 J	110	173
Dissolved Gases (µg/L)			
Methane	1200	1400	1800
Ethane	82	74	74
Ethene	14	12	10
VFAs (mg/L)			
Lactic Acid	0.7	1.1	<10
Pyruvic Acid	0.2 J	0.2 J	<10
Acetic Acid	51	57	15
Propionic Acid	46	48	18
Butyric Acid	0.6	0.9	<10
Anions (mg/L)			
Chloride	17.4	41.9	19
Nitrate	0.1 J	<10.0	<10.0
Nitrite	<0.2	<0.2	0.1 J
Sulfate	<0.2	< 0.2	0.4

Legend: NA = Not analyzed NS = Not sampled J = Estimated result below PQL but above LQL I = Inhibited <= Result not detected.



Glossary

Amendment Supplier: a component that fits inside the Bio-Trap unit at the top and/or bottom. This component is designed to slowly diffuse a desired amendment within a BioStim and/or a BioAug Unit during the incubation time.

Sampler: Individual components consisting either of a geochemical (GEO), contaminant of concern (COC) or microbial (MICRO) sampler. Geochemical samplers are essentially VOA vials with special septa that facilitate transfer. The microbial samplers are made from a smaller PVC pipe \sim 1" x 3 ½" and contains Bio-Sep® beads which serve as a microbial growth matrix.

COC Sampler: a passive diffusion bag designed for analysis of a variety of COCs, including chlorinated solvents and petroleum hydrocarbons

GEO Sampler: a 40 mL amber VOA with a nylon-based membrane permitting passive diffusion of anionic species

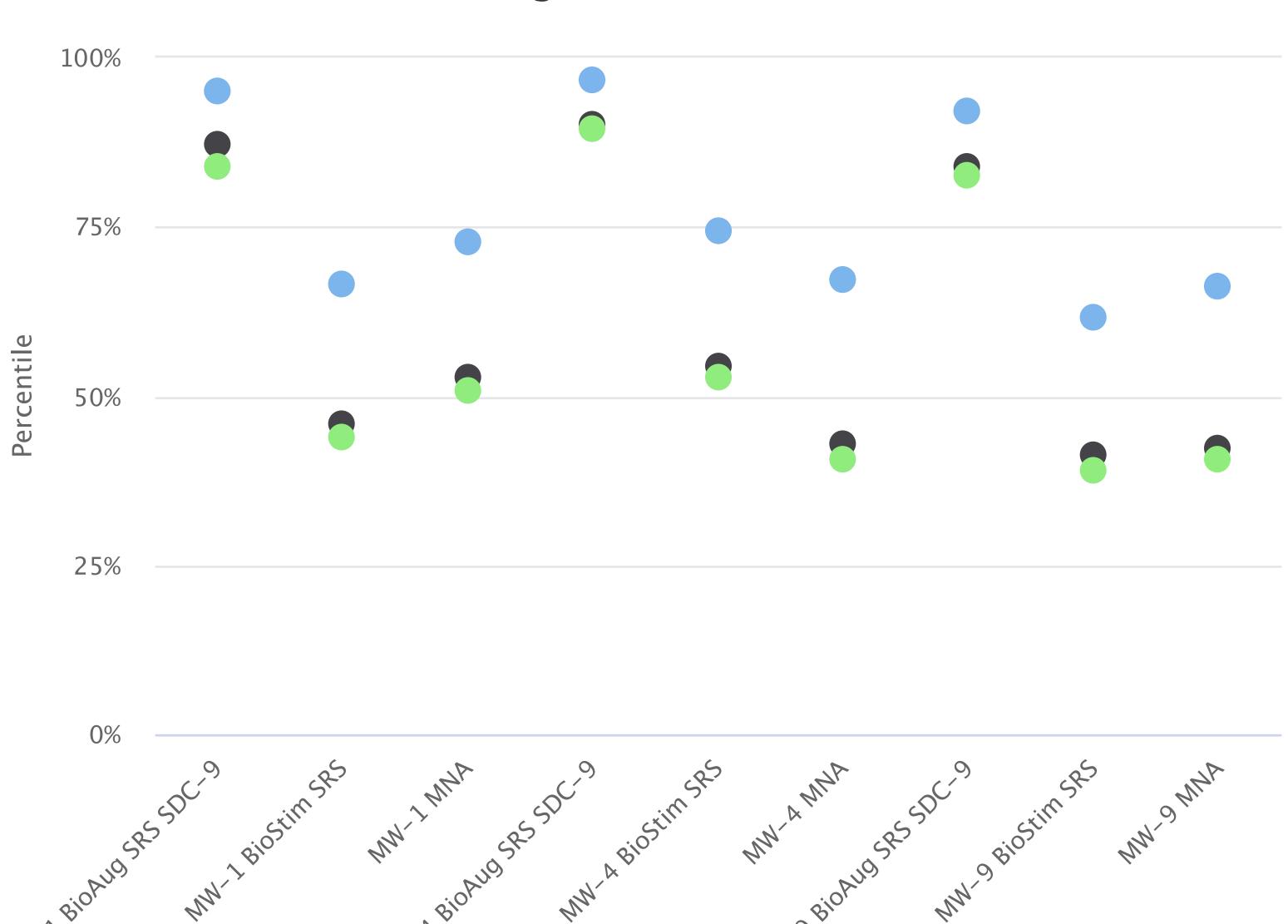
MICRO Sampler: a polyvinylchloride cassette containing Bio-Sep® beads which provide a large surface area for microbial growth. In addition to a matrix for microbial growth, the Bio-Sep® beads can be "baited" with bioaugmentation cultures or ¹³C-labeled compounds. Bio-Sep® beads were designed to allow extraction of phospholipids fatty acids and DNA for analysis of microbial communities.

Unit: 1.25" x 15" PVC housing that all of the samplers are place into for deployment. Units will have baffled end caps to separate different zones within the monitoring well. Typically, each unit will correspond to a treatment approach.

Assembly: Collections of Units for a particular monitoring well. Samplers (GEO, COC, and MICRO) are placed in each unit. Units are linked to form an Assembly. An entire Assembly (consisting of multiple units) is deployed in each well.

10

Target Percentiles



Sample





SAMPLING INSTRUCTIONS

Storage:

It is important to minimize the amount of time that Bio-Trap Samplers are stored prior to being installed in the field. The physical properties of the Bio-Trap Samplers that make them an ideal medium for collecting microbes also increase the chances of microbial or chemical contamination. **Bio-Trap Samplers need to remain sealed and refrigerated** (not frozen) until they can be installed in the field.

Note: Clean sterile gloves should be used at all times when handling Bio-Trap Samplers.

Installation:

- Prior to installing Bio-Trap Units, the monitoring well may need to be purged if it has not been sampled recently. If purging is
 necessary, MI recommends that three well volumes be removed to ensure contact with formation water and reduce well bore
 effect.
- Assemble the Bio-Trap Units as described in this protocol. Attach the top Bio-Trap Unit to a nylon line (not provided) and
 suspend the units at a depth where significant contaminant concentrations exist. If data is not available on the vertical
 distribution of contaminants, suspend the Bio-Trap Units in the middle of the saturated screened interval. If deploying multiple
 Bio-Trap Units, be sure that all units are placed within the screened interval and that they remain covered by water for the
 entire deployment period.
- Be sure not to suspend the bio-trap in the NAPL zone.

Helpful Hints

Do:

- Do wear gloves when handling all Bio-Trap components.
- Do remove and discard the clear end cap covers from all vials.
- Do keep assembled Bio-Traps in an upright position.

Do not:

- Do NOT remove the units from the well during the entire incubation period. Cross-talk can result from any disturbance to the units.
- Do NOT prepare multiple units at the same time. Preparing each unit separately minimizes the potential for mistakes to occur.
- Do NOT cut the rubber baffles. The baffles are sized to the well diameter to prevent cross-talk between the Units.

Remember:

- Amber vials are samplers
- Clear vials and sponges are suppliers
- Color dictates type of sampler or supplier



1

Supply List - Deployment

Following is a table detailing the supplies that were sent for deployment of the bio-trap units and the colors used for the amendment suppliers.

No.	Item	Image
9	Housing Unit	Commont.
3	Weight	
10	40 mL GEO Sampler*	
10	COC Sampler*	
6	Standard MICRO Sampler	
3	SDC-9 MICRO Sampler	
12	SRS Amendment Supplier	
3	Gray PVC Spacer	

^{*} Extra samplers provided in case of breakage during shipping



Terminology

Samplers are placed in each slotted PVC housing unit. Housing units (e.g. Control, BioStim-LACTATE, BioAug-DHC, etc.) are linked to form an Assembly. An entire Assembly (consisting of 2 or more units) is deployed in each well. **See Figure 1.**

Assembly: An assembly is a collection of two or more units (20" PVC housings) for deployment in a particular monitoring well.

Unit: A unit is a 1.25" x 20" slotted PVC housing to hold samplers. Units have baffled end caps to separate different zones within the monitoring well and are sized according to the well diameter. Typically each unit will correspond to a treatment (labeled "Control-MNA", "BioStim-LACTATE", "BioAug-DHC"). The metal nameplate is on the bottom of the Unit.

Samplers: ALL units will contain three samplers – One geochemical fingerprint sampler (GEO), one contaminant of concern sampler (COC), and one microbial population sampler (MICRO).

- The GEO sampler is a 40 mL <u>amber</u> VOA vial, with <u>Teal screw cap</u>.
- The COC sampler is a ~ 6 ½" Passive Diffusion Bag.
- The MICRO sampler is a ~ 1" x 1 1/2" slotted PVC pipe containing Bio-Sep beads.

Amendment Supplier: BioStim and/or BioAug units will include an "Amendment Supplier" which provides an electron donor (e.g. LACTATE, EOS), or an electron acceptor (e.g. ORC, PermeOx, nitrate, sulfate), or a nutrient source intended to stimulate microbial activity. Depending on the type of amendment, the amendment supplier will be one of the following:

- a sponge, or
- a nylon pouch, or
- a clear VOA vial

A sponge placed in the bottom of the unit is typically used as the amendment supplier for viscous amendments such as LACTATE or EOS. For powdered or granular amendments, a nylon pouch is often used as the amendment supplier. A clear VOA vial is used for dissolved amendment solutions. The Control-MNA units have a solid gray PVC spacer as a substitute for the amendment supplier.

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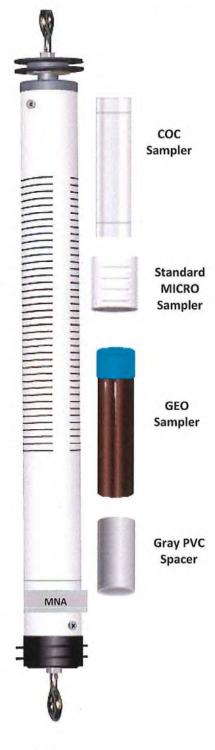


Pre-deployment Instructions

- Remove the clear end cap covers from all glass VOA vials. Colored screw caps should NOT be removed.
- The BOTTOM of the unit has the metal nameplate (e.g. MNA, LACTATE, BioAug-DHC, etc.)

Control - MNA unit assembly

- Twist off the bottom of unit labeled "MNA" to fill with samplers.
- · First, place the COC Sampler in the housing unit.
- Next, insert the Standard MICRO sampler. It does not matter which end goes in first. Be sure to wear gloves.
- Then, place the 40 mL amber VOA vial with the Teal screw cap (GEO Sampler) into the housing so that the cap is facing the MICRO sampler and the top of the unit.
- Please insert one gray PVC spacer as a substitute for an amendment supplier.
- Hand-tighten the threaded cap back onto unit until it is secure. Remember to store the unit in an upright position.
- A total of three (3) units of this configuration will be required.



Unit



Bio-Stim - SRS

- Twist off the bottom of unit labeled "BIOSTIM SRS" to fill with samplers.
- · First, place one of the SRS amendment supplier sponges into the unit.
- Then, place the COC Sampler in the housing unit.
- Next, insert the MICRO sampler. It does not matter which end goes in first.
 Be sure to wear gloves.
- Next, place the 40 mL VOA vial with the Teal screw cap (GEO Sampler) into the housing so that the cap is facing the MICRO sampler and the top unit.
- · Next, insert another SRS amendment supplier sponge into the unit.
- Carefully hand-tighten the threaded cap back onto unit until it is secure.
 Remember to store the unit in an upright position.
- A total of three (3) units of this configuration will be required.



Unit



Bio-Aug - SRS, SDC-9 culture

- Twist off the bottom of unit labeled "BIOAUG SRS SDC-9" to fill with samplers.
- · First, place one of the SRS amendment supplier sponges into the unit.
- · Then, place the COC Sampler in the housing unit.
- Next insert the SDC-9 MICRO sampler. It does not matter which end goes in first.

Be sure to wear gloves.

- Next, place the 40 mL VOA vial with the Teal screw cap (GEO Sampler) into the housing so that the cap is facing the MICRO sampler and the top unit.
- Next, insert another SRS amendment supplier sponge into the unit.
- Carefully hand-tighten the threaded cap back onto unit until it is secure.
 Remember to store the unit in an upright position.
- A total of three (3) units of this configuration will be required.



Unit



Bio-Trap Assembly and Deployment

Once all PVC units contain the required samplers, the units can be connected to form an assembly for in well deployment.

- Control-MNA Unit The control unit is usually placed at the top of the assembly (shallowest position in the well). Be sure to securely attach a nylon rope or cable to the top of this unit to suspend the assembly in the well. BioStim units will be attached with metal fasteners to the bottom of this unit.
- BioStim-Treatment A Unit Using the metal fasteners, attach the top of this unit to the bottom of the unit above. Repeat
- · Repeat as necessary

Please note: Microbial Insights will send a separate shipment arriving on a later date with the retrieval supplies.

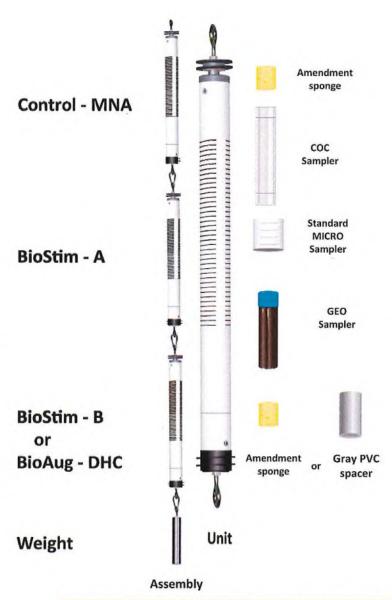


Illustration of Bio-Trap Assembly, Unit, and Samplers. Actual configuration may vary depending on the number of units per well

Fax: 865.573.8133 www.microbe.com



Supply List – Return Kit

Following is a list of the supplies that are being shipped separately (from the deployment supplies) for retrieval of the Bio-Trap Units and their return to Microbial Insights for analysis.

Item
40mL clear VOA w/White Cap TSP preserved
40 mL clear VOA w/Red Cap HCI preserved
GEO Sampler w/ Teal return Caps
Clear ~ 5" straws for COC sampler (use one per bag)
Labels for COC and GEO samplers
Bubble Bags for VOA vials
Small Zippered Bag labeled for MICRO samplers
Large Clear Zippered Bags for Unit Samplers (COC, GEO, and MICRO)
MI Chain of Custody Form



Bio-Trap Assembly Retrieval

- After the desired incubation period, carefully retrieve the assembly and remove the nylon line and the weights. Units contain
 glass vials that can be broken so please handle with care.
- Separate the Bio-Trap housing Units from each other and keep in an upright position.
- Open one Unit at a time and carefully remove all contents. DO NOT DISCARD THE PVC HOUSING UNIT.
- Label and Package all contents from one unit before opening another unit.
- Label the vials with the provided preprinted labels <u>directly onto all vials</u>. It is ok if the label overlaps the writing or covers the cap. Please include the well and unit type (Control, BioStim, BioAug, etc.) Please do not place the labels on the bubble wrap.
- Place the MICRO sampler (1" x 1 ½" slotted PVC containing bio-sep beads) into the silver zippered bag (provided) and label appropriately.
- Two 40-mL preserved vials are provided per unit to be filled with the water from the COC sampler bag.
 - o First, open the cap for the 40mL HCl preserved vial (with red cap) and, holding the COC sampler bag upright above the vial, poke the bag with one end of the clear straw provided, making sure that the other end is positioned inside the vial. This ensures the flow of the liquid from the bag to the vial. Be careful not to spill any liquid. Fill the vial completely so that there is no head space / air bubble and then screw the cap back on it. Please ensure COC vial is full before proceeding to the next step.
 - Next, fill the 40 mL TSP preserved vial (with white cap) in the same way.
- Then, remove the Teal screw cap from the 40 mL VOA vial (GEO sampler) and replace with the extra blue screw cap provided. Please note that it is okay for the GEO sampler to be exposed to air. The original membrane will be removed with the cap.
- Sponge and nylon pouch amendment suppliers can be discarded as investigation derived waste.
- Once all screw caps have been replaced, tightly wrap each vial in a bubble wrap bag.
- Place all of the vials and MICRO sampler from one unit into a large clear zippered bag so that <u>all samplers from a unit are in</u> one bag.
- Place large clear bag containing all samplers within a cooler packed with ice as quickly as possible after retrieval and for shipping.
- Complete the MI chain of custody form.
- Ship all samplers along with the chain of custody form to Microbial Insights on ice. Ship the Bio-Trap housing units, weights, well caps, and PVC spacers to Microbial Insights separately. See shipping instructions below:

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Shipping Instructions

MICRO samplers and all VOA vials need to be shipped on ice (or blue ice) for next day delivery. If regular ice is used, the <u>ice should be double bagged</u>. The Bio-Trap housing units, PVC spacers, well caps, and weights must be returned to Microbial Insights but should be shipped separately without ice.

Samples should be shipped to:

Sample Custodian Microbial Insights, Inc. 10515 Research Drive Knoxville, TN 37932-2536 (865) 573-8188

Saturday Delivery:

Due to the short hold time associated with this study, shipping samples for Saturday Delivery is not recommended.

MI Sample Cancellation Policy

MI understands that on rare occasions our clients may want to cancel an order or individual samples. However, because samples are processed promptly upon receipt the following cancellation surcharges will apply. If cancellation is within 24 hours of sample receipt there will be no charge. If cancellation is requested 24 hours up to 3 working days there will be a 50% surcharge to cover our processing to that point. After 3 working days the full charge will apply.

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		BioAug SRS SDC-9			×	×	-				^	×						×	×	_				
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In order for analysis to be completed correctly, it is vital that chain of custody is filled out correctly & that all relative information is provided. Failure to provide sufficient and/or correct information regarding reporting, invoicing & analyses requested information may result in delays for which MI will not be liable. * additional cost and sample preservation are associated with RNA samples.

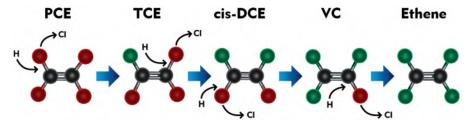


DHC Interpretation

1

Dehalococcoides 16S rRNA gene (qDHC)

Under anaerobic conditions, tetrachloroethene (PCE) and trichloroethene (TCE) can undergo sequential reductive dechlorination through the daughter products *cis*-dichloroethene (*cis*-DCE) and vinyl chloride to nontoxic ethene (1,2).



While a number of bacterial cultures capable of utilizing PCE and TCE as growth supporting electron acceptors have been isolated (3-7), *Dehalococcoides* spp. may be the most important because they are the only bacterial group that has been isolated to date which is capable of complete reductive dechlorination of PCE to ethene (8). In fact, the presence of *Dehalococcoides* spp. has been associated with complete dechlorination to ethene at sites across North America and Europe (9).

Status	Dehalococcoides spp.	Observation
	≥ 10 ⁴	Lu et al. proposed that a concentration of 1×10^4 DHC cells/mL could be used as a screening criterion to identify sites where reductive dechlorination will yield a generally useful biodegradation rate (10).
	(cells/mL)	Similarly, in an internal study conducted with nearly 1000 groundwater samples obtained from sites across the US, ethene production was observed in approximately 80% of samples in which CENSUS® qDHC results were greater than or equal to 10^4 DHC cells/mL.
	10 ¹ to < 10 ⁴	When vinyl chloride reductase genes (See DHC functional genes discussion below) are also detected, complete reductive dechlorination of PCE and TCE to ethene may still occur even with moderate DHC concentrations.
	(cells/mL)	When the DHC population is below the 10 ⁴ cells/mL criterion proposed by Lu et al. (10), project managers should carefully consider other site-specific data to determine whether subsurface conditions may be limiting reductive dechlorination. For example, the addition of an electron donor may be able to stimulate DHC growth and enhance anaerobic bioremediation.
	< 10 ¹ (cells/mL)	DHC concentrations are low suggesting that complete reductive dechlorination of PCE and TCE to ethene is unlikely to occur under existing conditions. Enhanced anaerobic bioremediation options (biostimulation or bioaugmentation) may need to be considered.

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DHC Functional Genes (tceA, bvcA, vcrA)

A "stall" where daughter products cis-DCE and vinyl chloride accumulate can occur at PCE- and TCE-impacted sites especially under MNA conditions. The accumulation of vinyl chloride, generally considered more carcinogenic than the parent compounds, is particularly problematic. Although elevated Dehalococcoides concentrations correspond to ethene production in numerous studies, the range of chlorinated ethenes metabolized and cometabolized varies among species and strains within the Dehalococcoides genus. For example, Dehalococcoides ethenogenes str. 195 metabolizes PCE, TCE, and cis-DCE and cometabolizes vinyl chloride (8) to produce ethene. Conversely, Dehalococcoides sp. CBDB1 utilizes PCE and TCE but does not cometabolize additional chloroethenes (11). Other Dehalococcoides strains, such as BAV1, GT and VS, are known to fully dechlorinate cis-DCE and VC to ethene (14,16,19). Quantification of reductive dehalogenase genes is used to more definitively confirm the potential for reductive dechlorination of TCE, cis-DCE, and vinyl chloride (12-15).

Functional Gene	Observation
TCE Reductase	
<i>tce</i> A gene	The <i>tce</i> A gene encodes the enzyme responsible for reductive dechlorination of TCE to <i>cis</i> -DCE in some strains of <i>Dehalococcoides</i> .
	Absence of <i>tce</i> A does not preclude the potential for reductive dechlorination of TCE in the field since the <i>tce</i> A gene is not universally distributed among all DHC and is not present in other microorganisms capable of reductive dechlorination of TCE (e.g. <i>Dehalobacter</i>).
	Detection of the <i>tce</i> A gene provides an additional line of evidence indicating the potential for dechlorination of TCE.
Vinyl Chloride Redu	ctase
<i>bvc</i> A gene	The <i>bvc</i> A gene encodes the vinyl chloride reductase enzyme responsible for reductive dechlorination of vinyl chloride to ethene by <i>Dehalococcoides</i> sp. str. BAV1 (16).
	Presence of bvcA gene indicates the potential for reductive dechlorination of VC to ethene.
	Absence of both bvcA and vcrA genes suggests VC may accumulate.
	An internal study with $^{\sim}1,000$ samples showed ethene production was observed in 80% of the samples that the DHC population was greater than or equal to 10^4 cells/mL. The <i>bvc</i> A gene was detected in over 50% of these samples.
	Van Der Zaan et al (17) noted that the bvcA gene was the only VC reductase gene detected at three of their sites.
	Alfred Spormann's laboratory at Stanford University (18) reported that the <i>bvc</i> A gene was the most abundant and active at the outflow of a PCE fed column study. This section of the column was in the DCE to VC stages of reductive dechlorination thus confirming the importance of the <i>bvc</i> A gene for complete reductive dechlorination.
<i>vcr</i> A gene	The <i>vcr</i> A gene encodes the vinyl chloride reductase enzyme responsible for reductive dechlorination of <i>cis</i> -DCE and vinyl chloride by <i>Dehalococcoides</i> sp. strain VS (14).

Presence of vcrA gene indicates the potential for reductive dechlorination of DCE and/or VC to ethene.

As with the bvcA gene, detection of the vcrA gene is associated with ethene production in internal studies (67%) and

Absence of both bvcA and vcrA genes suggest VC may accumulate.

vinyl chloride reduction in independent studies (14, 17).

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Reporting

3

Microbial Insights can provide a variety of data packages and reporting levels to suit the needs of any project. Data packages range from simple analytical reports with results only to more complex data packages that include a report narrative, analytical results, QC data, and supporting materials including all raw data and chain-of-custody documentation. The figure below shows our standard report and explains the way values are reported.

Microbial Insights, Inc.

CENSUS 2340 Stock Creek Blvd. Rockford, TN 37853-3044 Tel. (865) 573-8188 Fax. (865) 573-8133 **Client:** Company Name Unique Laboratory Identifier **MI Project Number:** Project: Your Project Name Date Samples Arrived Date Received: Sample Information Client Sample ID: Sample A Sample B Sample C 00/00/0000 00/00/0000 00/00/0000 Sample Date: cells/mL cells/mL cells/mL Units: "J" value Analyst: Intials Intials Intials Result is an estimated value. This data qualifier (flag) is used Dechlorinating Bacteria when the target gene is Dehalococcoides spp. DHC 1.84E+05 2.76E+02 2.28E+01 (J) detected but at a concentration or abundance below the practical quantification limit **Functional Genes** (PQL). tceA Reductase TCE 6.00E+01 3.23E+01 <4.00E-01 bvcA Reductase <4.00E-01 BVC 1.17E+04 1.81E+01 vcrA Reducatase <4.00E-01 VCR 8.42E+04 1.74E+02 Legend: J = Estimated gene copies below PQL but above LQL NA = Not Analyzed NS = Not Sampled I = Inhibited < = Result not detected < value The target gene was not detected at the limit of "I" value quantitation (LOQ) reported for that sample. QA Procedure indicated that the sample may have exhibited PCR inhibition. Although relatively rare, PCR inhibition can occur due to the presence of metals or humic acids at high concentrations in the sample.



Quality Assurance

Microbial Insights' comprehensive Quality Assurance (QA) Program is the foundation of all laboratory analyses, ensuring that our clients receive high-quality analytical services that are timely, reliable, and meet their intended purpose in a cost effective manner. MI is committed to providing quality data that surpasses regulatory and industry standards, thus enabling the client to make well-informed decisions. MI maintains strict standard operating procedures and QA/QC measures throughout all of the analyses offered. The following Table details specific QA/QC procedures that are used for CENSUS.

QA/QC	Description
Date of Extraction	DNA and RNA extractions are performed the day the samples are received by MI to minimize the possibility of any changes to the microbial community prior to analysis.
Laboratory Method Blanks	An extraction blank (no sample added) is processed alongside each set of field samples from DNA extraction through CENSUS® analysis to ensure that cross contamination has not occurred. Although MI has never experienced this issue, the detection of the CENSUS® target (e.g. Dehalococcoides) in an extraction blank is direct evidence of cross contamination with a sample or contamination of a reagent and would invalidate the results. If this were to occur, MI would re-extract the sample. If not possible to re-extract, MI would contact the client immediately and notate it on the laboratory report.
Laboratory Control Samples (LCS)	A laboratory control sample (LCS) or positive control (target DNA) is included with each CENSUS® plate to confirm amplification and as a continuing calibration check.
Negative Controls	A negative control (no DNA) is included with each CENSUS plate to ensure that cross contamination has not occurred during amplification. As with the extraction blank, detection of CENSUS target (e.g. DHC) in a negative control is direct evidence of contamination and would invalidate the results. If this were to occur, MI would rerun the analysis.

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How to Use Estimated Percentile Ranks from the Microbial Insights Database

The MI Database and Client Portal

The Microbial Insights Database is the largest collection of field concentrations of key microorganisms and functional genes currently containing qPCR and QuantArray results for more than 32,000 unique groundwater, soil, and sediment samples from all 50 states and 33 countries worldwide. Driven by field samples, the database reflects the impacts of common contaminants, geochemical conditions, and site management practices on critical microbial populations.

With your report, you received a passcode enabling you to retrieve estimates of the <u>percentile ranks</u> of your results based on those compiled in the MI database at <u>no additional charge</u>. When accessing the database, you will be asked to provide background information about the sample (e.g. contaminant concentrations) to aid in understanding the links between environmental conditions and microbial populations. As with all client information provided to MI, site specific data will be treated as confidential.

Is that low, medium or high?

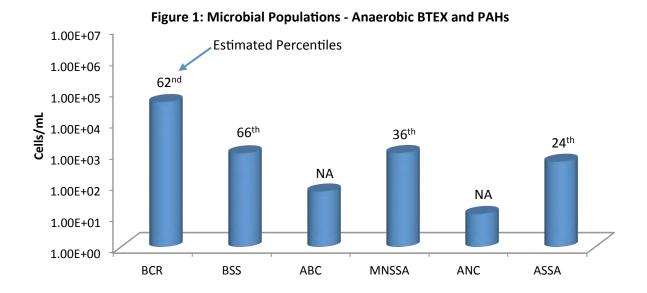
In practice, biodegradation depends not just on the presence but the <u>actual concentrations</u> of the contaminant degrading microorganisms. Simply put, qPCR and QuantArray results demonstrating high concentrations of target microorganisms or functional genes suggest in situ selection, enrichment and growth of those specific contaminant degraders and therefore a greater probability that monitored natural attenuation (MNA) or bioremediation will be successful.

Is that a low, medium, or high concentration? The estimated percentile ranks retrieved from the MI Database answer that question by comparing your qPCR and QuantArray results to those of the literally thousands of other environmental samples submitted to MI for analysis over the last 20+ years.

Using the Estimated Percentile - Interpretation Examples

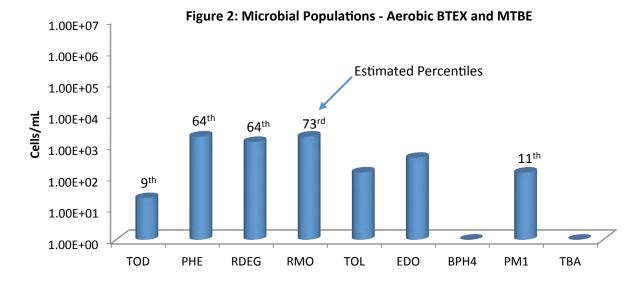
MNA Assessment – Petroleum Hydrocarbon Site:

Whenever possible, interpretation of qPCR and QuantArray results should include comparisons between samples obtained from background and impacted wells. The estimated percentile ranks however provide an additional avenue for comparison and evaluation of treatment options as shown below.



Anaerobic BTEX and PAH Biodegradation (Figure 1):

- With moderate concentrations of functional genes involved in anaerobic BTEX metabolism detected, the QuantArray-Petro® results were encouraging in terms of evaluating biodegradation potential under existing site conditions.
- More specifically, benzylsuccinate synthase (BSS) was detected on the order of nearly 10³ cells/mL indicating the presence of a substantial population (66th percentile) capable of anaerobic biodegradation of toluene and other alkyl substituted benzenes.
- Naphthyl-2-methylsuccinate synthase (MNSSA) and alkylsuccinate synthase (ASSA) genes were
 also detected indicating the potential for anaerobic biodegradation of 2-methylnaphthalene and
 normal alkanes.
- The concentration of MNSSA genes would be considered modest with an estimated percentile of 36th.
- While the percentile rank for MNSSA would be "below average", a number of additional factors should be considered.
 - First, anaerobic hydrocarbon degraders are less prevalent than aerobic BTEX degraders and overall detection frequencies for many genes involved in anaerobic hydrocarbon biodegradation are less than 50%.
 - Therefore, the detection of genes like BSS, MNSSA, ASSA, anaerobic benzene carboxylase (ABC), and anaerobic naphthalene carboxylase (ANC) even at low concentrations is certainly noteworthy and inherently "better than average".
 - The estimated percentiles for all assays are based only on samples where the concentration of the target gene was greater than the practical quantitation limit (PQL).
 - o For less commonly detected targets like many of the genes involved in anaerobic hydrocarbon biodegradation this is an especially important consideration.
 - Excluding samples where a gene target is below the PQL ensured that the median concentrations of less commonly detected targets would not be unduly biased low by the fact that the gene is not detected in most samples.
- Anaerobic benzene carboxylase (ABC) and naphthalene carboxylase (ANC) genes were also detected indicating the presence of bacterial populations capable of anaerobic biodegradation of benzene and naphthalene.
- For newly identified genes like ABC and ANC, estimated percentile ranks are not yet available due to the limited number of field samples that have been analyzed to date.
- However, like MNSSA and other genes involved in anaerobic hydrocarbon biodegradation, ABC and ANC detection frequencies are relatively low so the detection of these genes even at low concentrations should be considered when evaluating biodegradation potential under existing site conditions.

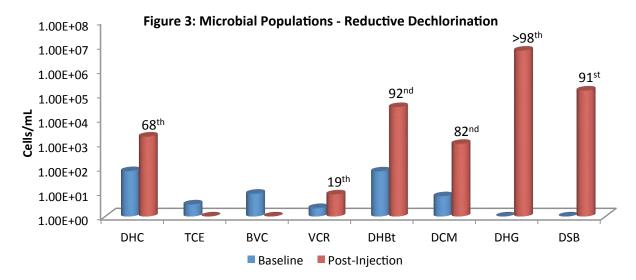


Aerobic BTEX and MTBE Biodegradation (Figure 2):

- With growing evidence that aromatic oxygenases function at low dissolved oxygen concentrations, aerobic BTEX biodegradation pathways should also be evaluated when considering MNA.
- Again, the QuantArray-Petro results were encouraging genes encoding the first step in multiple pathways for aerobic BTEX biodegradation were detected indicating the presence of a diverse population of aerobic BTEX degraders.
- However, aerobic BTEX degraders are often considered ubiquitous. Therefore answering the question "Is that low, medium or high?" becomes especially important when evaluating aerobic BTEX biodegradation at petroleum hydrocarbon sites.
- In this case, the estimated percentile ranks of the concentrations of toluene/benzene monooxygenase (RMO and RDEG) and phenol hydroxylase (PHE) genes ranged from the 64th to 73rd percentile.
- In other words, the concentrations of RMO, RDEG, and PHE detected in this groundwater sample were greater than the concentrations detected in 64% to 73% of all other groundwater samples where these genes were analyzed and detected above the PQL.
- Aerobic BTEX degraders are common in the environment, but in this sample concentrations of toluene/benzene monooxygenase genes could be viewed as "better than average" when compared to the MI Database.

Biostimulation – Chlorinated Solvent Site:

Whenever possible, interpretation of qPCR and QuantArray results should include comparisons between baseline and post-injection monitoring events as shown below (Figure 3). The estimated percentile ranks however provide an additional avenue for comparison and evaluation of remedy performance.



- During the baseline groundwater sampling event, Dehalococcoides and vinyl chloride reductase genes were detected indicating the potential for complete reductive dechlorination of PCE and TCE to ethene.
- However, the *Dehalococcoides* concentration was well below the 10⁴ cells/mL recommended by Lu et al. (2006) for generally effective rates of reductive dechlorination.
- Based on qPCR results as well as traditional groundwater monitoring, biostimulation with electron donor addition was selected as the site management plan.
- By the first monitoring event after injection, populations of halorespiring bacteria had increased substantially in response to electron donor addition.
 - o *Dehalobacter* populations increased by more than two orders of magnitude to post-injection concentrations greater than 10⁴ cells/mL (92nd percentile).
 - Dehalogenimonas (10⁶ cells/mL) and Desulfitobacterium (10⁵ cells/mL) which had not been detected prior electron donor addition were present at concentrations greater than observed in over 90% of other groundwater samples where these halorespiring bacteria were detected.
- After injection, Dehalococcoides populations increased by more than an order of magnitude to a concentration of over 10³ cells/mL (68th percentile) demonstrating growth of this key group of halorespiring bacteria.
- Despite a substantial increase and a "better than average" concentration, the *Dehalococcoides* population was still below the 10⁴ cells/mL threshold and vinyl chloride reductase gene copies were low (19th percentile).
 - o In terms of electron donors and acceptors, the metabolic capabilities of *Dehalococcoides* are rather specialized (hydrogen utilizing obligate halorespiring bacteria) so the median concentration is low. With a low median concentration across the database, a "better than average" *Dehalococcoides* concentration in a given sample may not exceed the 10⁴ cells/mL threshold established for effective reductive dechlorination (Lu et al. 2006) and ethene production (Microbial Insights, unpublished data).

- In this case, the initial growth of *Dehalococcoides* was substantial but may have been somewhat hindered by competition with sulfate reducing bacteria (Figure 4 below).
 - The baseline population of sulfate reducing bacteria was moderate (10⁴ cells/mL; 63rd percentile). Consistent with an observed decreased in dissolved sulfate concentrations, populations of sulfate reducing bacteria increased and were detected at a relatively high concentration (81st percentile) after electron donor addition.
 - After injection, methanogen populations also increased to a relatively high concentration (83rd percentile) suggesting generation of methanogenic conditions.
- With sulfate depletion and generation of highly anaerobic conditions more conducive to reductive dechlorination, *Dehalococcoides* populations may continue to increase and exceed the 10⁴ *Dehalococcoides* cells/mL threshold in subsequent monitoring events.
- Overall, QuantArray analysis conclusively demonstrated that electron donor addition stimulated growth of halorespiring bacteria with the estimated percentiles retrieved from the MI Database providing the "low, medium or high" perspective to the observed changes in microbial populations.

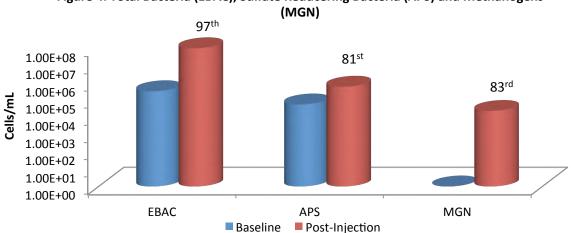


Figure 4: Total Bacteria (EBAC), Sulfate Reducering Bacteria (APS) and Methanogens

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Lu, X., J.T. Wilson, and D.H. Kampbell. 2006. Relationship between *Dehalococcoides* DNA in ground water and rates of reductive dechlorination at field scale. *Water Research* 40 no. 16: 3131-3140.



APPENDIX G

Validation Packages

(Available Upon

Request)



APPENDIX H

Laboratory Packages

(Available Upon

Request)